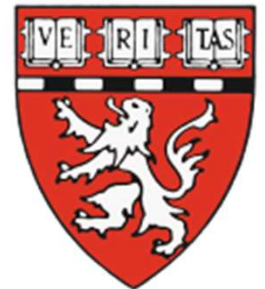


# **THE 2020 WORLD HEALTH ORGANIZATION CLASSIFICATION OF SOFT TISSUE TUMORS: AN UPDATE**

**Jason L Hornick, MD, PhD  
Director of Surgical Pathology  
and Immunohistochemistry  
Brigham and Women's Hospital  
Professor of Pathology  
Harvard Medical School  
Boston, MA, USA**





WHO Classification of Tumours 5th Edition: Soft Tissue and Bone Tumours  
2nd Editorial Board meeting: 6-8 May 2019, IARC, Lyon FRANCE



# Outline

- **New tumor types**
- **New concepts/revised nomenclature**
- **New genetics and novel diagnostic markers**

# WHO 2020: NEW TUMOR TYPES

## Adipocytic tumors

Atypical spindle cell/pleomorphic lipomatous tumor

Myxoid pleomorphic liposarcoma

## Fibroblastic/myofibroblastic tumors

*EWSR1-SMAD3*-positive fibroblastic tumor (emerging)

Angiofibroma of soft tissue

Superficial CD34-positive fibroblastic tumor

## Vascular tumors

Epithelioid hemangioendothelioma with *YAP1-TFE3*

## Smooth muscle tumors

Inflammatory leiomyosarcoma

## Tumors of uncertain differentiation

NTRK-rearranged spindle cell neoplasm (emerging)

## Undifferentiated small round cell sarcomas of bone and soft tissue

*CIC*-rearranged sarcoma

Sarcomas with *BCOR* genetic alterations

Round cell sarcomas with *EWSR1*-non-ETS fusions

# WHO 2020: NEW TUMOR TYPES

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## Undifferentiated small round cell sarcomas of bone and soft tissue

*CIC*-rearranged sarcoma

Sarcomas with *BCOR* genetic alterations

Round cell sarcomas with EWSR1-non-ETS fusions

# “Spindle Cell Liposarcoma”

The American Journal of Surgical Pathology 18(9): 913-921, 1994

## Spindle Cell Liposarcoma, A Hitherto Unrecognized Variant of Liposarcoma Analysis of Six Cases

Angelo P. Dei Tos, M.D., Thomas Mentzel, M.D.,  
Paul L. Newman, M.R.C.Path.,  
and Christopher D.M. Fletcher, M.D., M.R.C.Path.

MODERN PATHOLOGY (2010) 23, 729-736

## Well-differentiated spindle cell liposarcoma (‘atypical spindle cell lipomatous tumor’) does not belong to the spectrum of atypical lipomatous tumor but has a close relationship to spindle cell lipoma: clinicopathologic, immunohistochemical, and molecular analysis of six cases

Thomas Mentzel<sup>1</sup>, Gabriele Palmedo<sup>1</sup> and Cornelius Kuhnen<sup>2</sup>

*Am J Surg Pathol* • Volume 37, Number 9, September 2013

## Fibrosarcoma-like Lipomatous Neoplasm

### *A Reappraisal of So-called Spindle Cell Liposarcoma Defining a Unique Lipomatous Tumor Unrelated to Other Liposarcomas*

*Andrea T. Deyrup, MD, PhD,\* Frederic Chibon, PhD,† Louis Guillou, MD,‡  
Pauline Lagarde, MD,† Jean-Michel Coindre, MD,†§ and Sharon W. Weiss, MD||*

*Am J Surg Pathol* • Volume 41, Number 2, February 2017

# Atypical Spindle Cell Lipomatous Tumor

## *Clinicopathologic Characterization of 232 Cases Demonstrating a Morphologic Spectrum*

*Adrian Mariño-Enriquez, MD, PhD,\* Alessandra F. Nascimento, MD,\* Azra H. Ligon, PhD,\*  
Cherwei Liang, MD,† and Christopher D.M. Fletcher, MD, FRCPath\**

**M:F ratio: 3:2**

**Median age: 54 yr**

### Anatomic locations

Lower limb	36%
Upper limb	27%
Head/neck/face	10%
Retroperitoneum	<1%

### Depth

Subcutaneous	56%
Deep/subfascial	44%

**Local recurrence: 13%**

*Am J Surg Pathol* • Volume 41, Number 2, February 2017

# Atypical Spindle Cell Lipomatous Tumor

## *Clinicopathologic Characterization of 232 Cases Demonstrating a Morphologic Spectrum*

*Adrian Mariño-Enriquez, MD, PhD,\* Alessandra F. Nascimento, MD,\* Azra H. Ligon, PhD,\*  
Cherwei Liang, MD,† and Christopher D.M. Fletcher, MD, FRCPath\**

Marker	Positive
CD34	64%
S100	40%
Desmin	22%
RB1 (loss)	57%

Marker	Positive
MDM2	6%
CDK4	5%
Both	0%



*Am J Surg Pathol* • Volume 41, Number 11, November 2017

## “Atypical” Pleomorphic Lipomatous Tumor

*A Clinicopathologic, Immunohistochemical and Molecular Study of 21 Cases, Emphasizing its Relationship to Atypical Spindle Cell Lipomatous Tumor and Suggesting a Morphologic Spectrum (Atypical Spindle Cell/Pleomorphic Lipomatous Tumor)*

*David Creytens, MD, PhD,\*† Thomas Mentzel, MD, PhD,‡ Liesbeth Ferdinande, MD, PhD,\*† Evelyne Lecoutere, MD,\* Joost van Gorp, MD, PhD,§ Lilit Atanesyan, PhD,|| Karel de Groot, MSc,|| Suvi Savola, PhD,|| Nadine Van Roy, PhD,†¶ Jo Van Dorpe, MD, PhD,\*† and Uta Flucke, MD, PhD#*

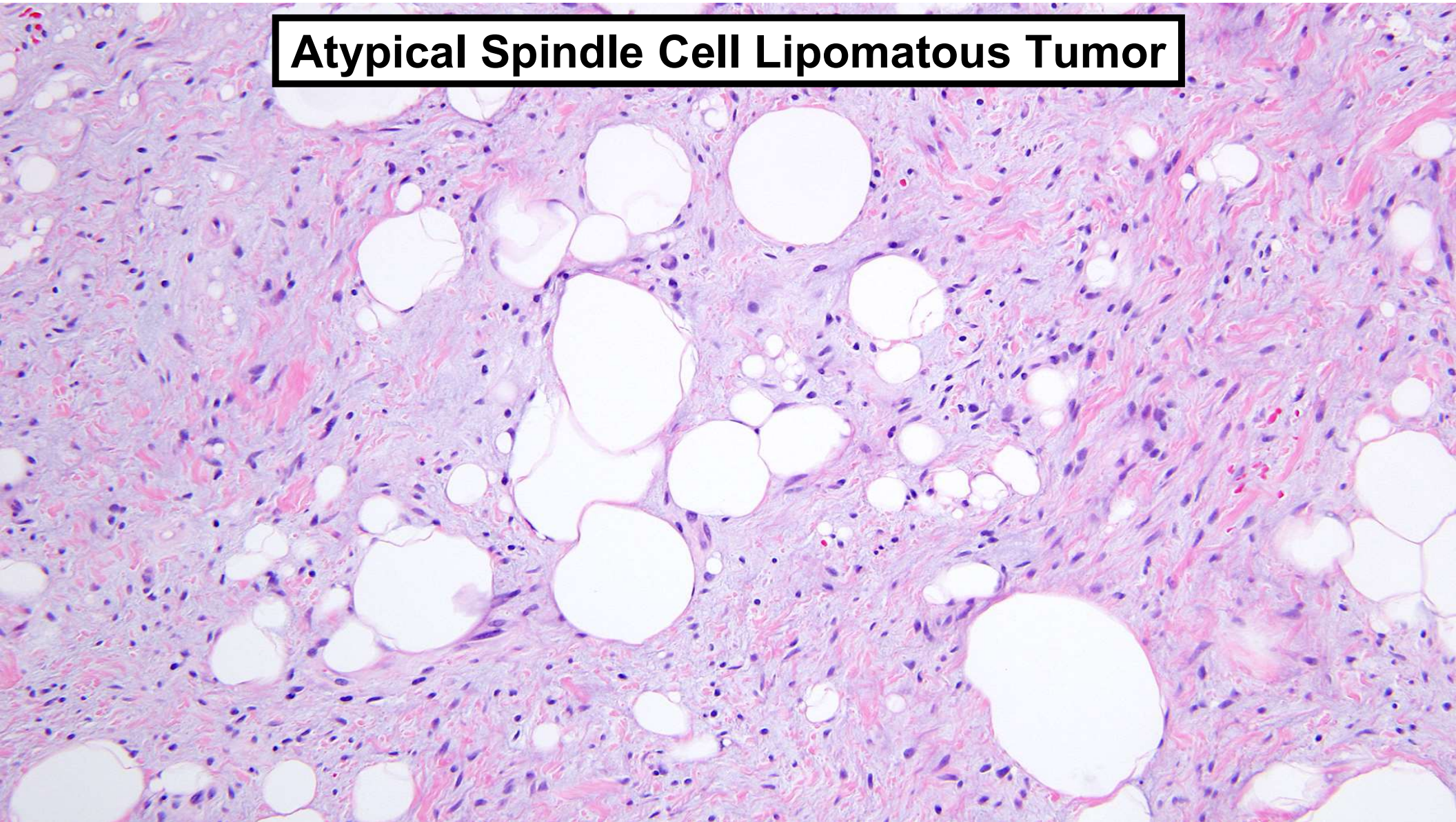
*Am J Surg Pathol* • Volume 45, Number 9, September 2021

## Atypical Pleomorphic Lipomatous Tumor

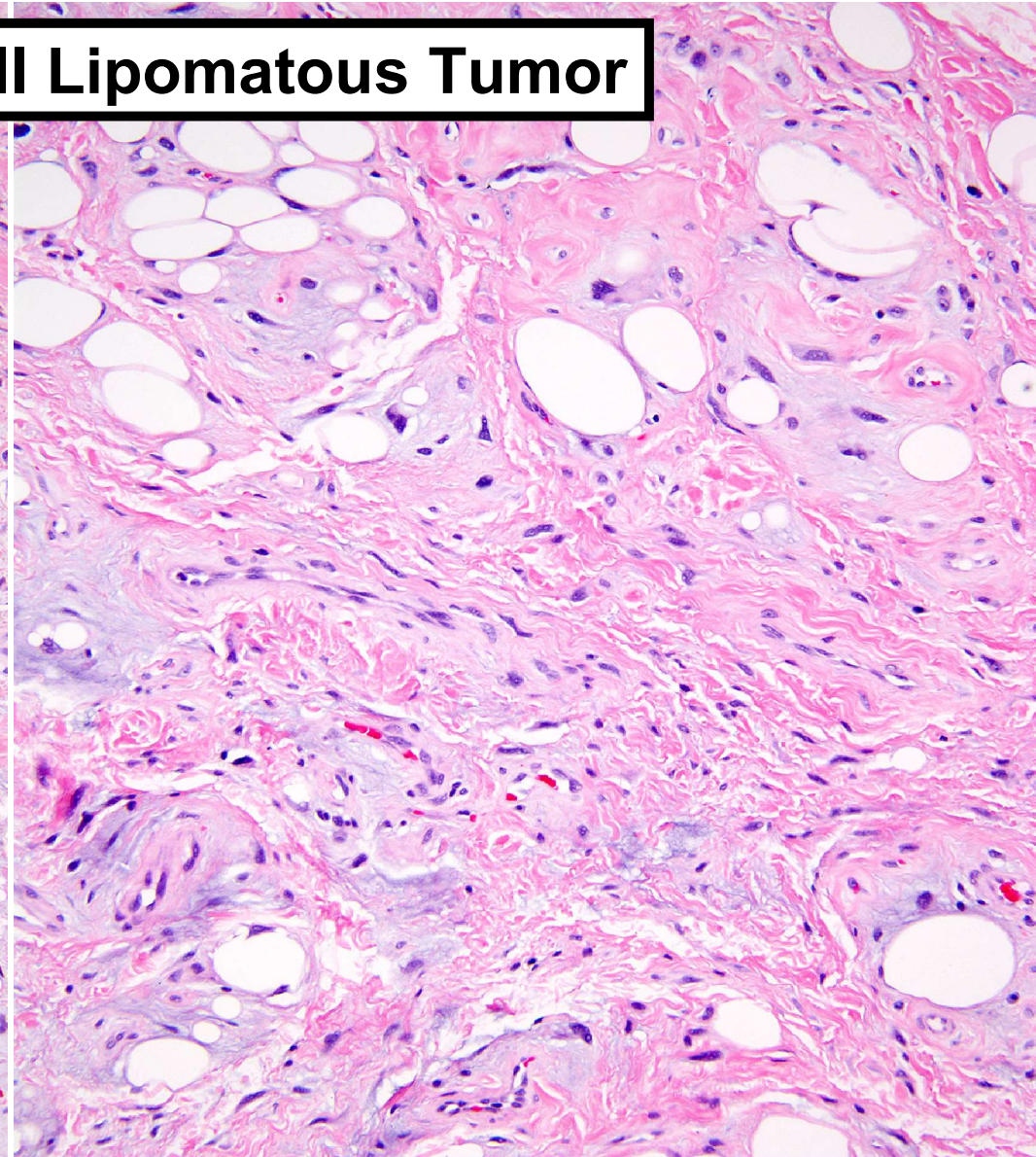
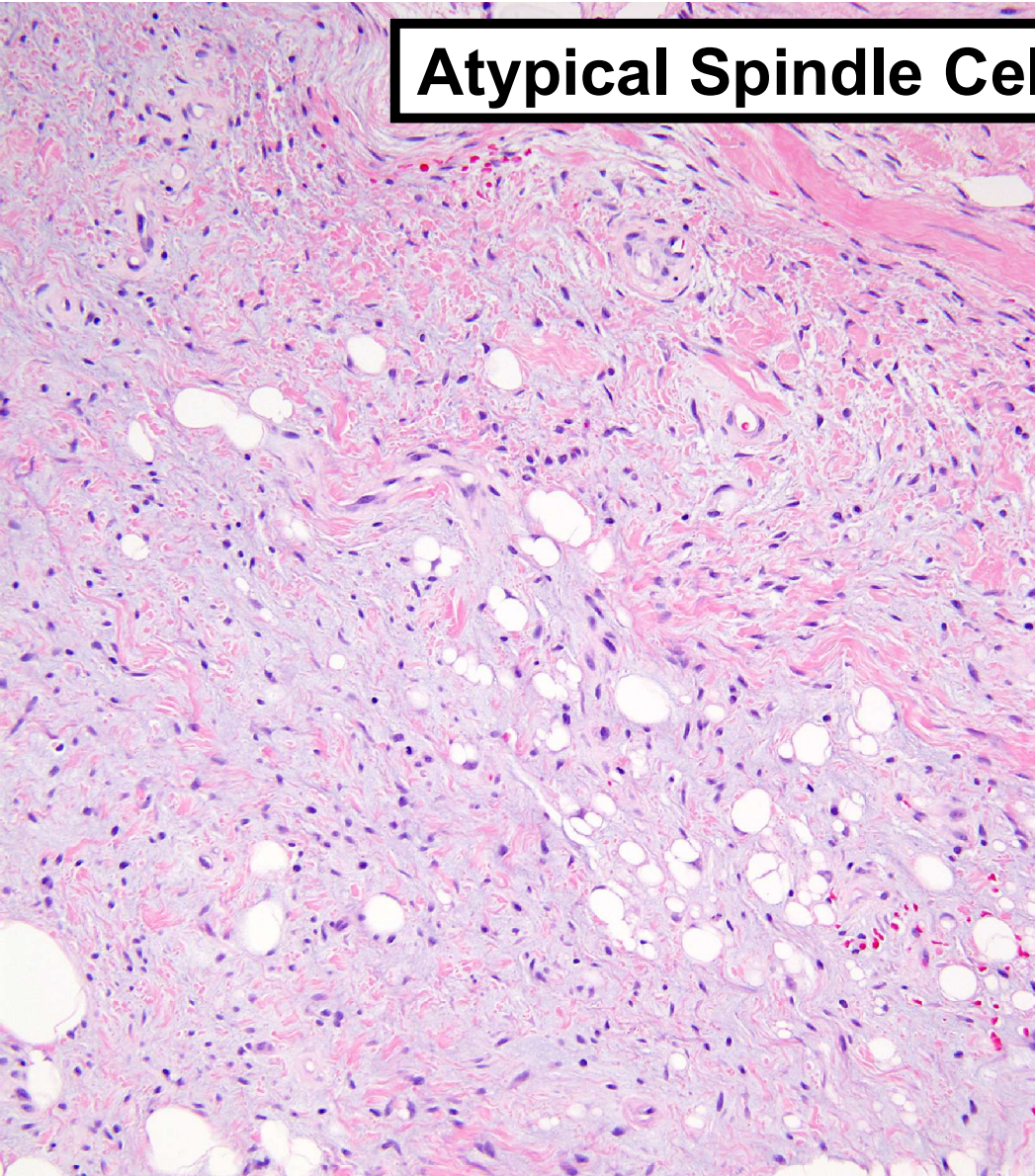
*Expanding Our Current Understanding in a Clinicopathologic Analysis of 64 Cases*

*William J. Anderson, MBChB, Christopher D.M. Fletcher, MD, FRCPath, and Vickie Y. Jo, MD*

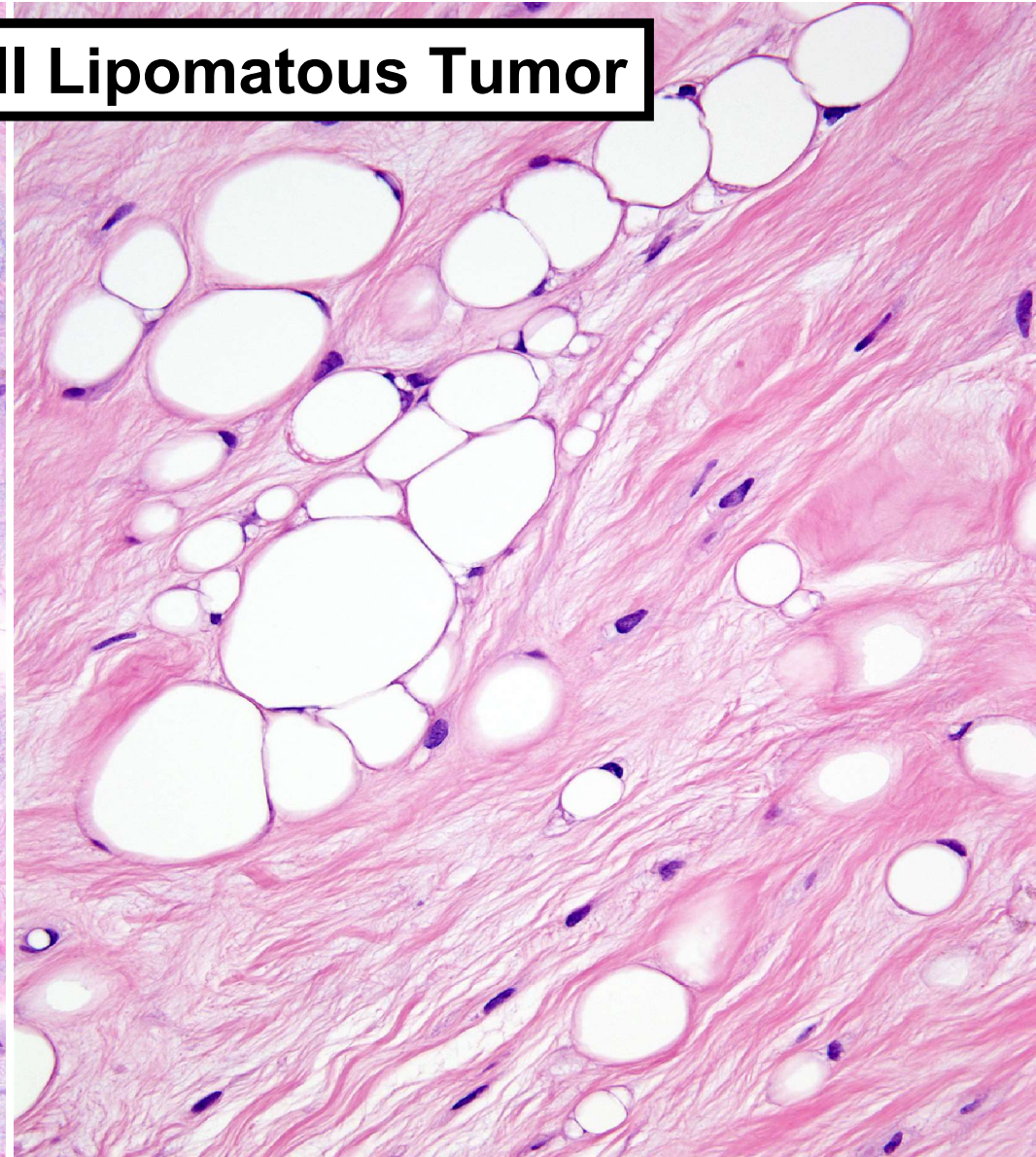
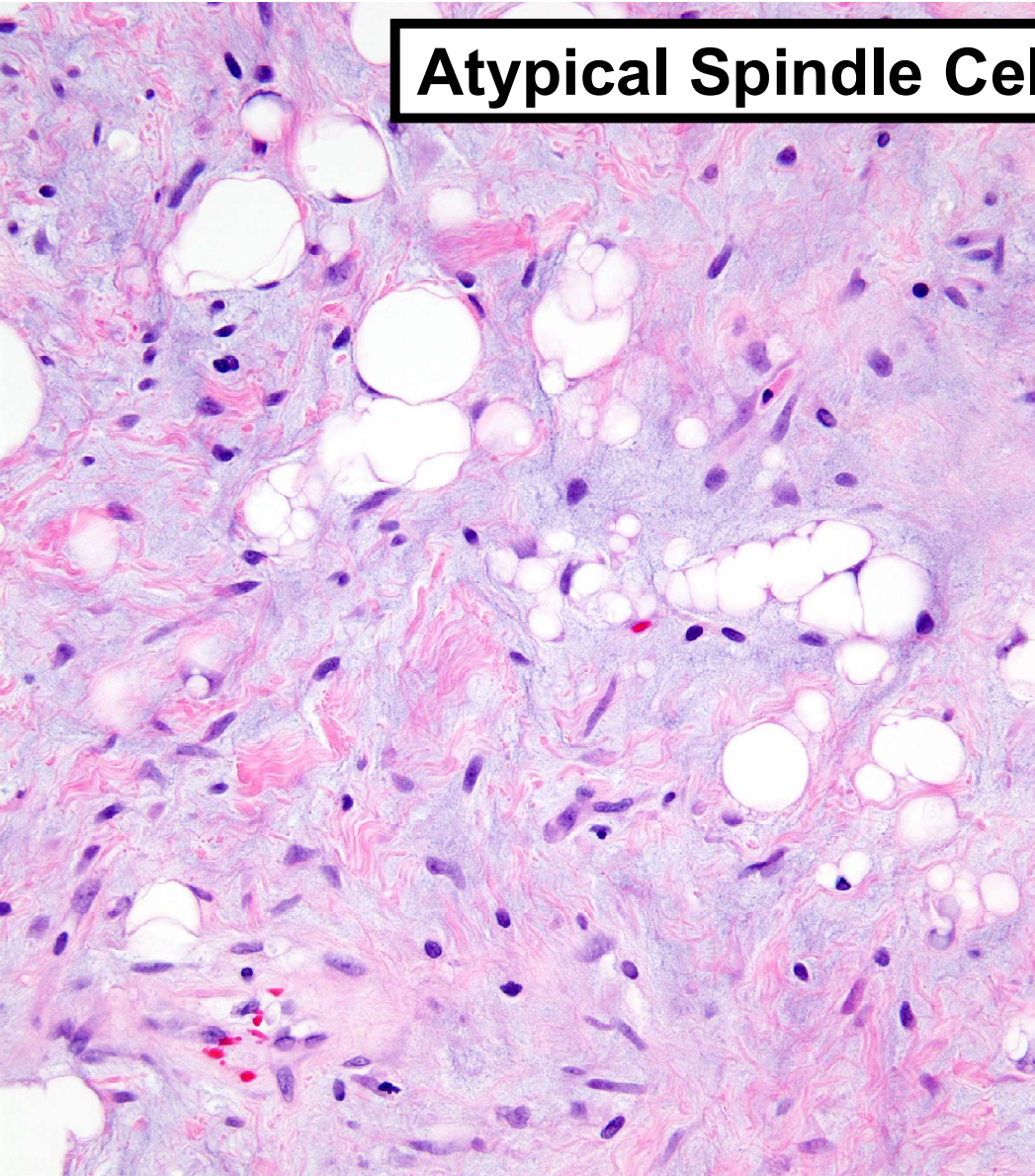
**Atypical Spindle Cell Lipomatous Tumor**



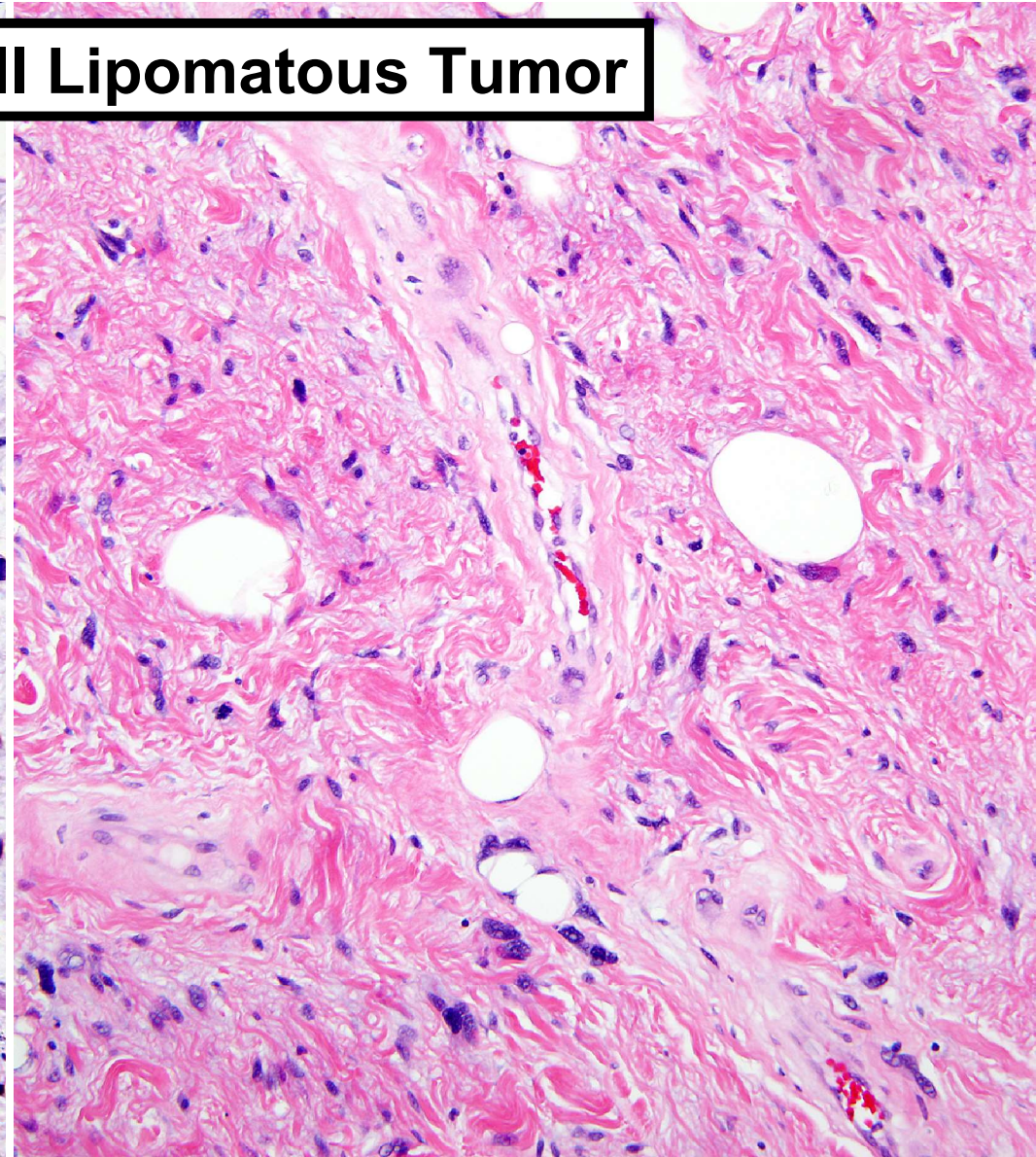
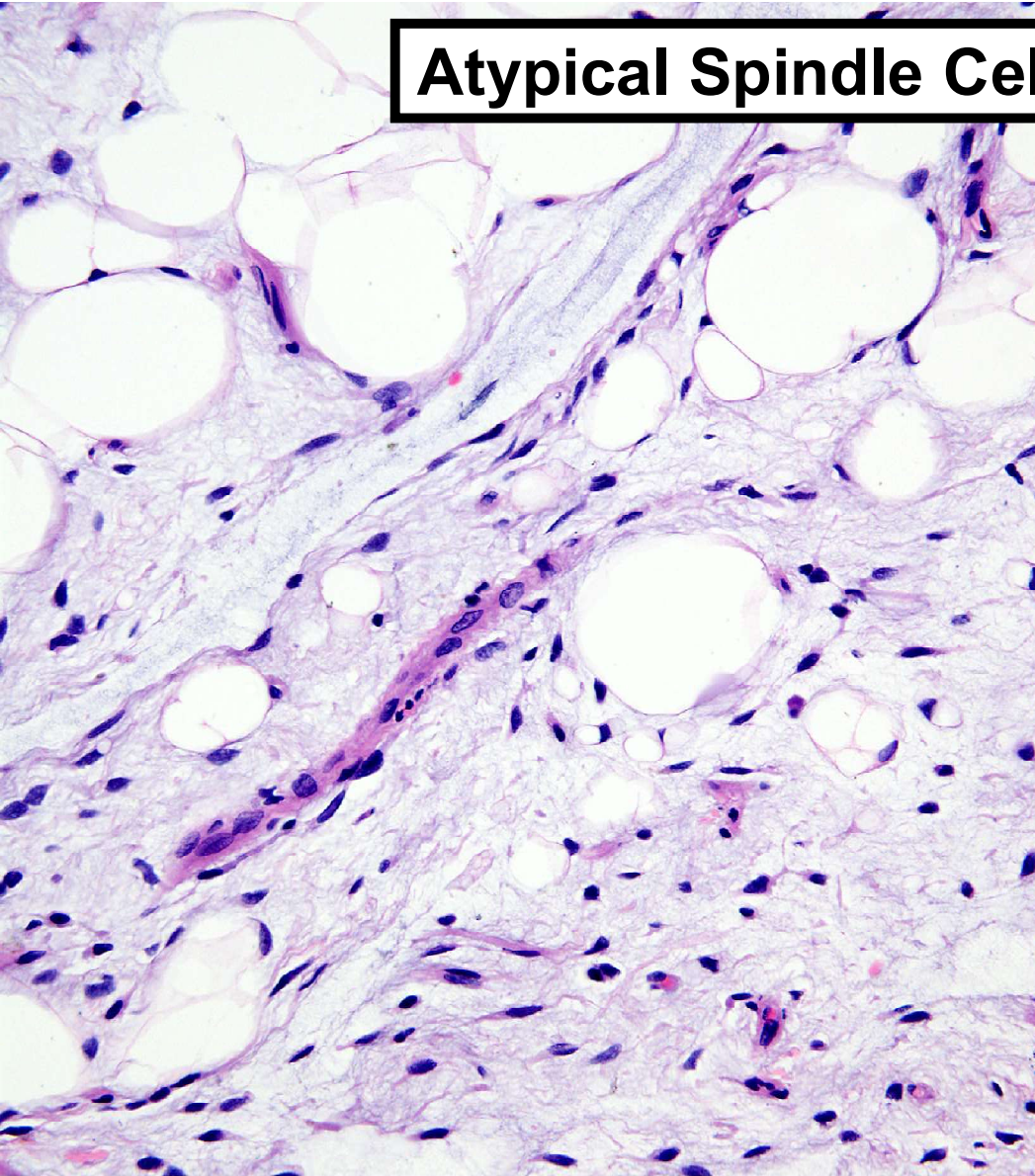
# Atypical Spindle Cell Lipomatous Tumor



# Atypical Spindle Cell Lipomatous Tumor



# Atypical Spindle Cell Lipomatous Tumor



## Liposarcomas in Young Patients

*A Study of 82 Cases Occurring in Patients Younger Than 22 Years of Age*

*Rita Alaggio, MD,\* Cheryl M. Coffin, MD,† Sharon W. Weiss, MD,‡ Julia A. Bridge, MD,§  
Josephine Issakov, MD,|| Andre M. Oliveira, MD,¶ and Andrew L. Folpe, MD¶¶*

*Am J Surg Pathol • Volume 33, Number 5, May 2009*

## Liposarcomas of the Mediastinum and Thorax

*A Clinicopathologic and Molecular Cytogenetic  
Study of 24 Cases, Emphasizing Unusual  
and Diverse Histologic Features*

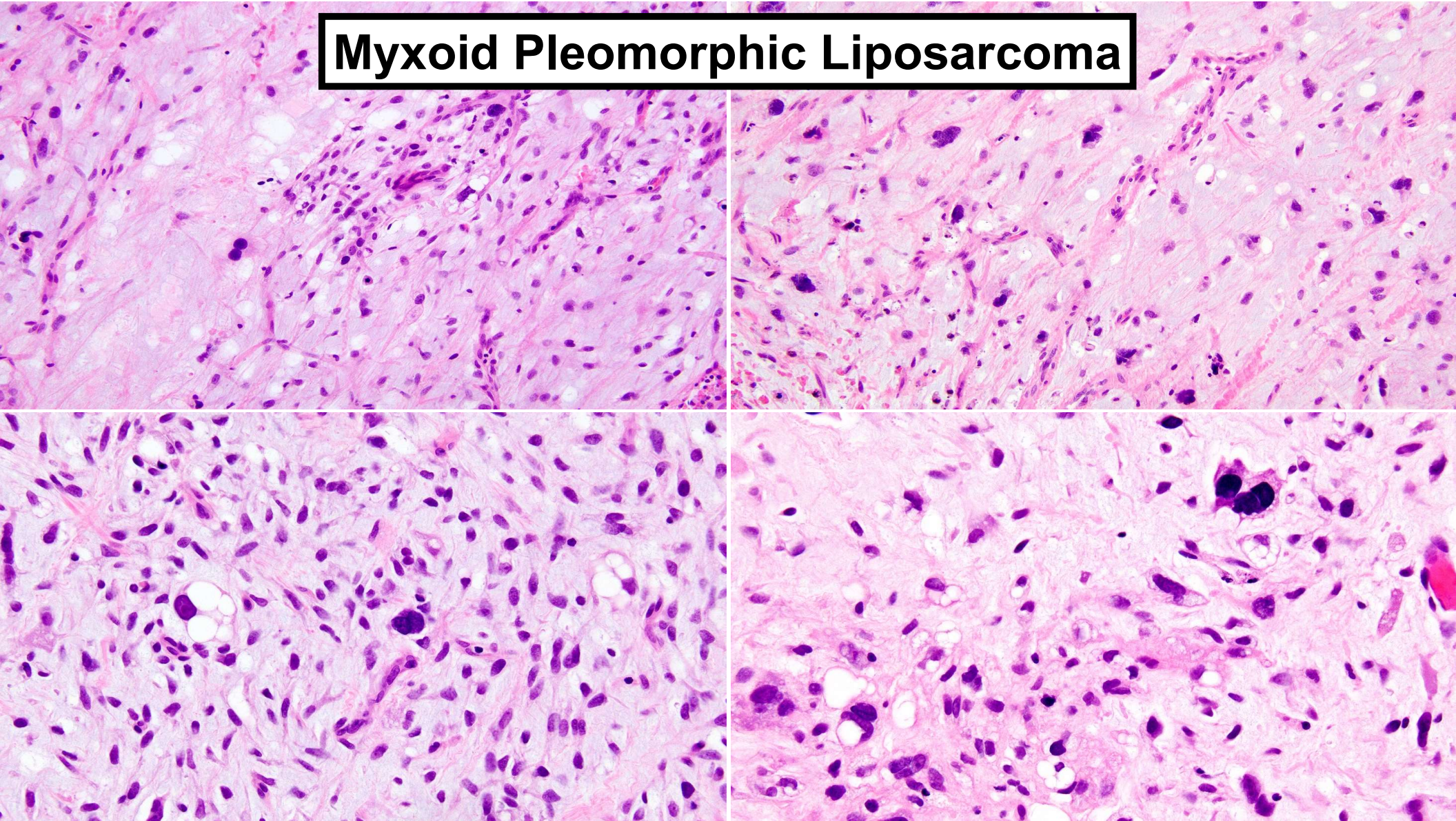
*Jennifer M. Boland, MD,\* Thomas V. Colby, MD,† and Andrew L. Folpe, MD\**

*Am J Surg Pathol • Volume 36, Number 9, September 2012*

# Myxoid pleomorphic liposarcoma

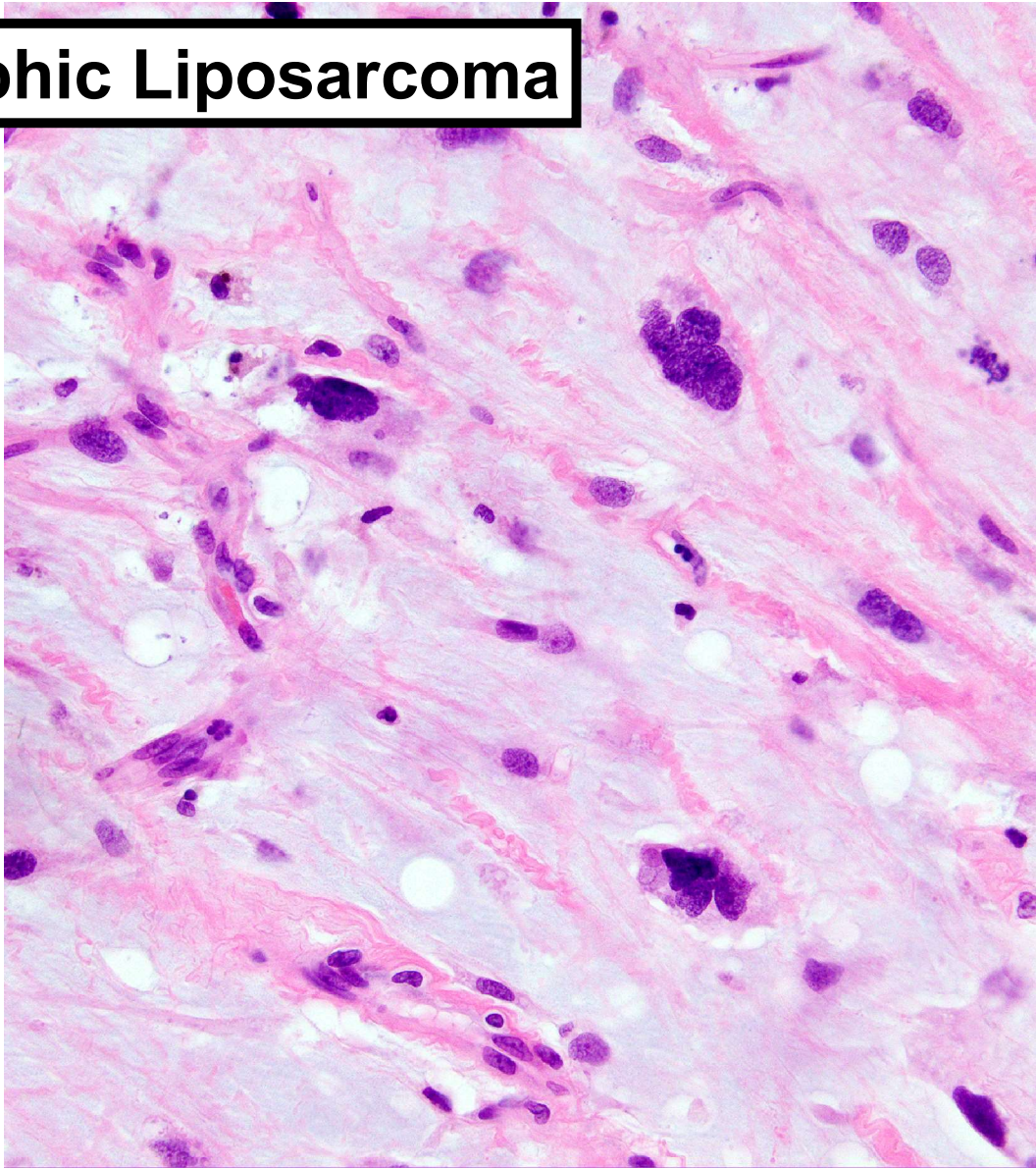
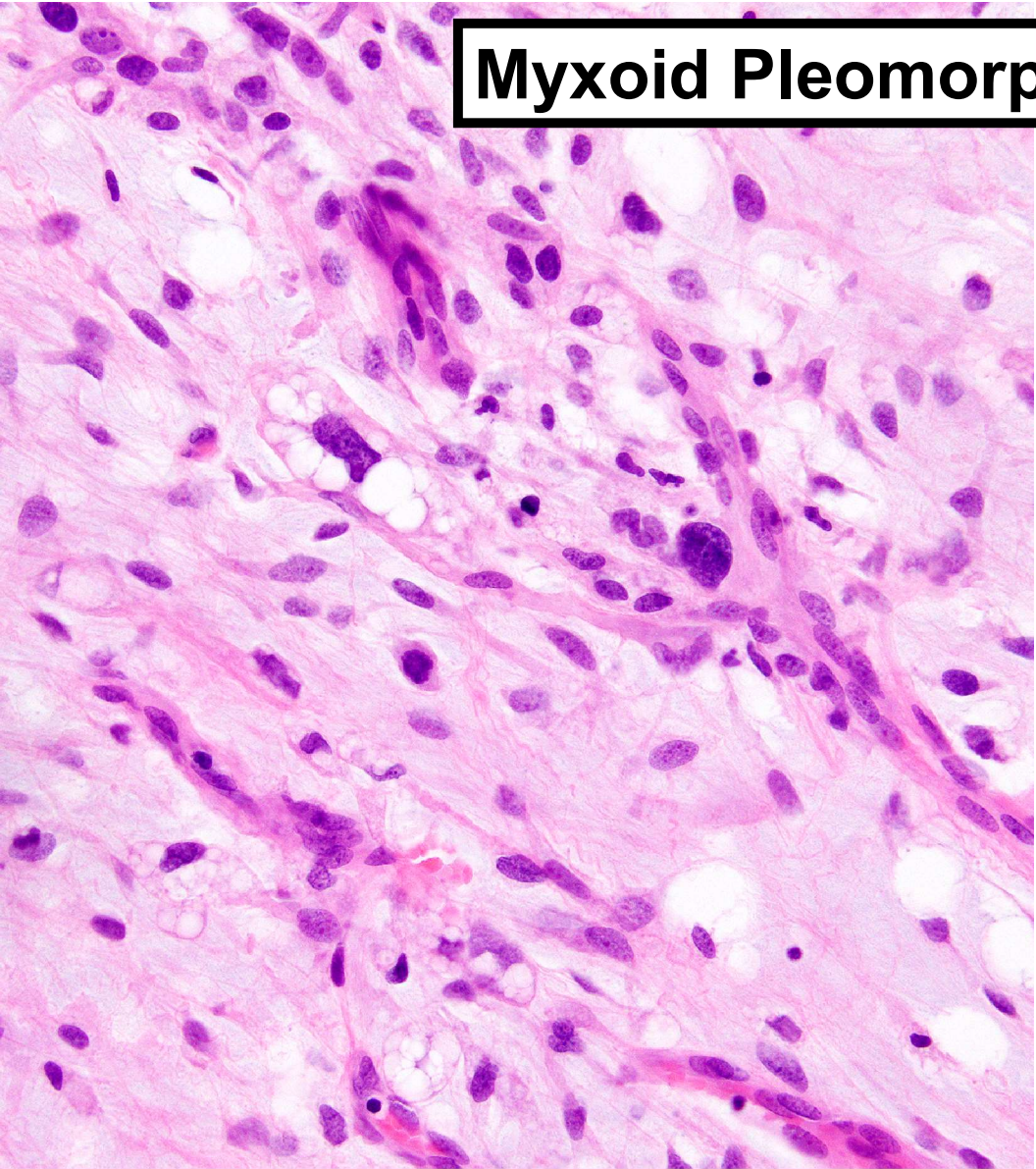
- **Predilection for the mediastinum of children and young adults**
- **Histology: admixture of hypocellular zones with myxoid stroma, bland nuclei, and delicate branching vessels and areas with atypia and pleomorphism**
- **Lack *DDIT3* rearrangements and *MDM2* amplification**
- **Some cases associated with Li-Fraumeni syndrome (germline *TP53* mutation)**
- **Clinically aggressive with high rate of local recurrence and distant metastasis**

# Myxoid Pleomorphic Liposarcoma





**Myxoid Pleomorphic Liposarcoma**



*Am J Surg Pathol* • Volume 36, Number 4, April 2012

# Angiofibroma of Soft Tissue: Clinicopathologic Characterization of a Distinctive Benign Fibrovascular Neoplasm in a Series of 37 Cases

*Adrián Mariño-Enríquez, MD and Christopher D. M. Fletcher, MD, FRCPath*

**M:F ratio: 2:1**

**Median age: 49 yr**

## Anatomic locations

Lower limb 62%

Upper limb 27%

## Depth

Subcutaneous 56%

Deep/subfascial 44%

## Marker

## Positive

EMA 44%

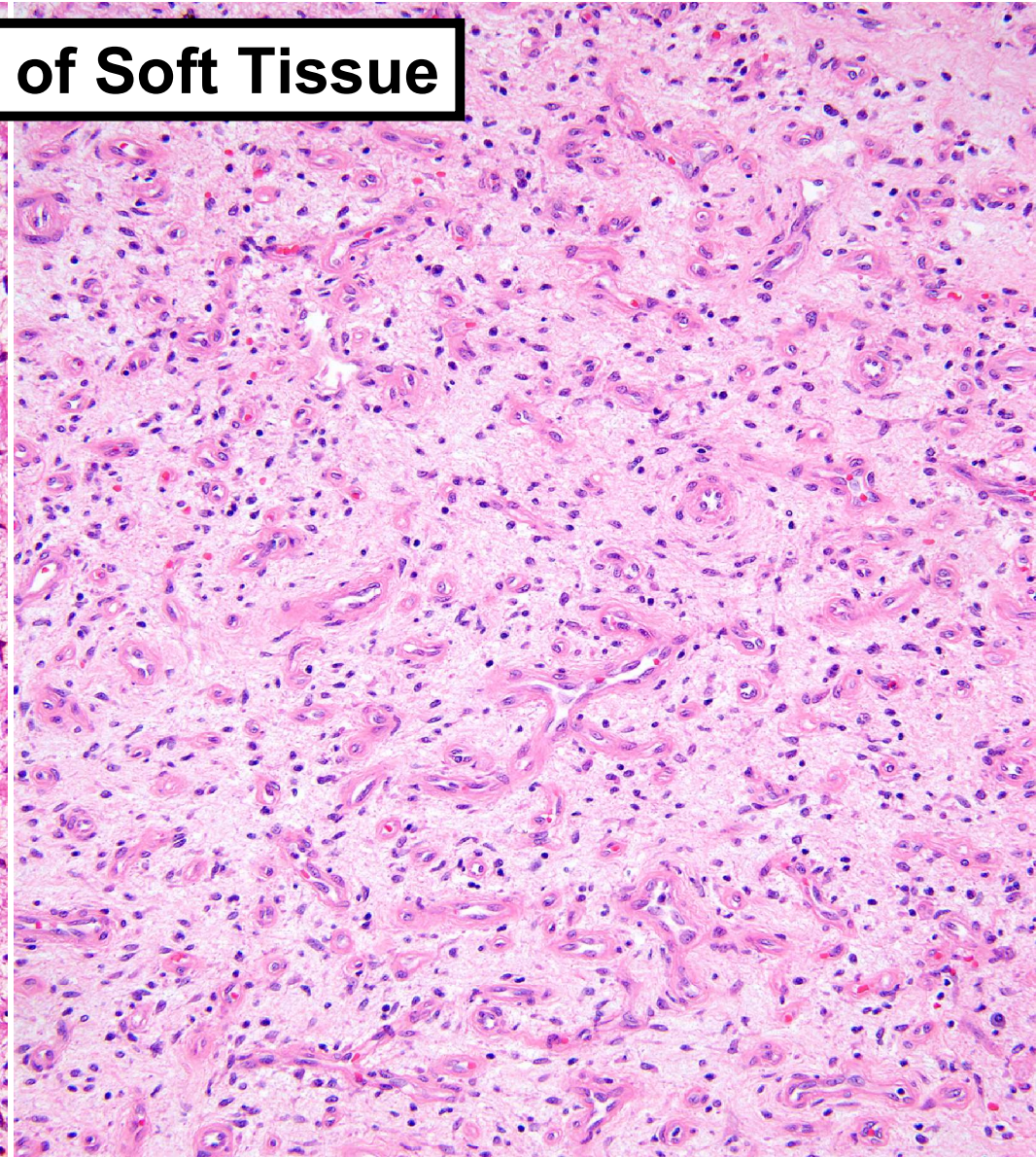
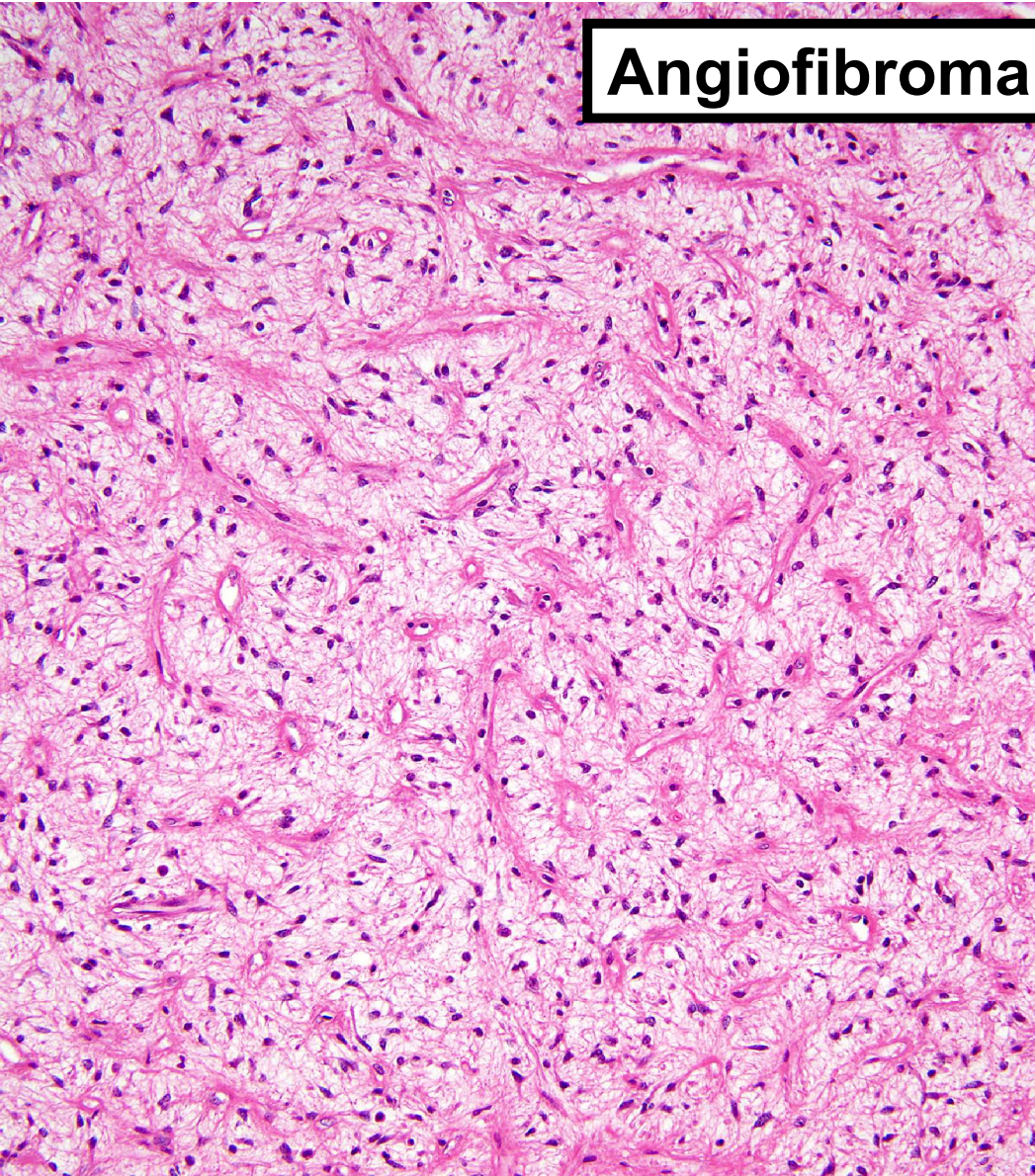
CD34 15%

SMA 15%

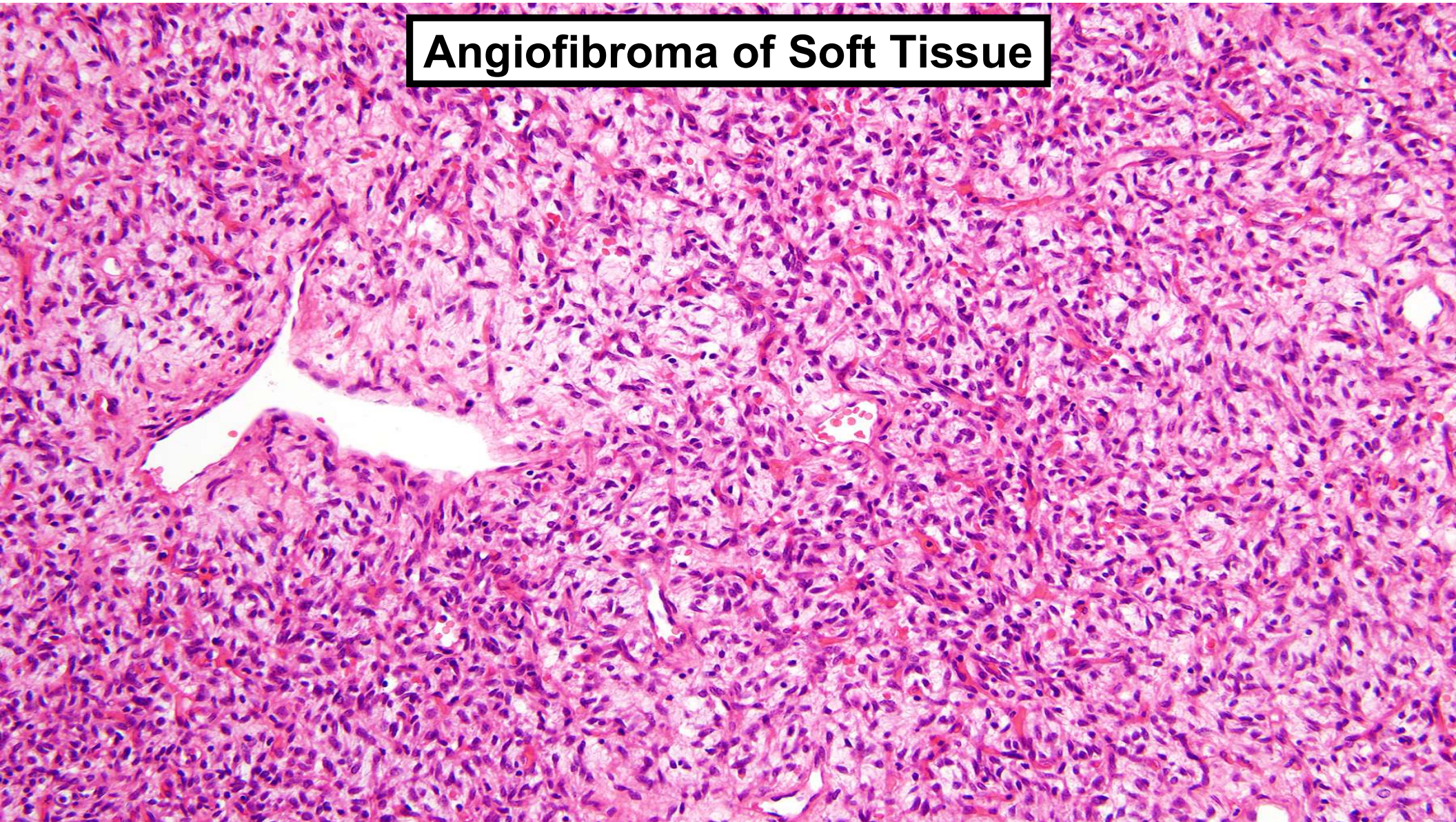
Desmin 10%

**Low rate local recurrence**

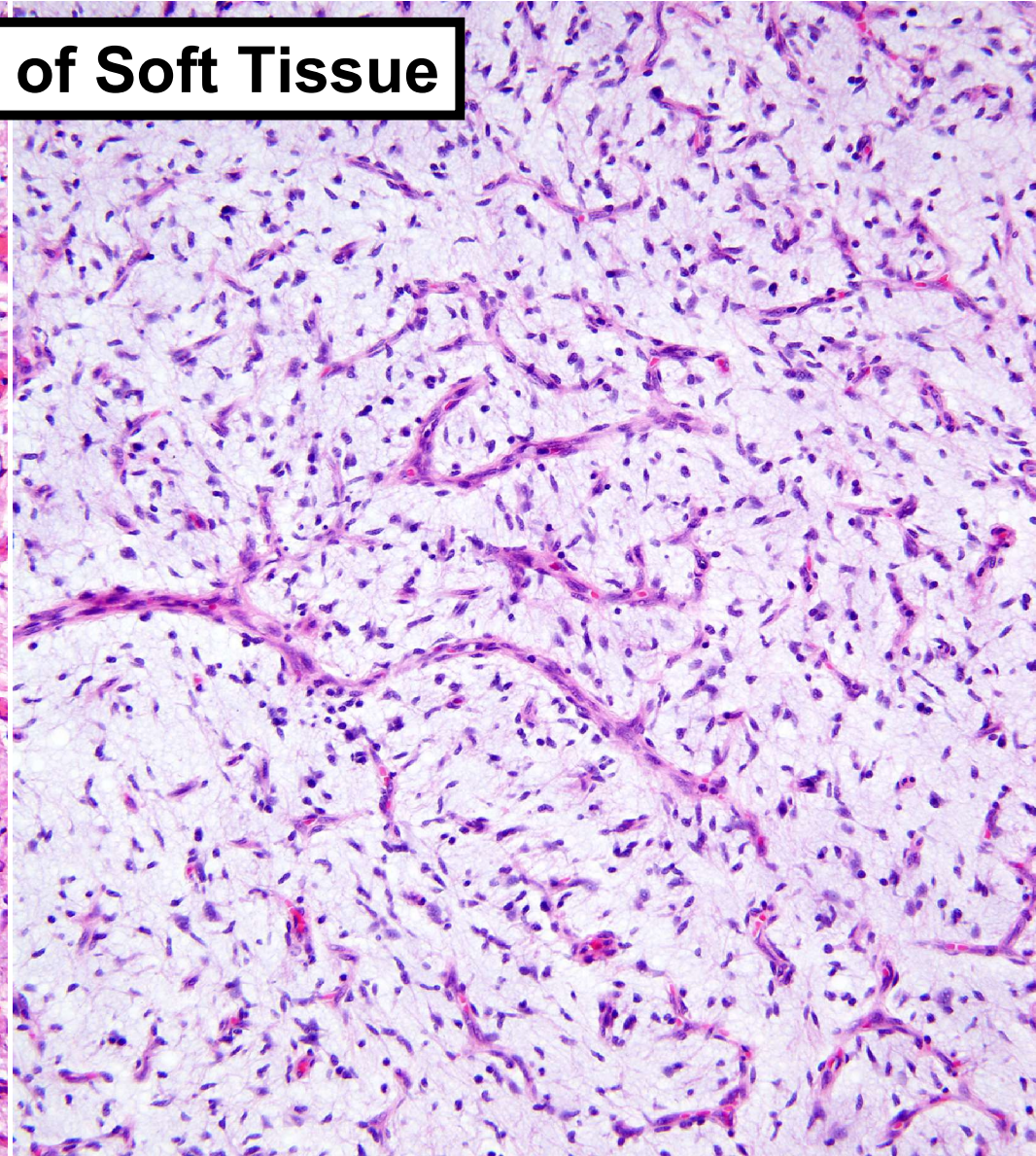
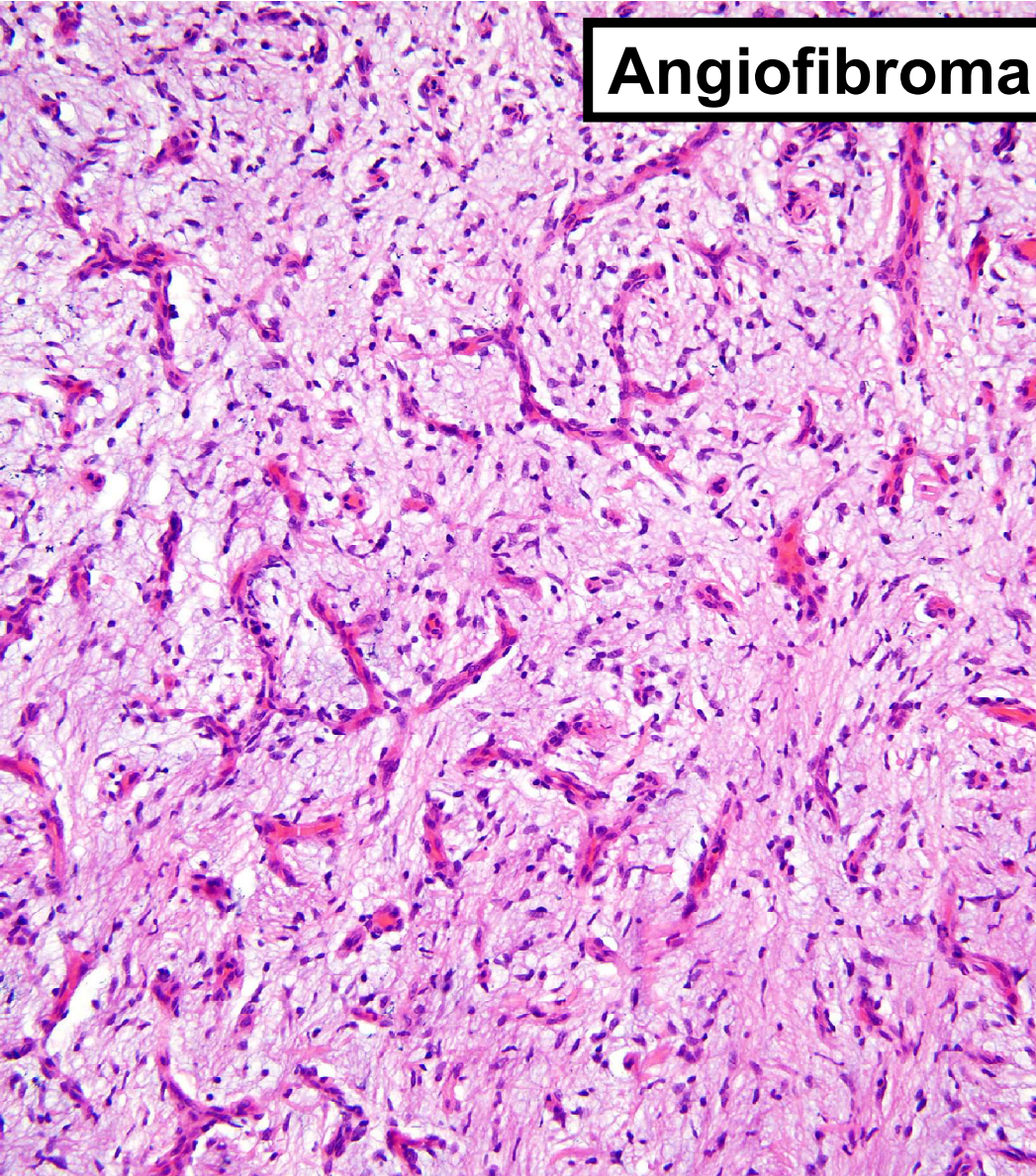
# Angiofibroma of Soft Tissue



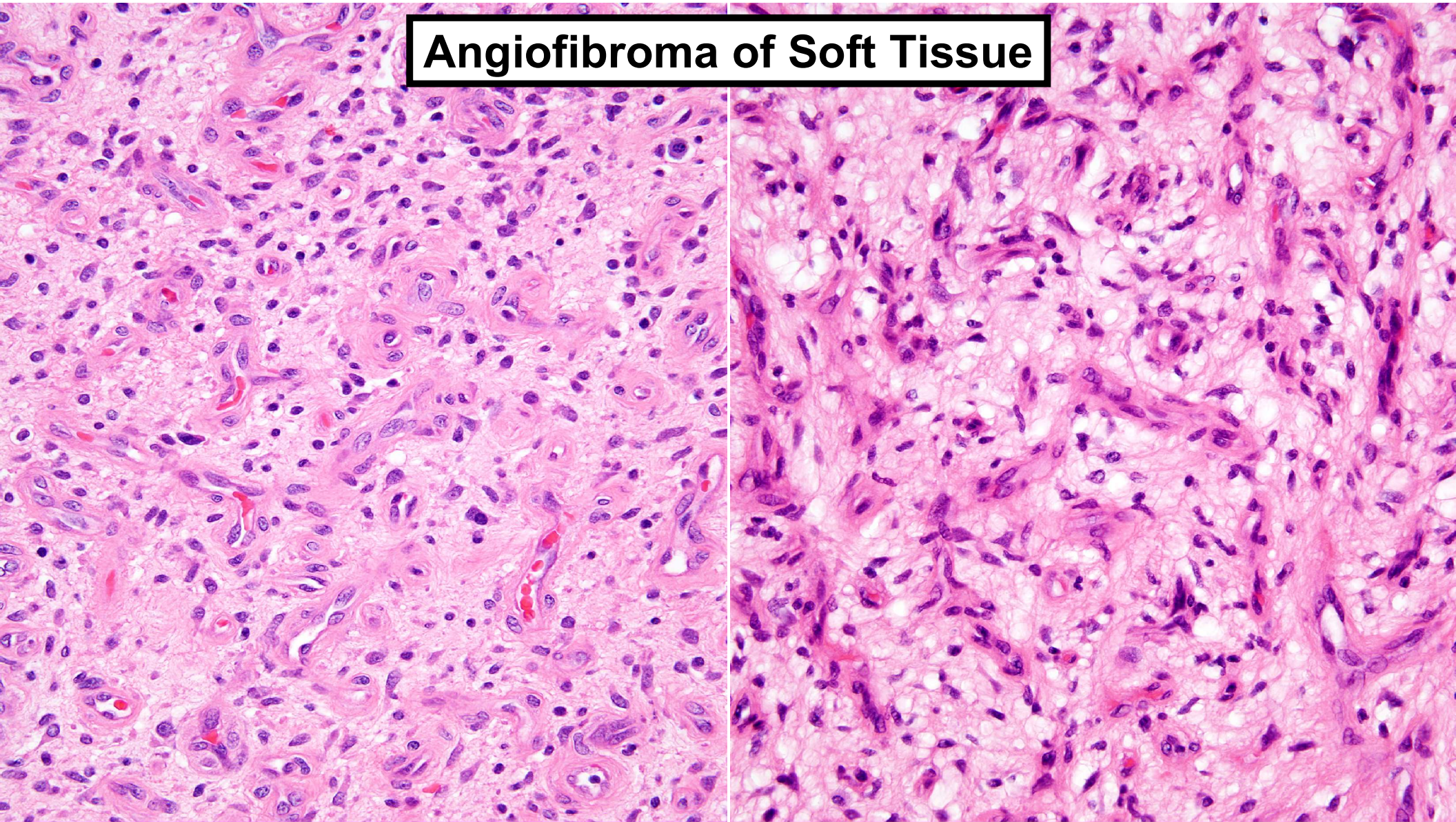
## Angiofibroma of Soft Tissue



## Angiofibroma of Soft Tissue

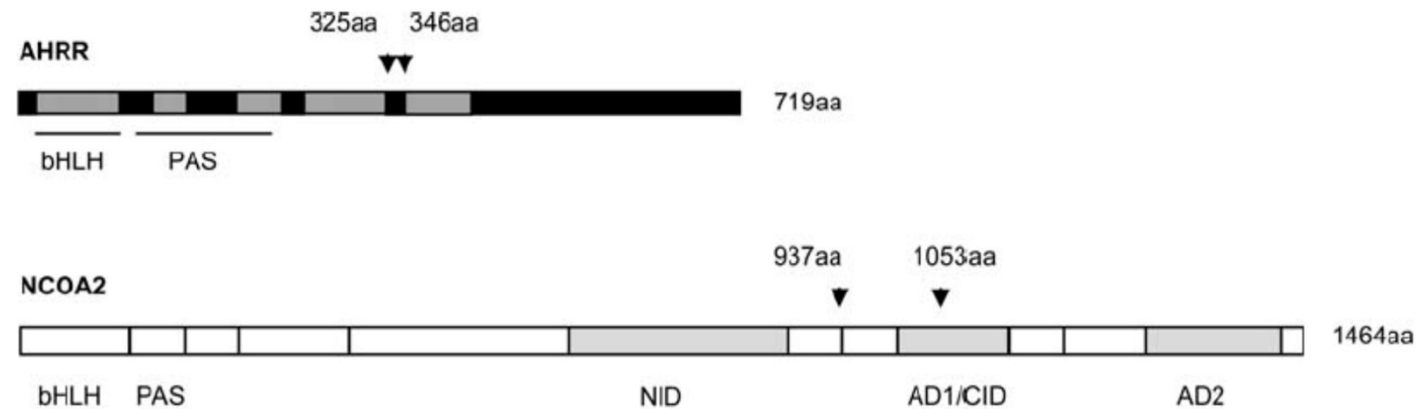
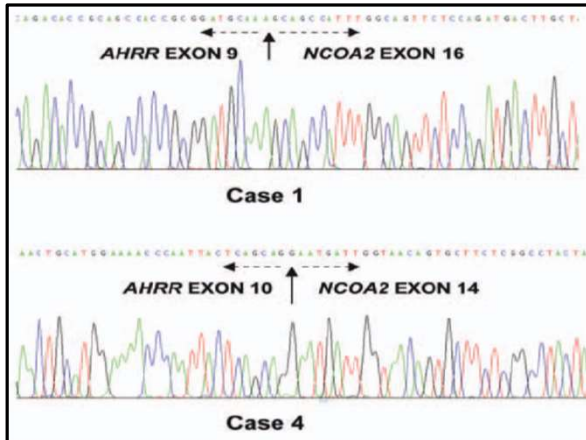


## Angiofibroma of Soft Tissue



# Fusion of the *AHRR* and *NCOA2* Genes Through a Recurrent Translocation $t(5;8)(p15;q13)$ in Soft Tissue Angiofibroma Results in Upregulation of Aryl Hydrocarbon Receptor Target Genes

Yuesheng Jin,<sup>1</sup> Emely Möller,<sup>1</sup> Karolin H. Nord,<sup>1</sup> Nils Mandahl,<sup>1</sup> Fredrik Vult Von Steyern,<sup>2</sup> Henryk A. Domanski,<sup>3</sup> Adrian Mariño-Enríquez,<sup>4</sup> Linda Magnusson,<sup>1</sup> Jenny Nilsson,<sup>1</sup> Raf Scot,<sup>5</sup> Christopher D. M. Fletcher,<sup>4</sup> Maria Debiec-Rychter,<sup>6</sup> and Fredrik Mertens<sup>1\*</sup>

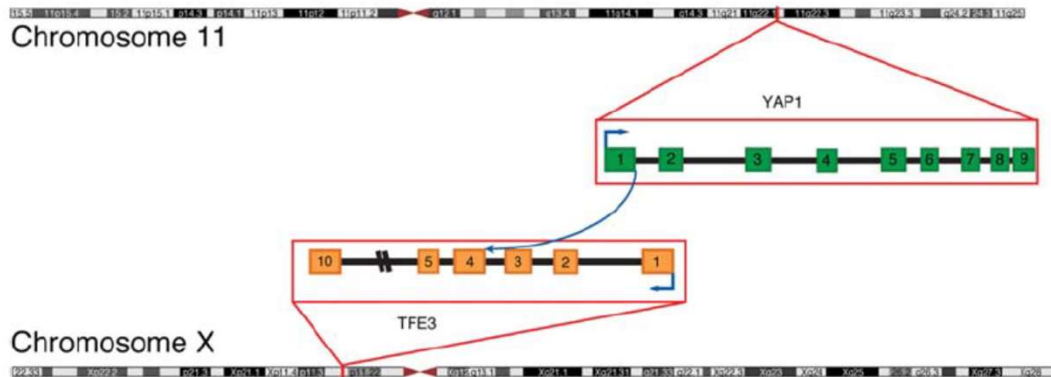


# Novel *YAPI-TFE3* Fusion Defines a Distinct Subset of Epithelioid Hemangioendothelioma

Cristina R. Antonescu,<sup>1\*</sup> Francois Le Loarer,<sup>1</sup> Juan-Miguel Mosquera,<sup>2</sup> Andrea Sboner,<sup>2,3</sup> Lei Zhang,<sup>1</sup> Chun-Liang Chen,<sup>1</sup> Hsiao-Wei Chen,<sup>1</sup> Nursat Pathan,<sup>4</sup> Thomas Krausz,<sup>5</sup> Brendan C. Dickson,<sup>6</sup> Ilan Weinreb,<sup>7</sup> Mark A. Rubin,<sup>2</sup> Meera Hameed,<sup>1</sup> and Christopher D. M. Fletcher<sup>8\*</sup>

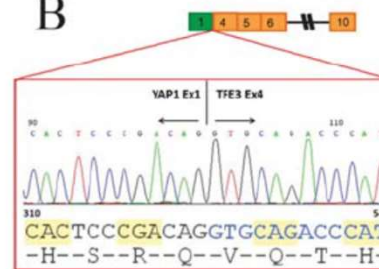
A

Chromosome 11

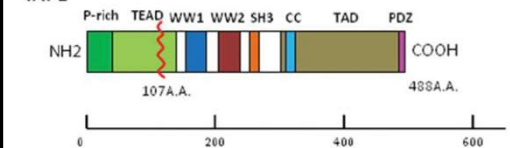


Chromosome X

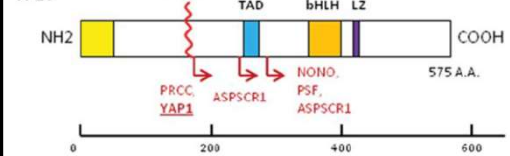
B



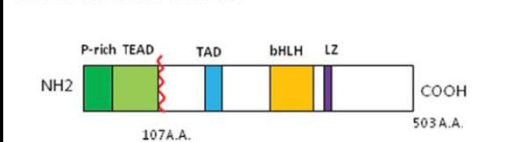
YAP1



TFE3

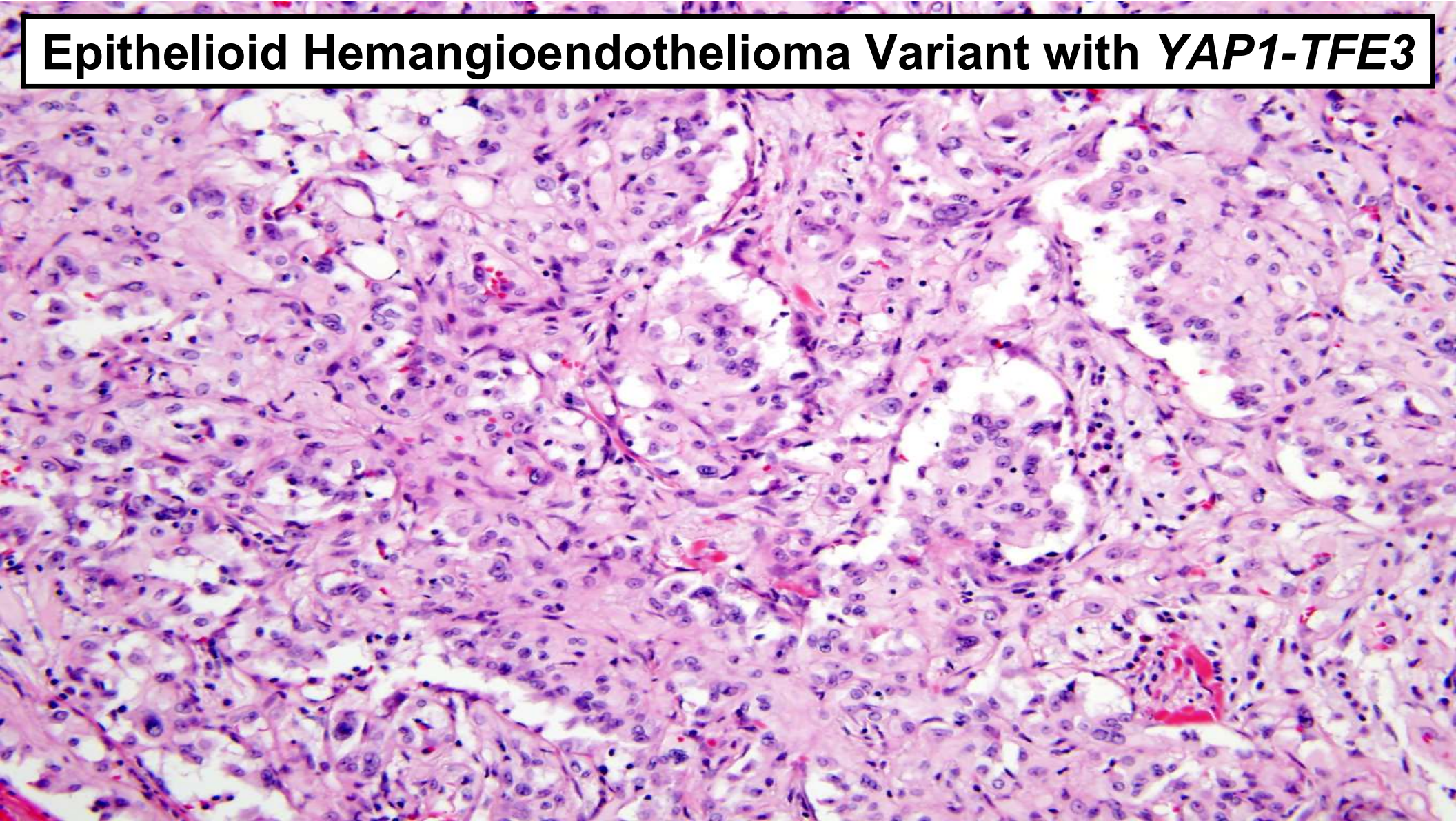


Fusion YAP1-Ex1-TFE3-Ex4

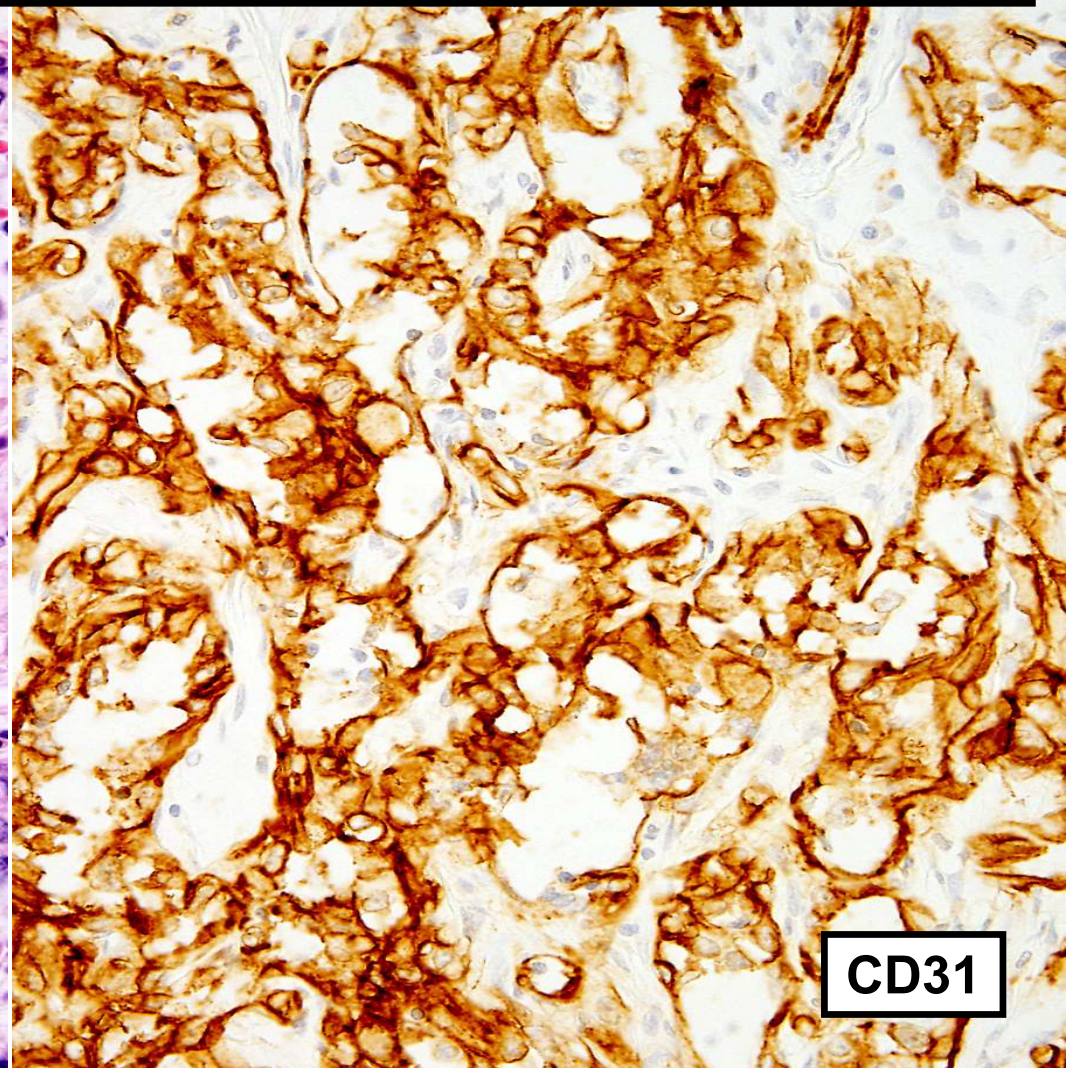
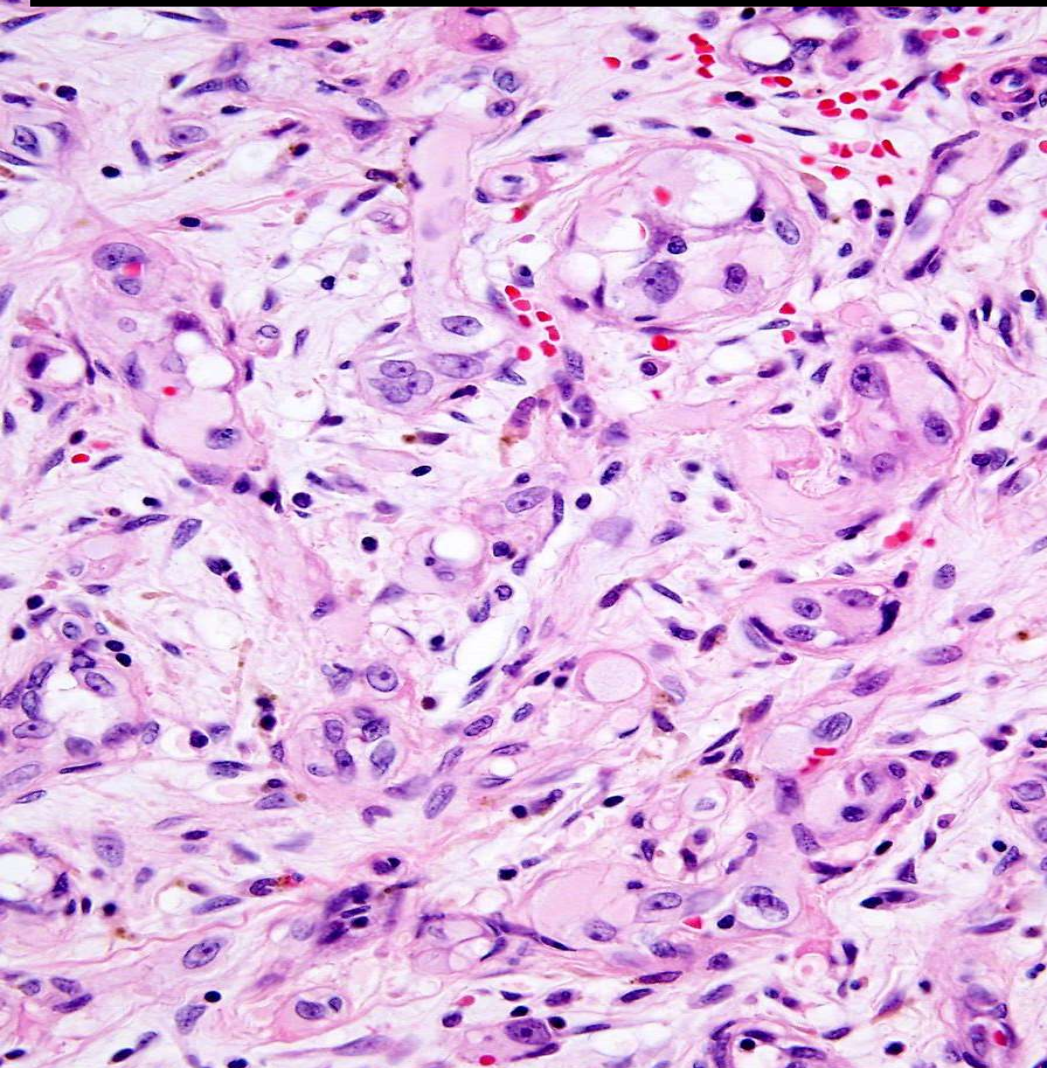




**Epithelioid Hemangioendothelioma Variant with *YAP1-TFE3***

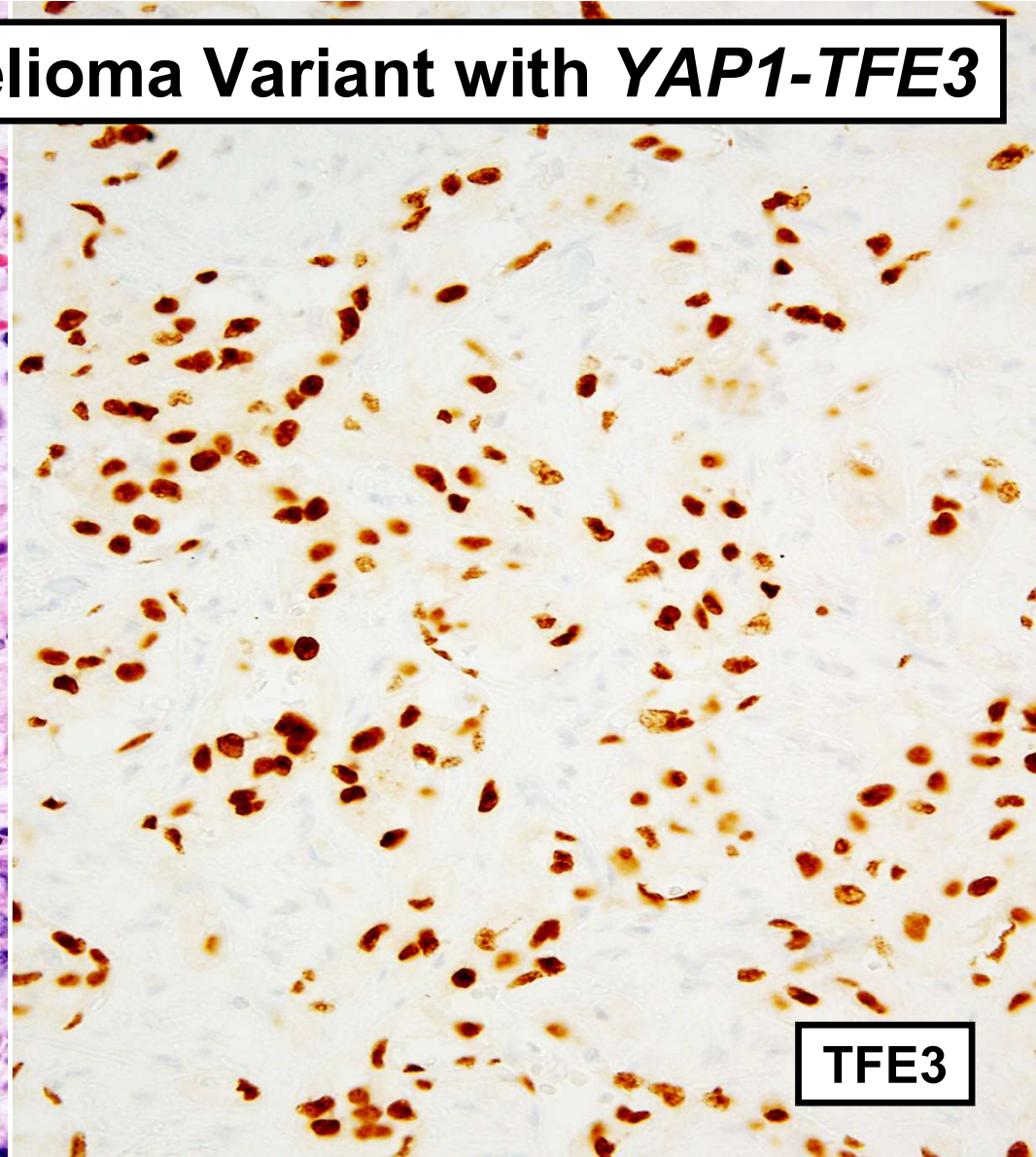
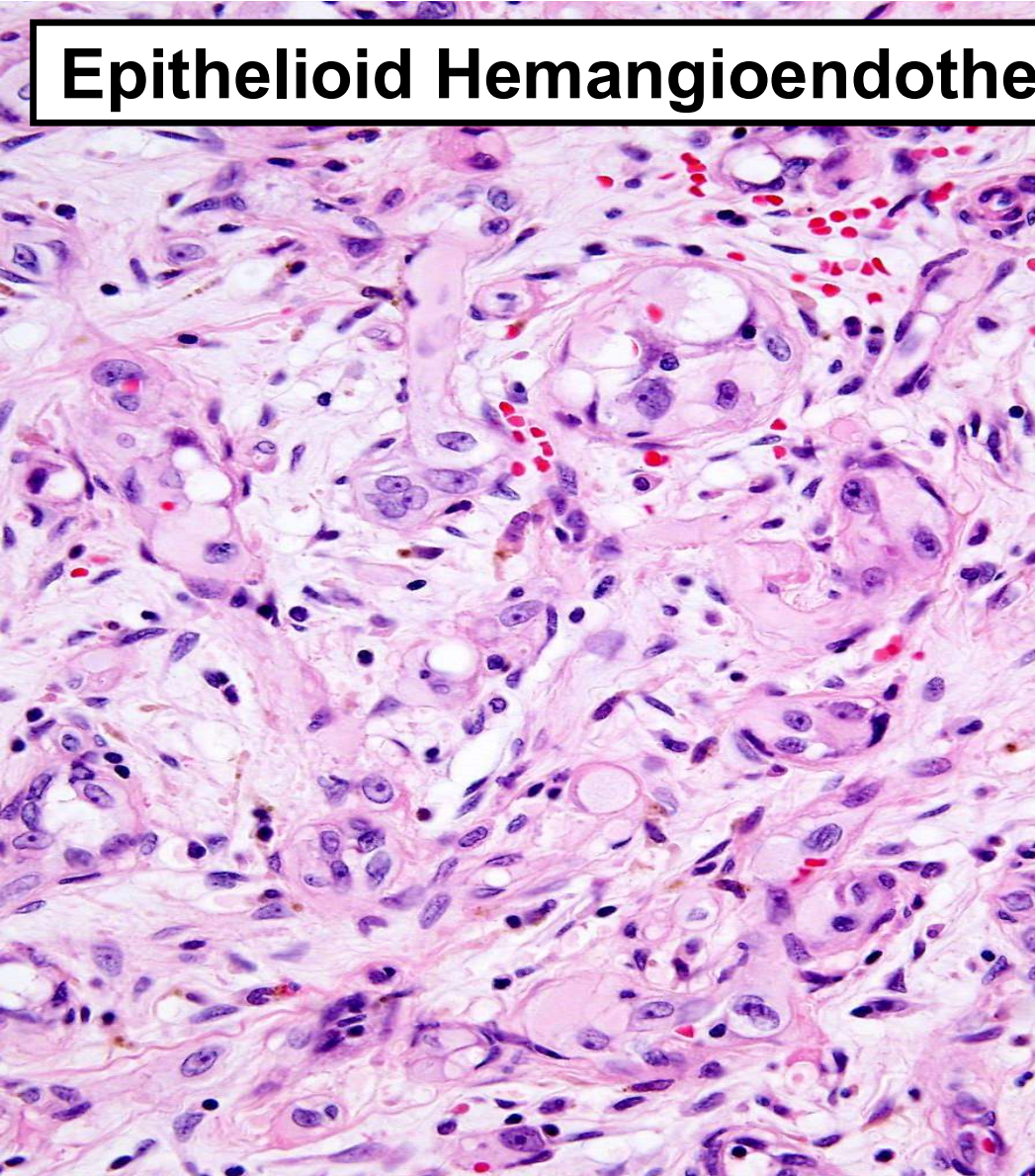


**Epithelioid Hemangioendothelioma Variant with *YAP1-TFE3***

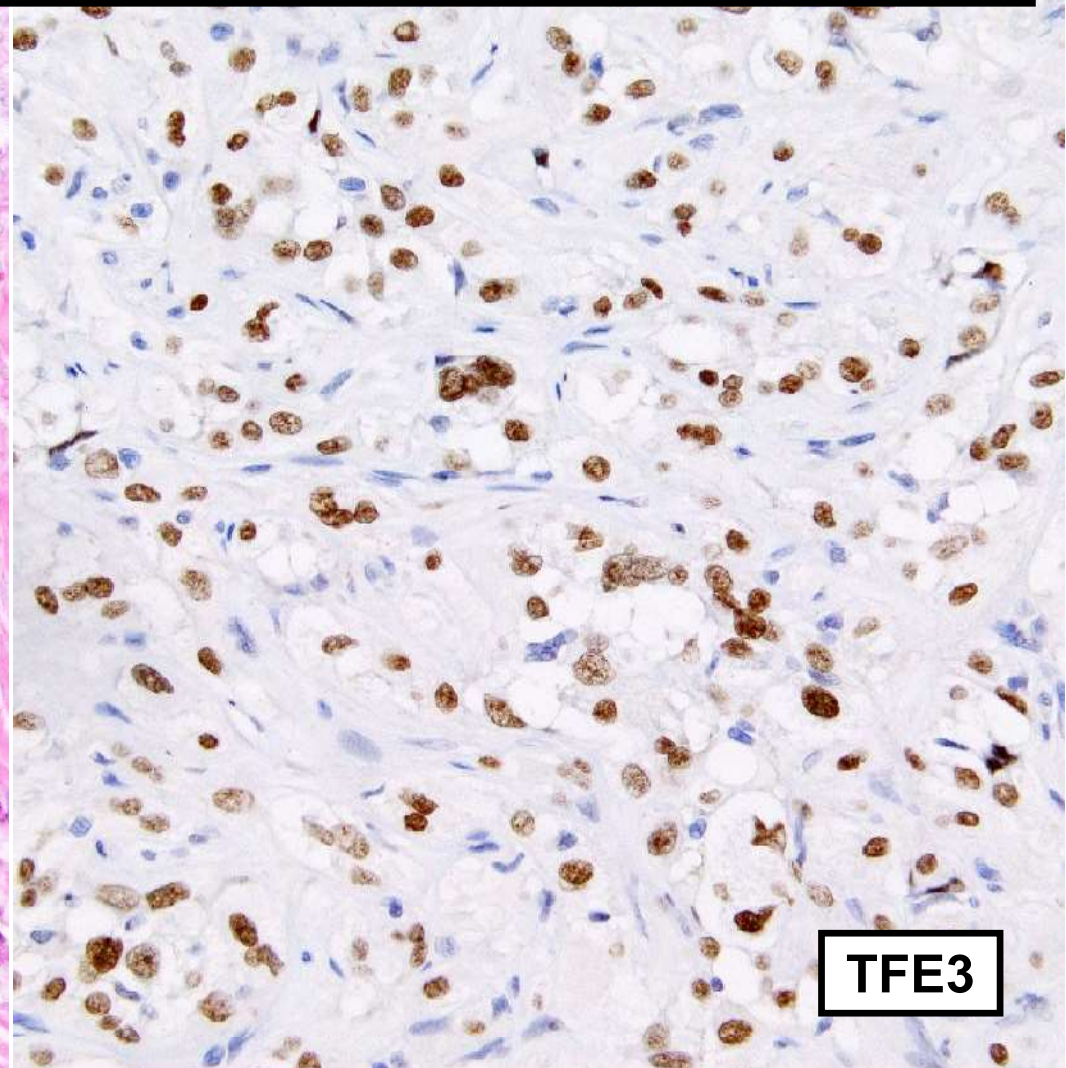
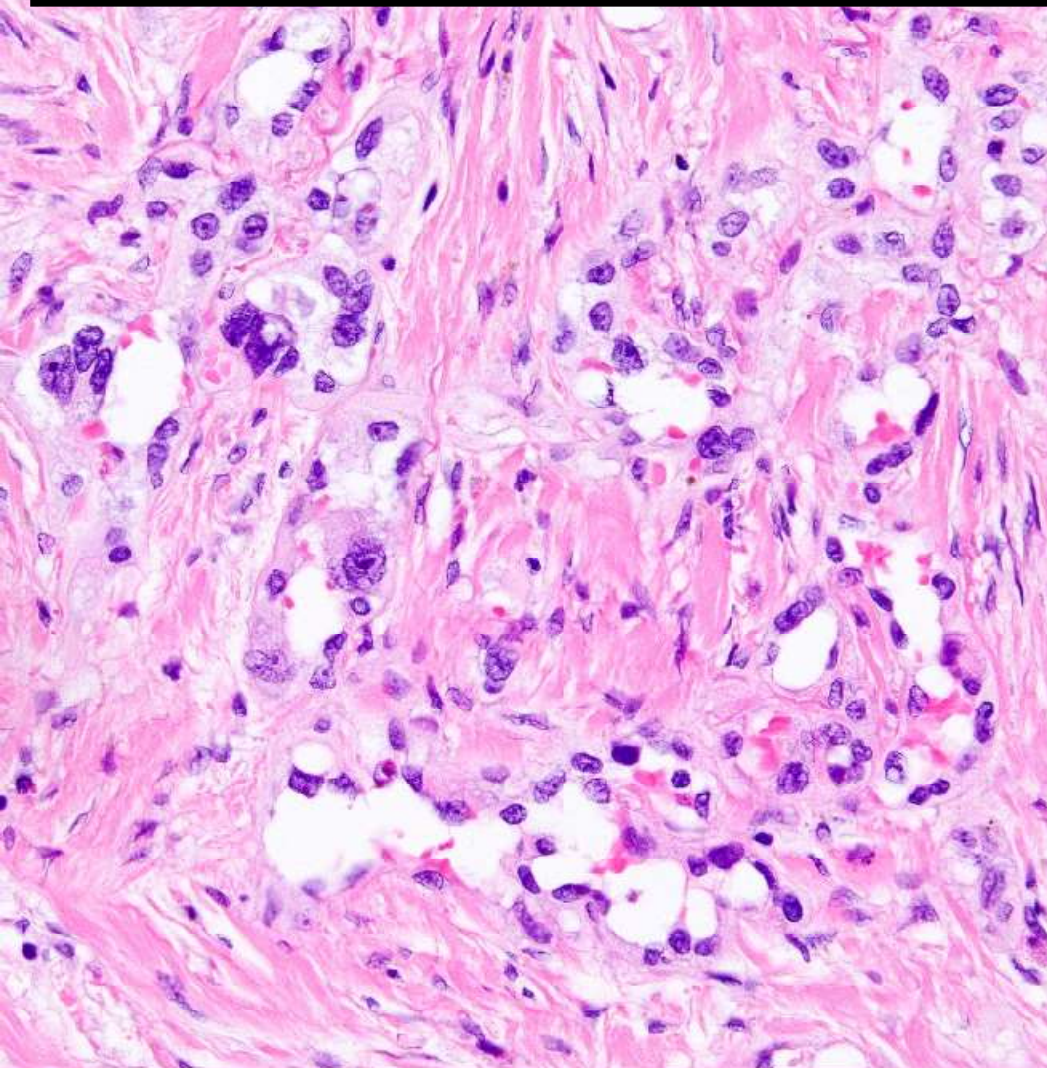


**CD31**

**Epithelioid Hemangioendothelioma Variant with *YAP1-TFE3***





# Epithelioid Hemangioendothelioma Variant with *YAP1-TFE3*

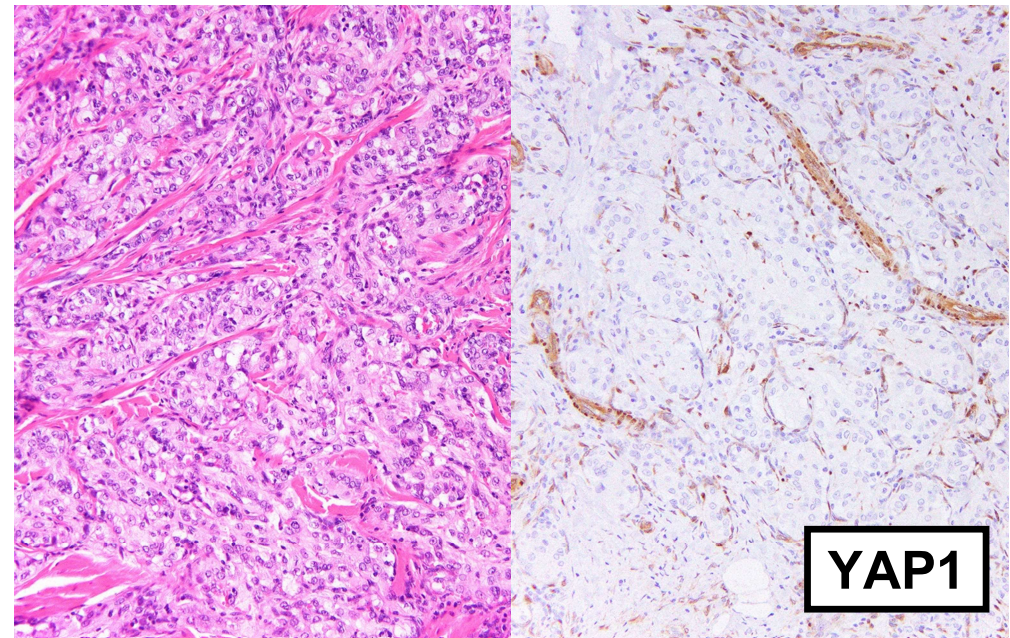
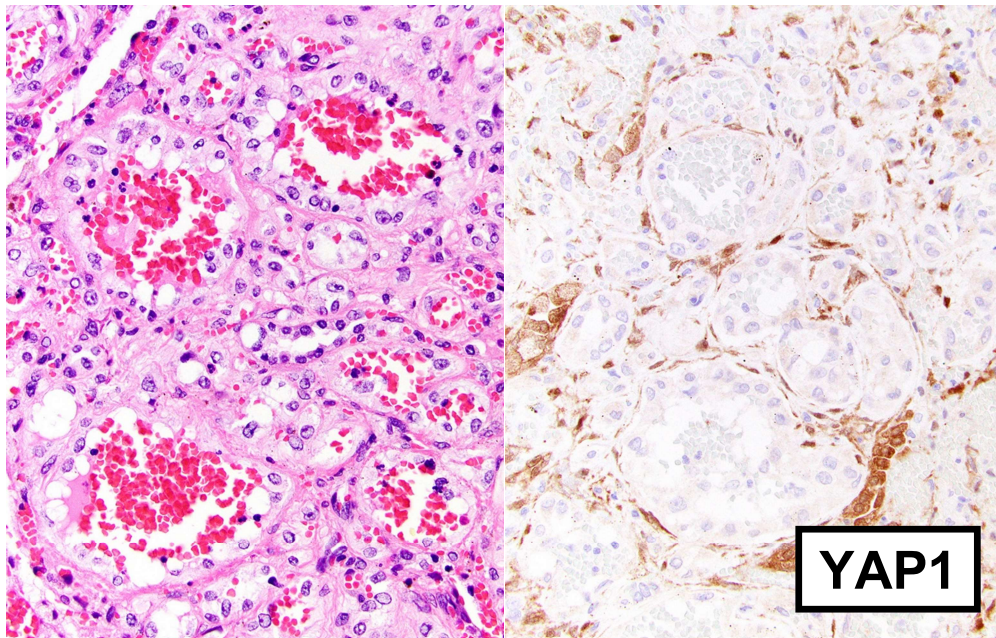


TFE3

# Loss of expression of YAP1 C-terminus as an ancillary marker for epithelioid hemangioendothelioma variant with *YAP1-TFE3* fusion and other YAP1-related vascular neoplasms

William J. Anderson<sup>1</sup>, Christopher D. M. Fletcher<sup>1</sup> and Jason L. Hornick<sup>1</sup>  

Modern Pathology (2021) 34:2036 – 2042



# WHO 2020: NEW CONCEPTS

<b>Tumor types</b>	<b>What's new?</b>
<b>Malignant melanotic nerve sheath tumor</b>	<b>New nomenclature</b>
<b>Solitary fibrous tumor</b>	<b>Risk stratification</b>

# **Malignant Melanotic Nerve Sheath Tumor**

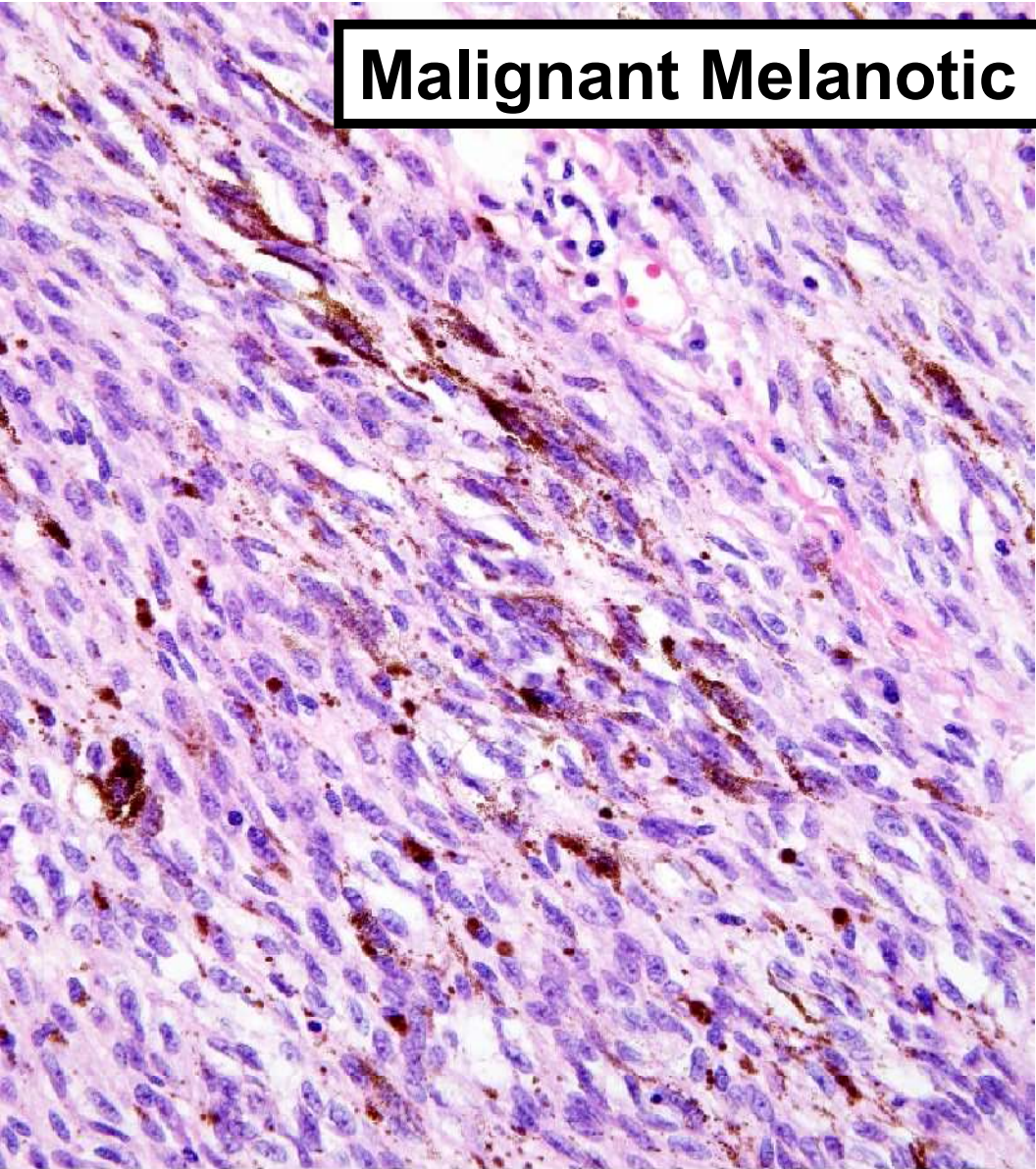
- **Previously known as melanotic schwannoma**
- **Rare peripheral nerve sheath tumor composed of Schwann cells with melanocytic differentiation**
- **Usually associated with spinal or autonomic nerves**
- **Variable association with Carney complex**
- **Locally aggressive with significant metastatic potential**
- **Behavior difficult to predict**

# Malignant Melanotic Nerve Sheath Tumor

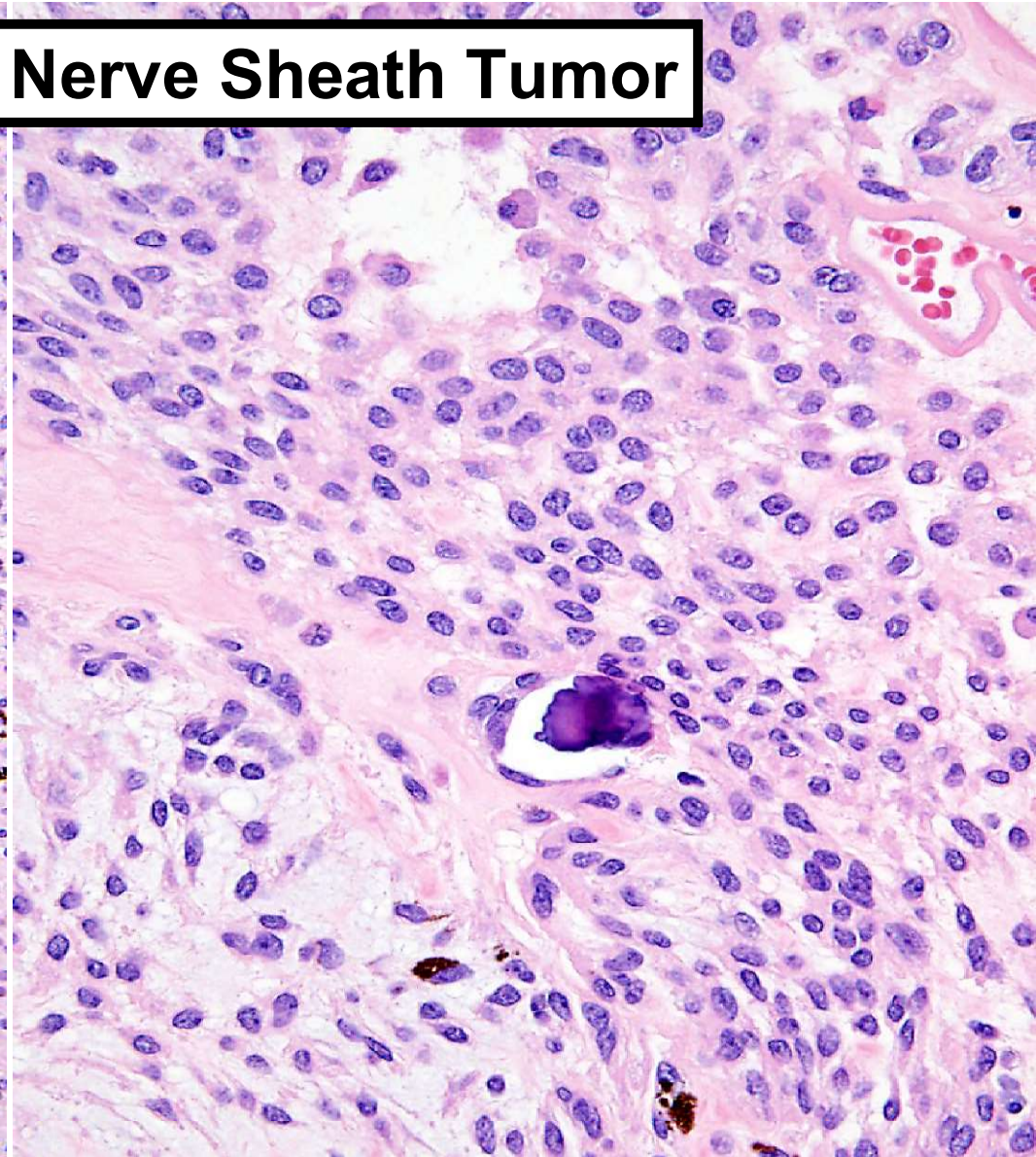
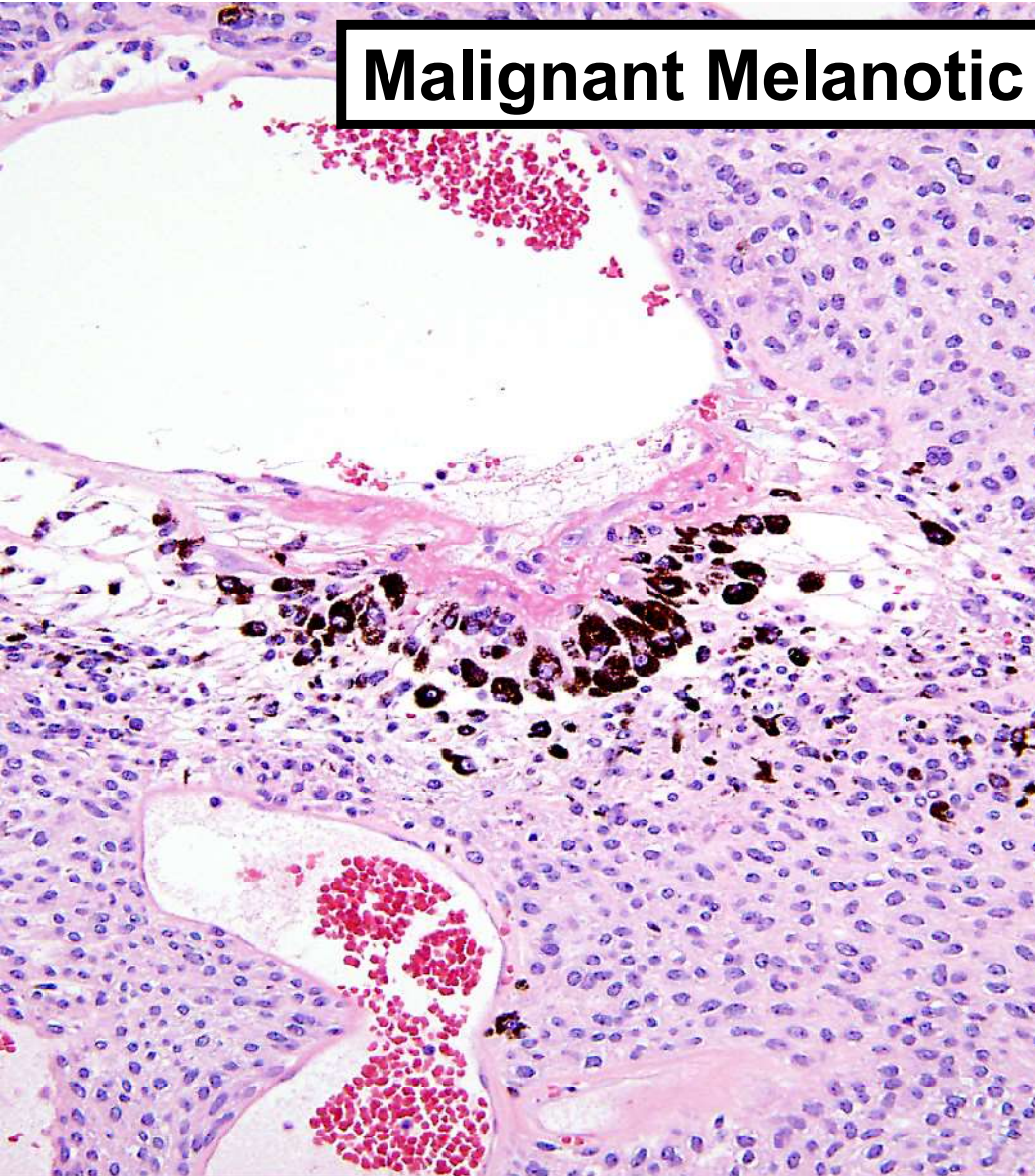




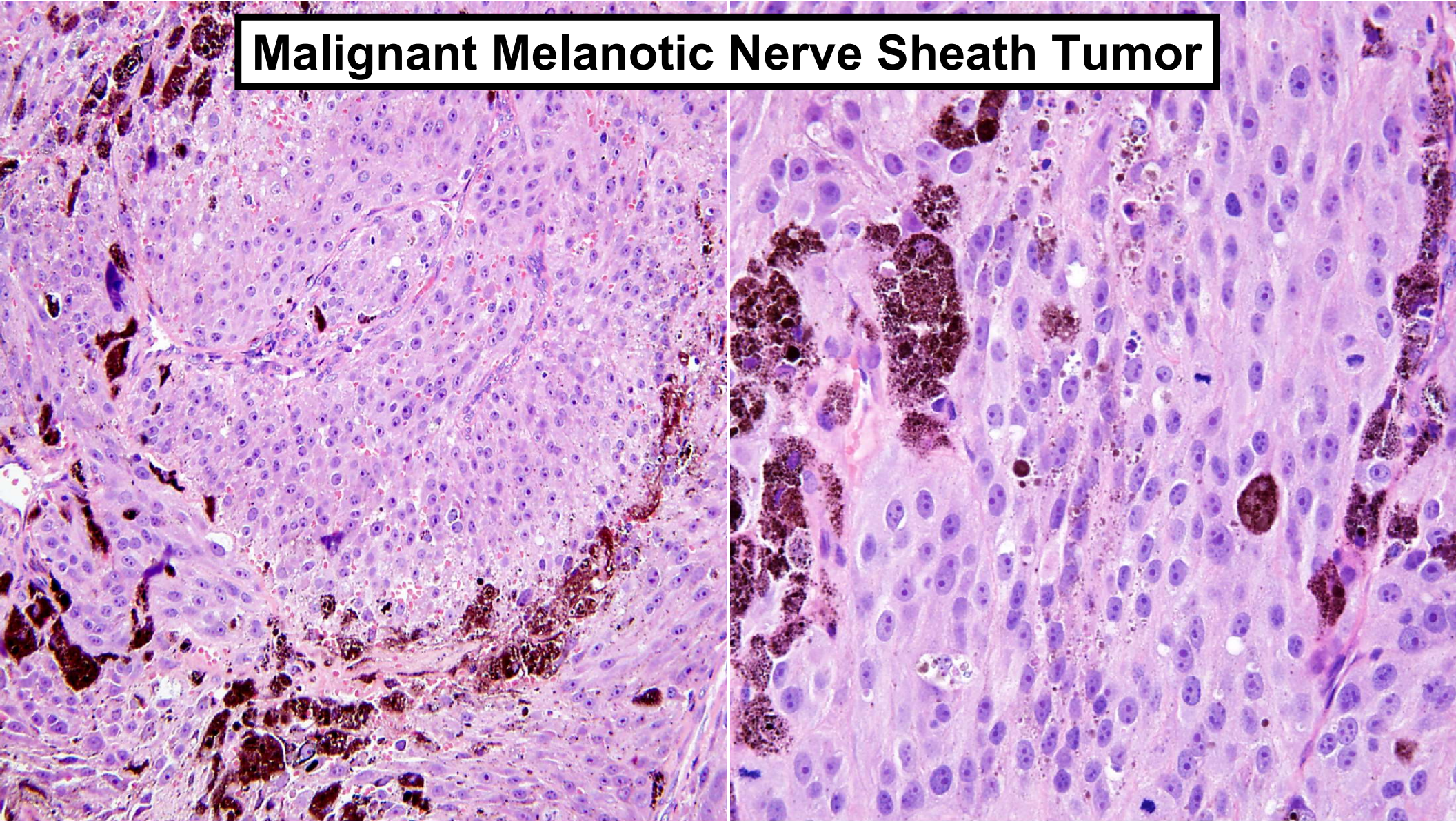
# Malignant Melanotic Nerve Sheath Tumor



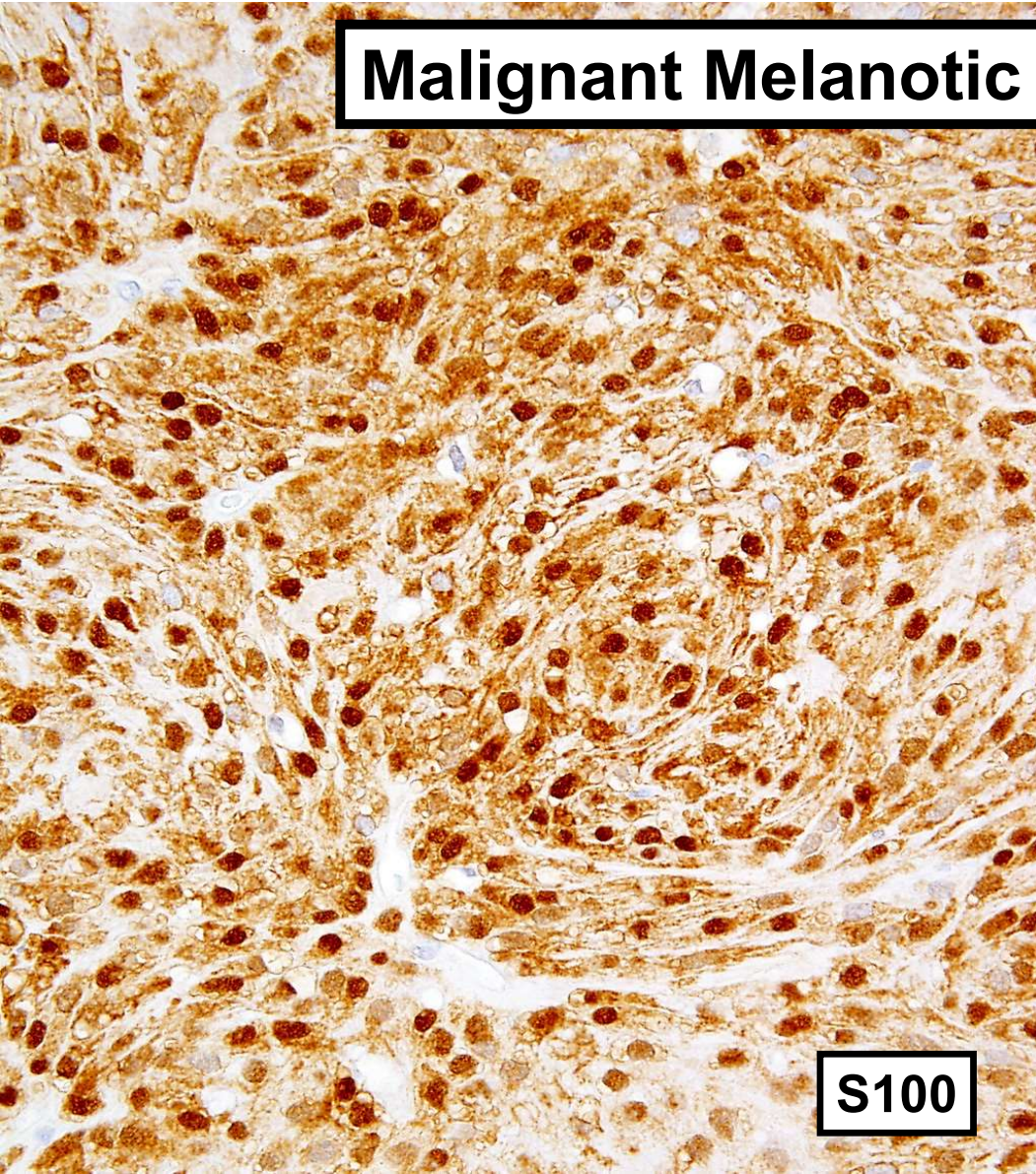
# Malignant Melanotic Nerve Sheath Tumor



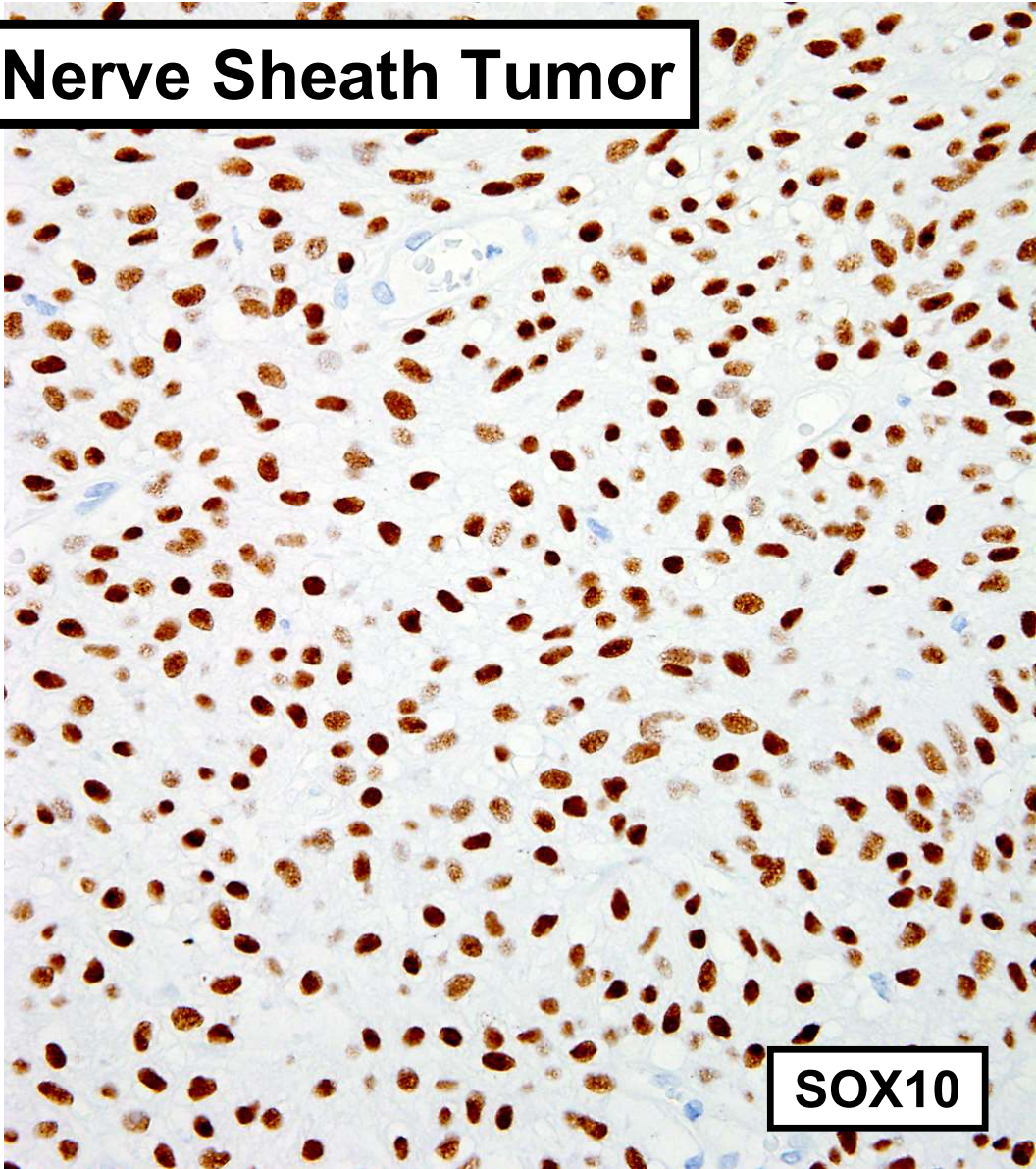
# Malignant Melanotic Nerve Sheath Tumor



# Malignant Melanotic Nerve Sheath Tumor



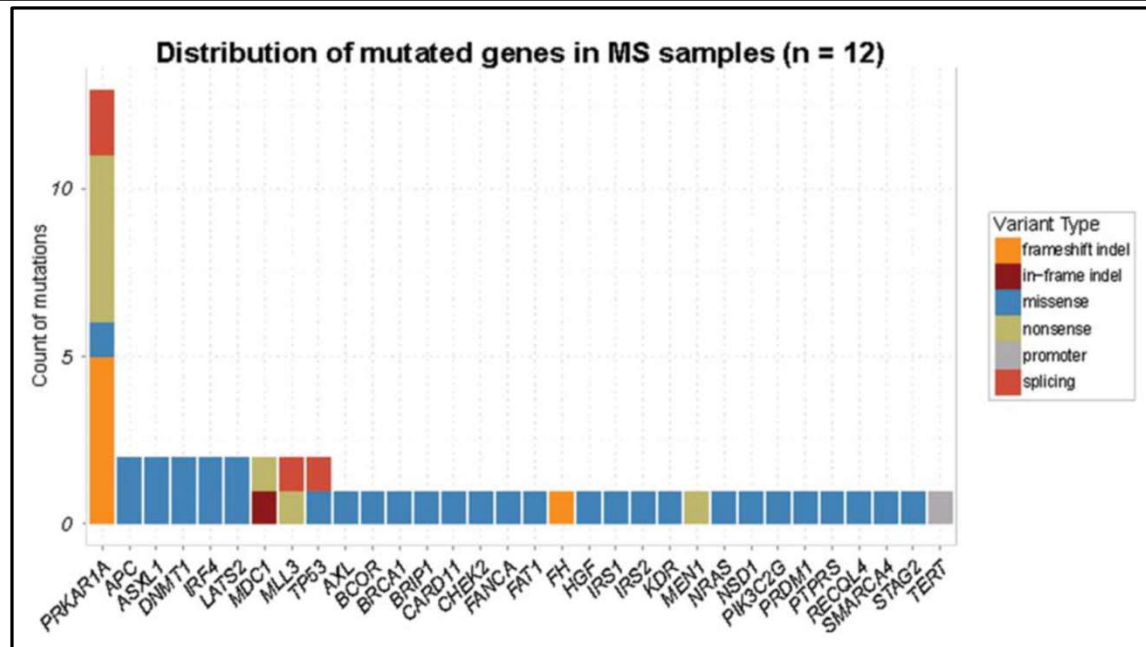
S100



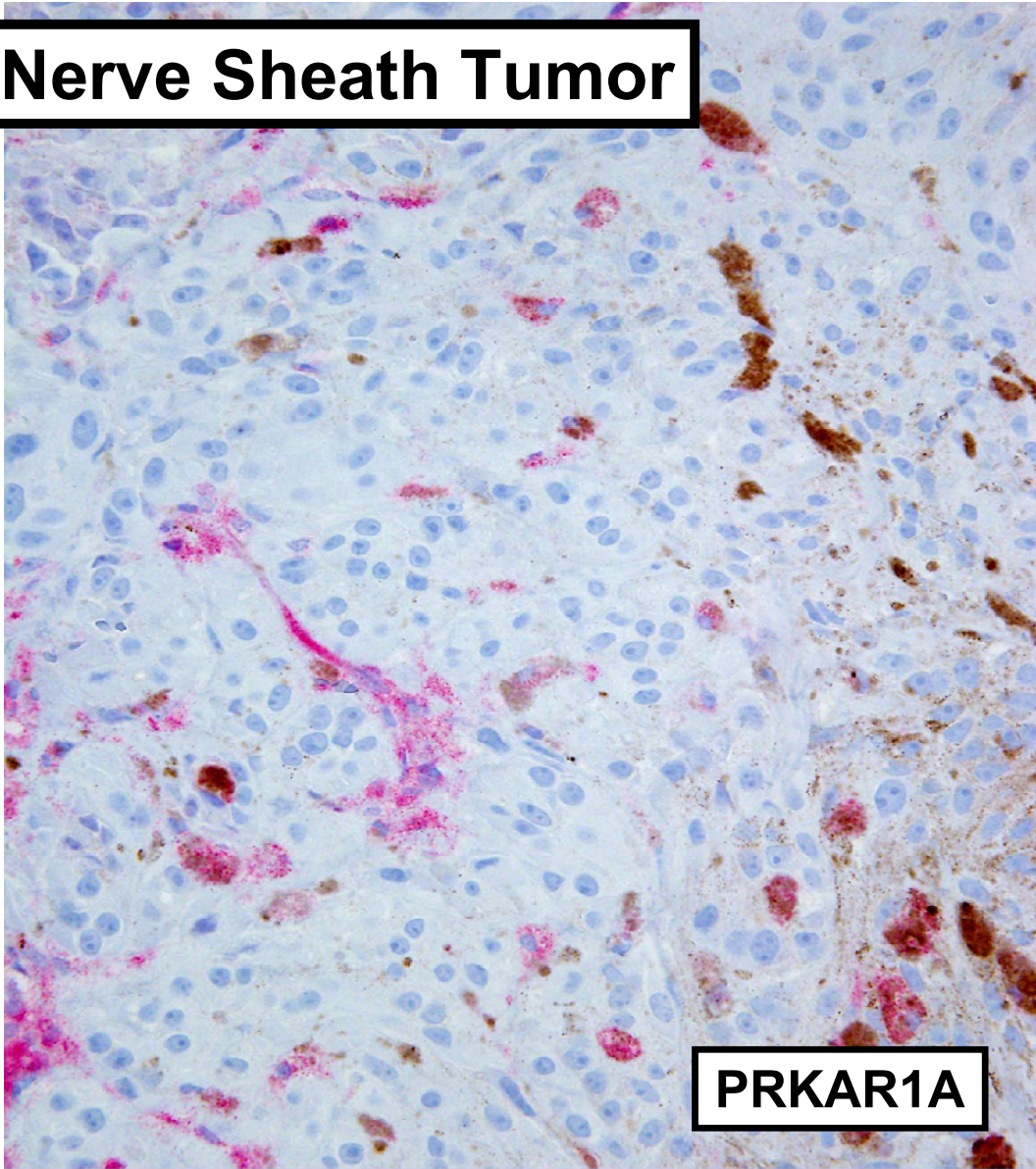
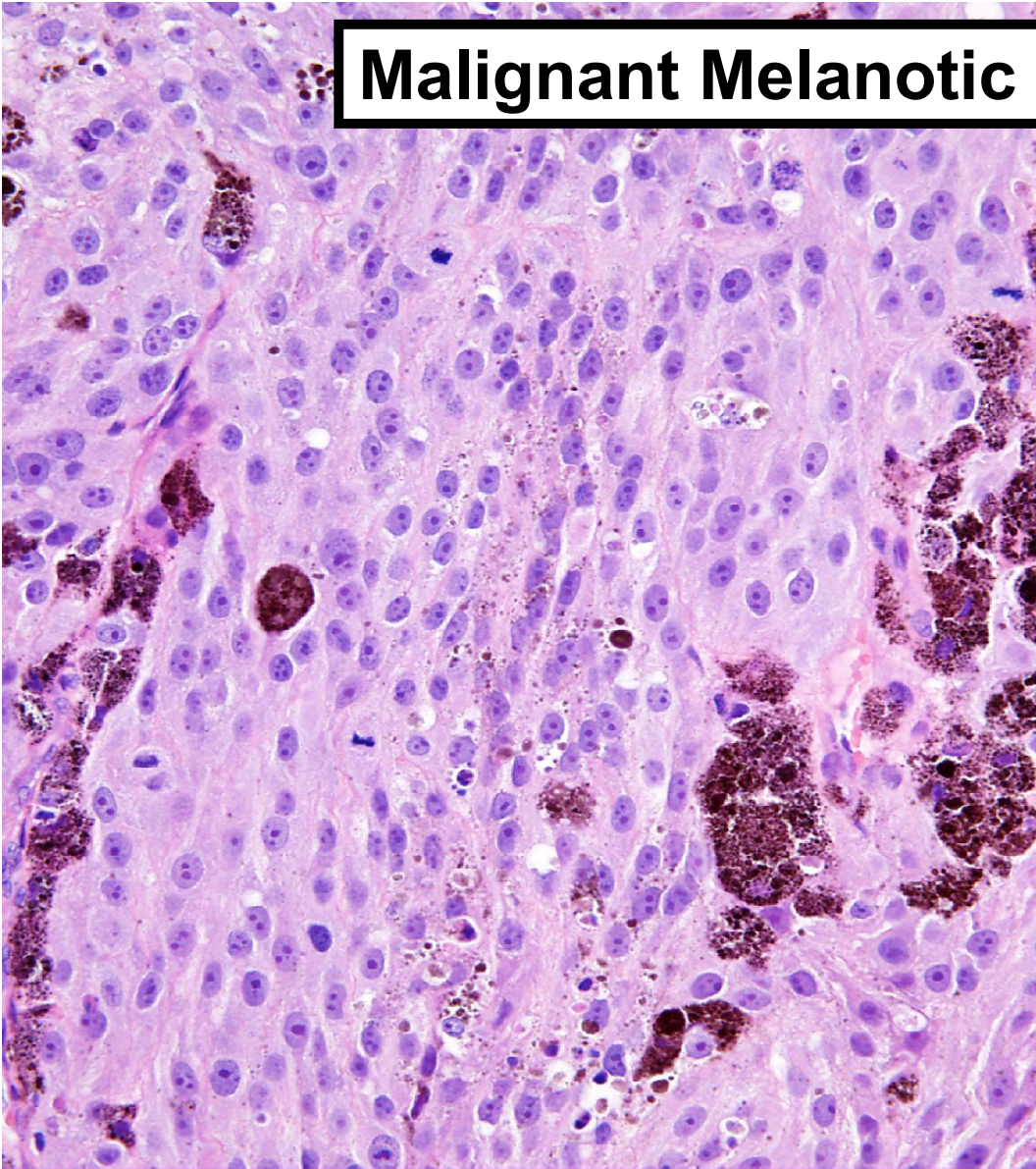
SOX10

# Consistent Copy Number Changes and Recurrent *PRKARIA* Mutations Distinguish Melanotic Schwannomas from Melanomas: SNP-Array and Next Generation Sequencing Analysis

Lu Wang,<sup>1</sup> Ahmet Zehir,<sup>1</sup> Justyna Sadowska,<sup>1</sup> Nengyi Zhou,<sup>1</sup> Marc Rosenblum,<sup>1</sup> Klaus Busam,<sup>1</sup> Narasimhan Agaram,<sup>1</sup> William Travis,<sup>1</sup> Maria Arcila,<sup>1</sup> Snjezana Dogan,<sup>1</sup> Michael F. Berger,<sup>1,2</sup> Donovan T. Cheng,<sup>1</sup> Marc Ladanyi,<sup>1,2</sup> Khedoudja Nafa,<sup>1</sup> and Meera Hameed<sup>1\*</sup>



# Malignant Melanotic Nerve Sheath Tumor



**PRKAR1A**

*Am J Surg Pathol* • Volume 38, Number 1, January 2014

## Malignant Melanotic Schwannian Tumor

*A Clinicopathologic, Immunohistochemical, and Gene Expression Profiling Study of 40 Cases, With a Proposal for the Reclassification of “Melanotic Schwannoma”*

*Jorge Torres-Mora, MD,\* Sarah Dry, MD,† Xinmin Li, PhD,† Scott Binder, MD,† Mitul Amin, MD,‡ and Andrew L. Folpe, MD\**

**Local recurrence: 35%**  
**Metastasis: 44%**

**TABLE 2.** Correlation of Pathologic Features With Metastases

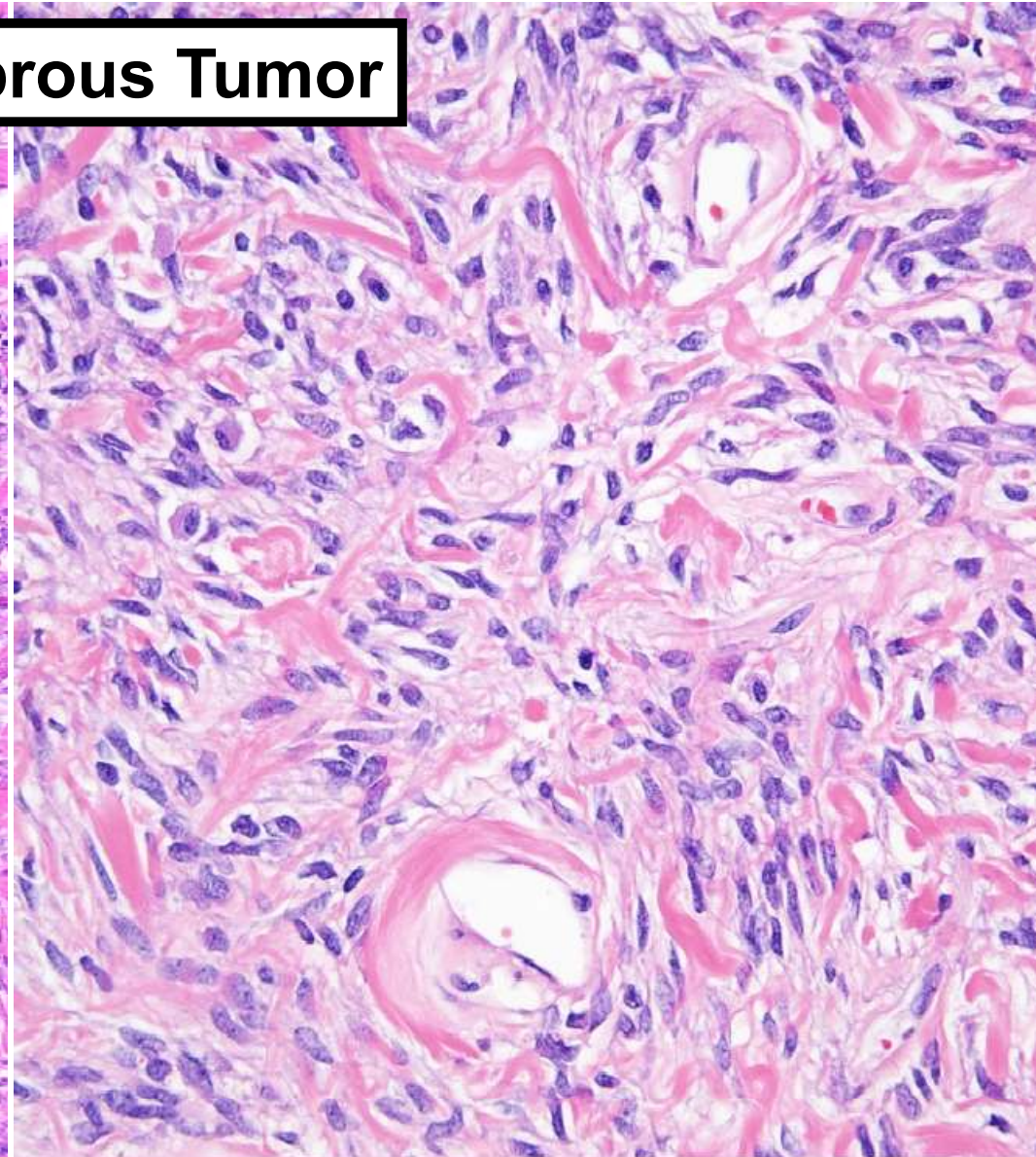
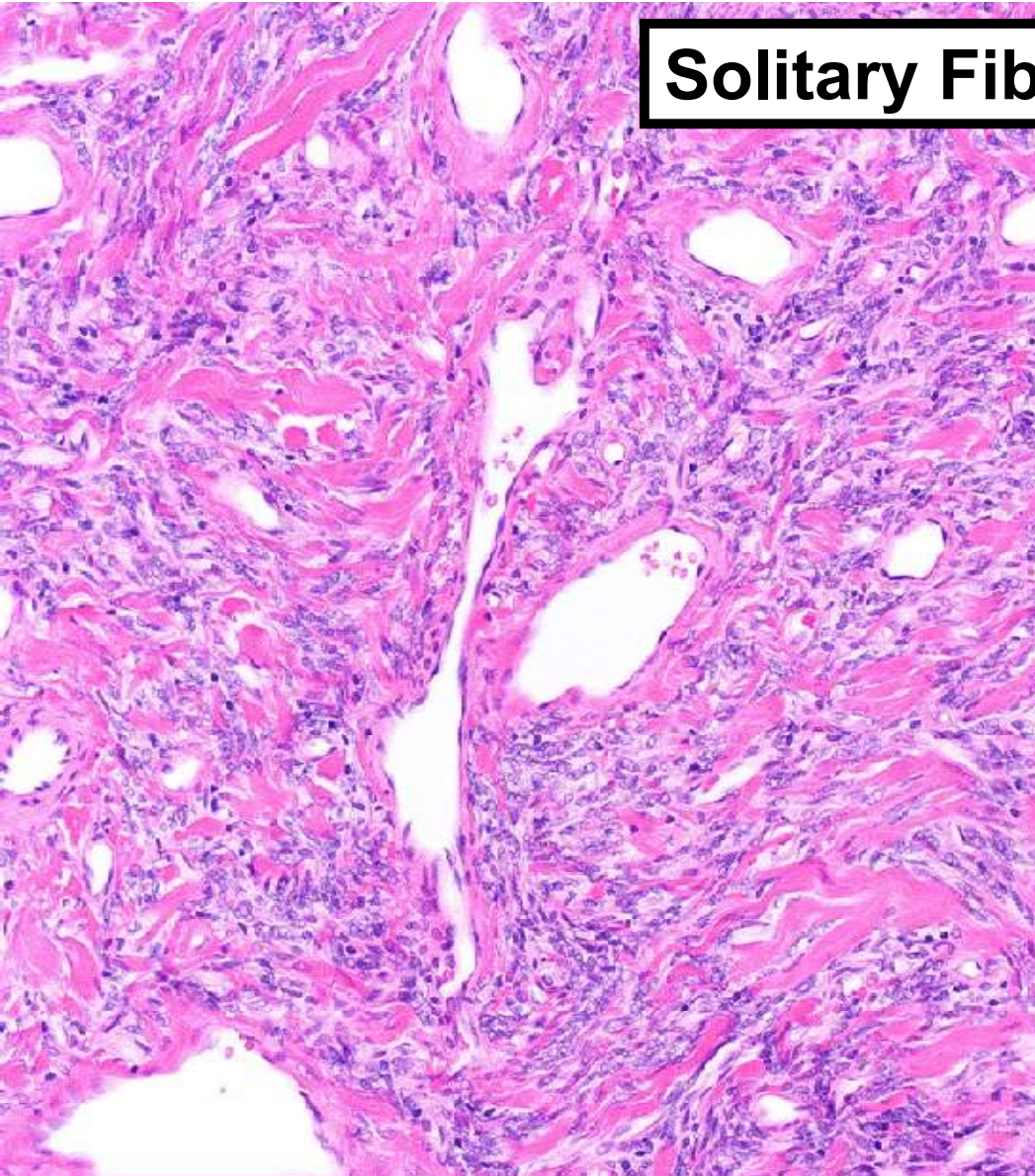
Pathologic Feature	Metastases	<i>P</i> (Fisher Exact Test)
Mitotic activity > 1/10 HPF		
Present	5/5	0.008
Absent	6/20	
Necrosis		
Present	3/7	1 (NS)
Absent	8/18	
Macronucleoli		
Present	5/11	1 (NS)
Absent	6/14	
Small cell change		
Present	2/3	0.56 (NS)
Absent	9/22	
Nuclear pleomorphism		
Present	4/8	1 (NS)
Absent	7/17	
Psammoma bodies		
Present	4/10	1 (NS)
Absent	7/15	

# **Solitary Fibrous Tumor**

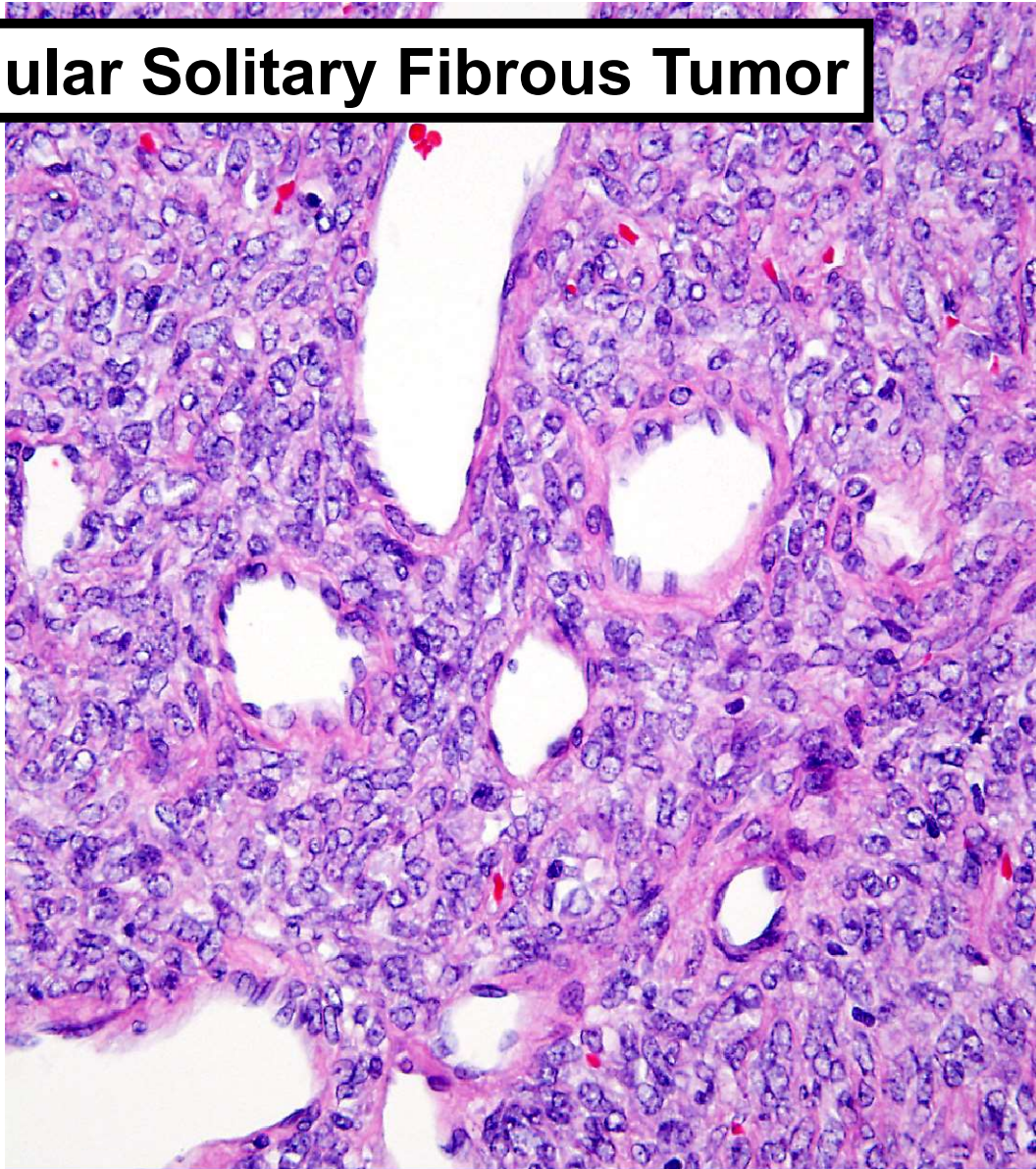
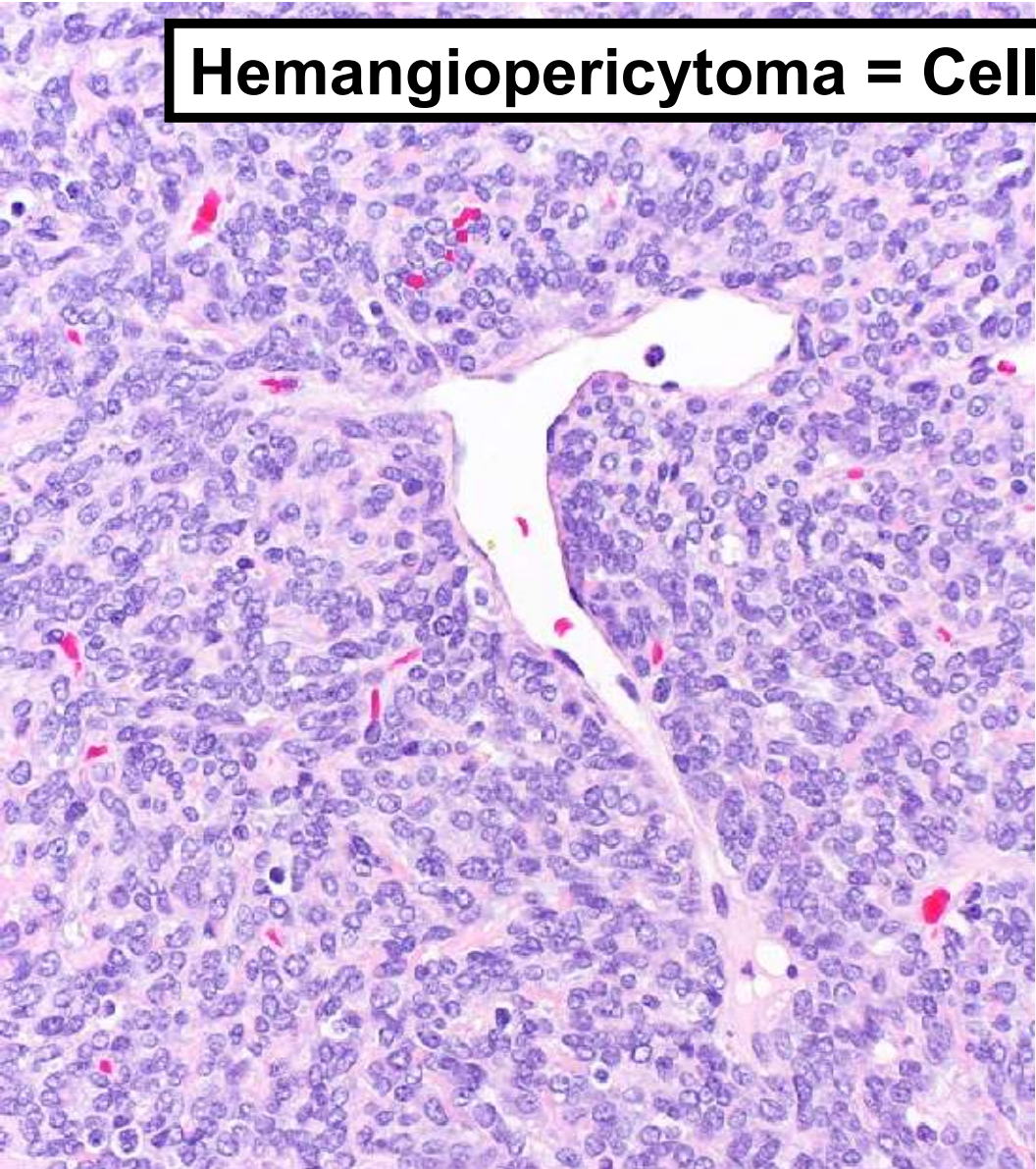
- **Anatomically ubiquitous fibroblastic neoplasm (pleura, retroperitoneum, abdomen, head & neck)**
- **“Patternless” architecture, varying cellularity, prominent stromal collagen, dilated branching (“staghorn”) vessels**
- **“Hemangiopericytoma” synonymous with SFT (uniform hypercellularity)**
- **CD34 positive in 95% of cases, but not specific**
- **Notoriously difficult to predict clinical behavior**
- **Mitotic rate  $\geq 4$  per 10 HPF = malignant?**



# Solitary Fibrous Tumor



**Hemangiopericytoma = Cellular Solitary Fibrous Tumor**



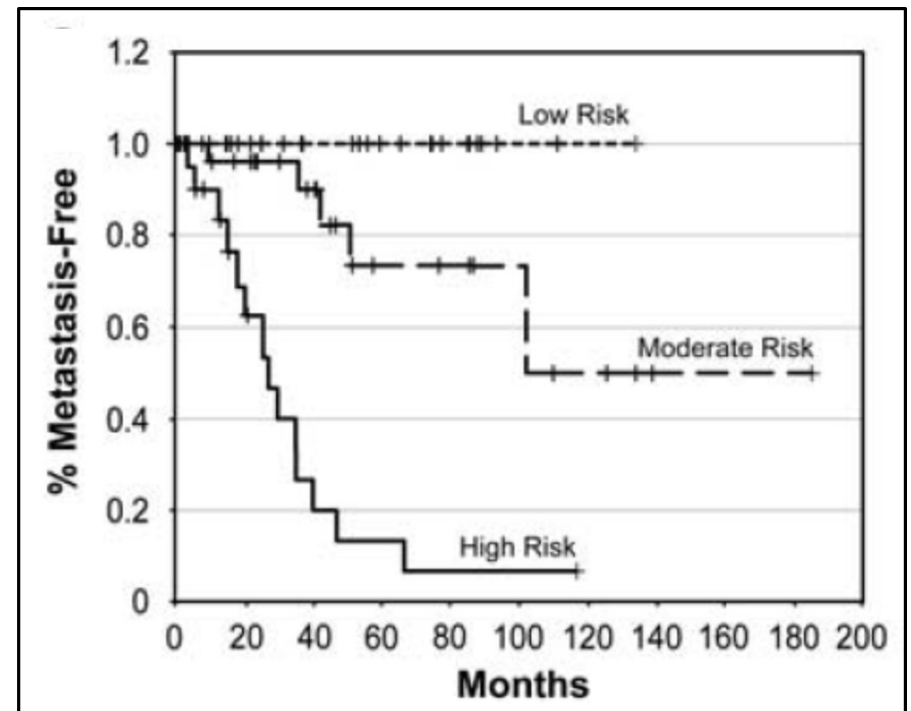
MODERN PATHOLOGY (2012) 25, 1298–1306

## Solitary fibrous tumor: a clinicopathological study of 110 cases and proposed risk assessment model

Elizabeth G Demicco<sup>1</sup>, Min S Park<sup>2</sup>, Dejka M Araujo<sup>2</sup>, Patricia S Fox<sup>3</sup>, Roland L Bassett<sup>3</sup>, Raphael E Pollock<sup>4</sup>, Alexander J Lazar<sup>1,5</sup> and Wei-Lien Wang<sup>1</sup>

**Table 5** Risk stratification model

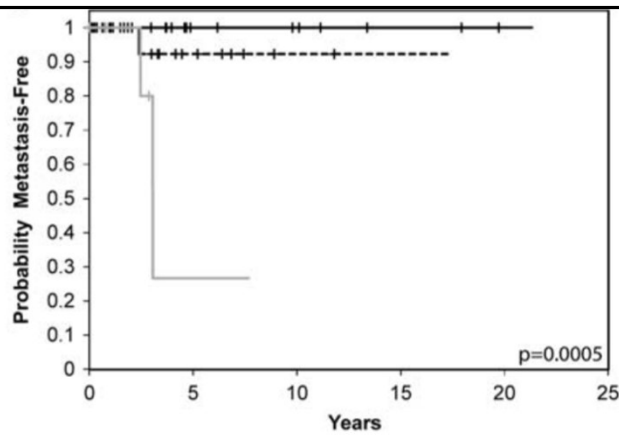
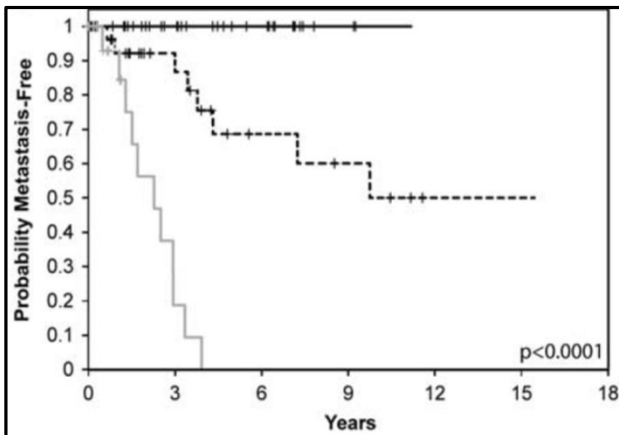
Risk factor	Score
<i>Age</i>	
<55	0
≥55	1
<i>Tumor size (cm)</i>	
<5	0
5 to <10	1
10 to <15	2
≥15	3
<i>Mitotic figures (/10 high-power fields)</i>	
0	0
1–3	1
≥4	2
<i>Risk</i>	<i>Total score</i>
Low	0–2
Moderate	3–4
High	5–6



# Risk assessment in solitary fibrous tumors: validation and refinement of a risk stratification model

Elizabeth G Demicco<sup>1</sup>, Michael J Wagner<sup>2</sup>, Robert G Maki<sup>3,4</sup>, Vishal Gupta<sup>5</sup>, Ilya Iofin<sup>6</sup>, Alexander J Lazar<sup>7</sup> and Wei-Lien Wang<sup>7</sup>

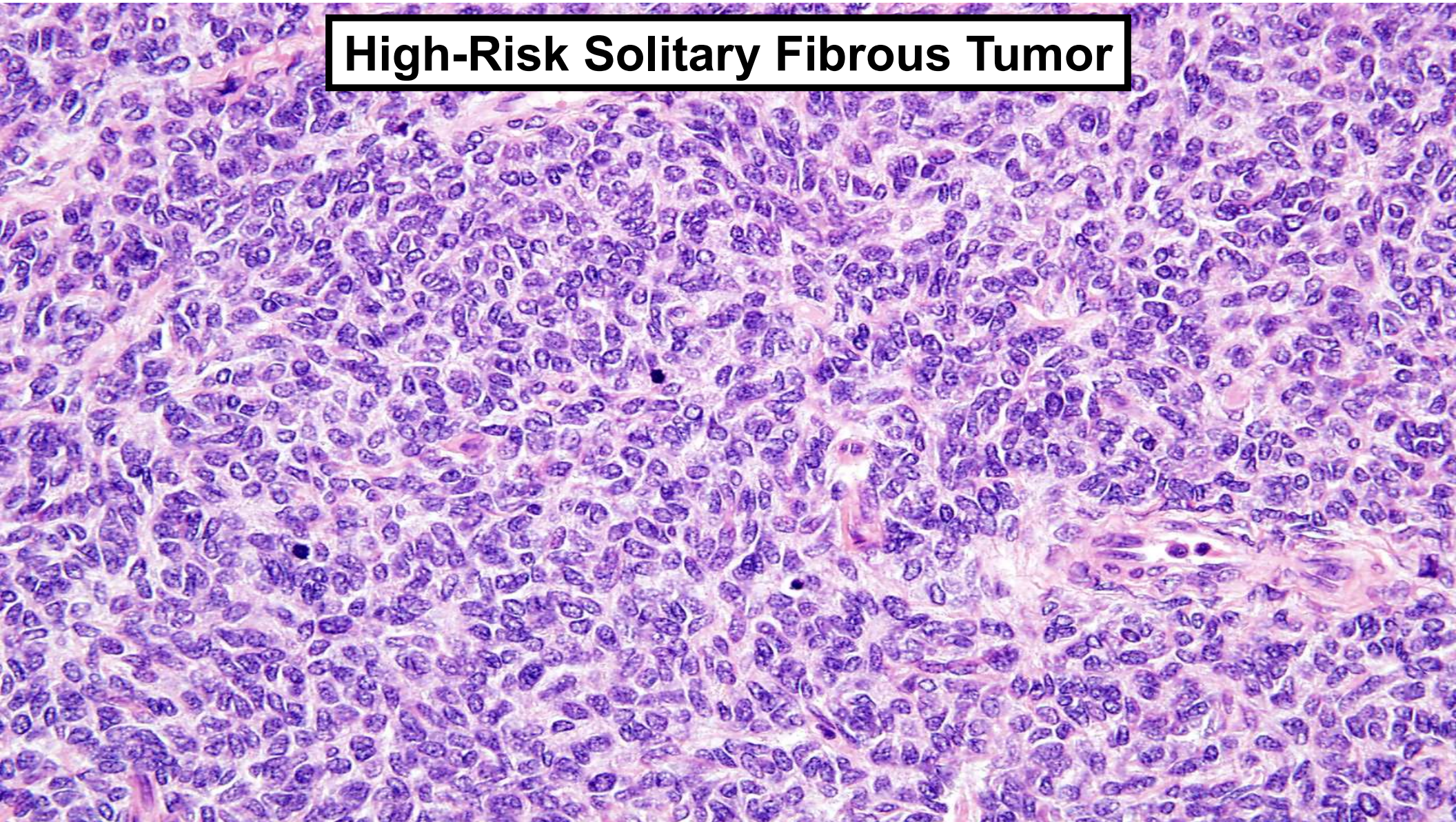
Risk factor	Score
<i>Age</i>	
< 55	0
≥ 55	1
<i>Tumor size (cm)</i>	
< 5	0
5 to < 10	1
10 to < 15	2
≥ 15	3
<i>Mitotic count (/10 high-power fields)</i>	
0	0
1–3	1
≥ 4	2
<i>Tumor necrosis</i>	
< 10%	0
≥ 10%	1
<i>Risk class</i>	<i>Total score</i>
Low	0–3
Intermediate	4–5
High	6–7



+ Low Risk (n=37)  
 + Intermediate Risk (n=30)  
 + High Risk (n=15)

+ Low Risk (n=28)  
 + Intermediate Risk (n=15)  
 + High Risk (n=7)

# High-Risk Solitary Fibrous Tumor



# WHO 2020: NEW GENETICS

## Fibroblastic/myofibroblastic tumors

Tumor type	New genetic alterations
Fibrous hamartoma of infancy	<i>EGFR</i> mutations
Calcifying aponeurotic fibroma	<i>FN1::EGF</i>
Lipofibromatosis	<i>FN1::EGF</i> , other RTK or EGFR ligand fusions
Dermatofibrosarcoma protuberans	<i>COL6A3::PDGFD</i> , <i>EMILIN2::PDGFD</i>
Solitary fibrous tumor	<i>NAB2::STAT6</i>
Myxoinflammatory fibroblastic sarcoma	<i>BRAF</i> fusions
Infantile fibrosarcoma	<i>EML4::NTRK3</i> , <i>NTRK1</i> , <i>NTRK2</i> , <i>BRAF</i> , <i>MET</i> fusions

## Pericytic (perivascular) tumors

Tumor type	New genetic alterations
Glomus tumor	<i>MIR143::NOTCH1/2/3</i>
Myopericytoma/myofibroma	<i>PDGFRB</i> mutations, <i>SRF::RELA</i> (cellular myofibroma)

# WHO 2020: NEW GENETICS

## Vascular tumors

Tumor type	New genetic alterations
Epithelioid hemangioma	<i>FOS</i> and <i>FOSB</i> fusions
Pseudomyogenic hemangioendothelioma	<i>SERPINE1::FOSB</i> , <i>ACTB::FOSB</i>

## Skeletal muscle tumors

Tumor type	New genetic alterations
Spindle cell/sclerosing rhabdomyosarcoma	<i>MYOD1</i> mutations (adolescents/adults)
	<i>SRF::NCOA2</i> , <i>TEAD1::NCOA2</i> , <i>VGLL2::NCOA2</i> , <i>VGLL2::CITED2</i> (congenital/infantile)
	<i>EWSR1::TFCP2</i> , <i>FUS::TFCP2</i> (intraosseous; spindle cell and epithelioid)

# WHO 2020: NEW GENETICS

## Vascular tumors

Tumor type	New genetic alterations
Epithelioid hemangioma	<i>FOS</i> and <i>FOSB</i> fusions
Pseudomyogenic hemangioendothelioma	<i>SERPINE1::FOSB</i> , <i>ACTB::FOSB</i>

## Skeletal muscle tumors

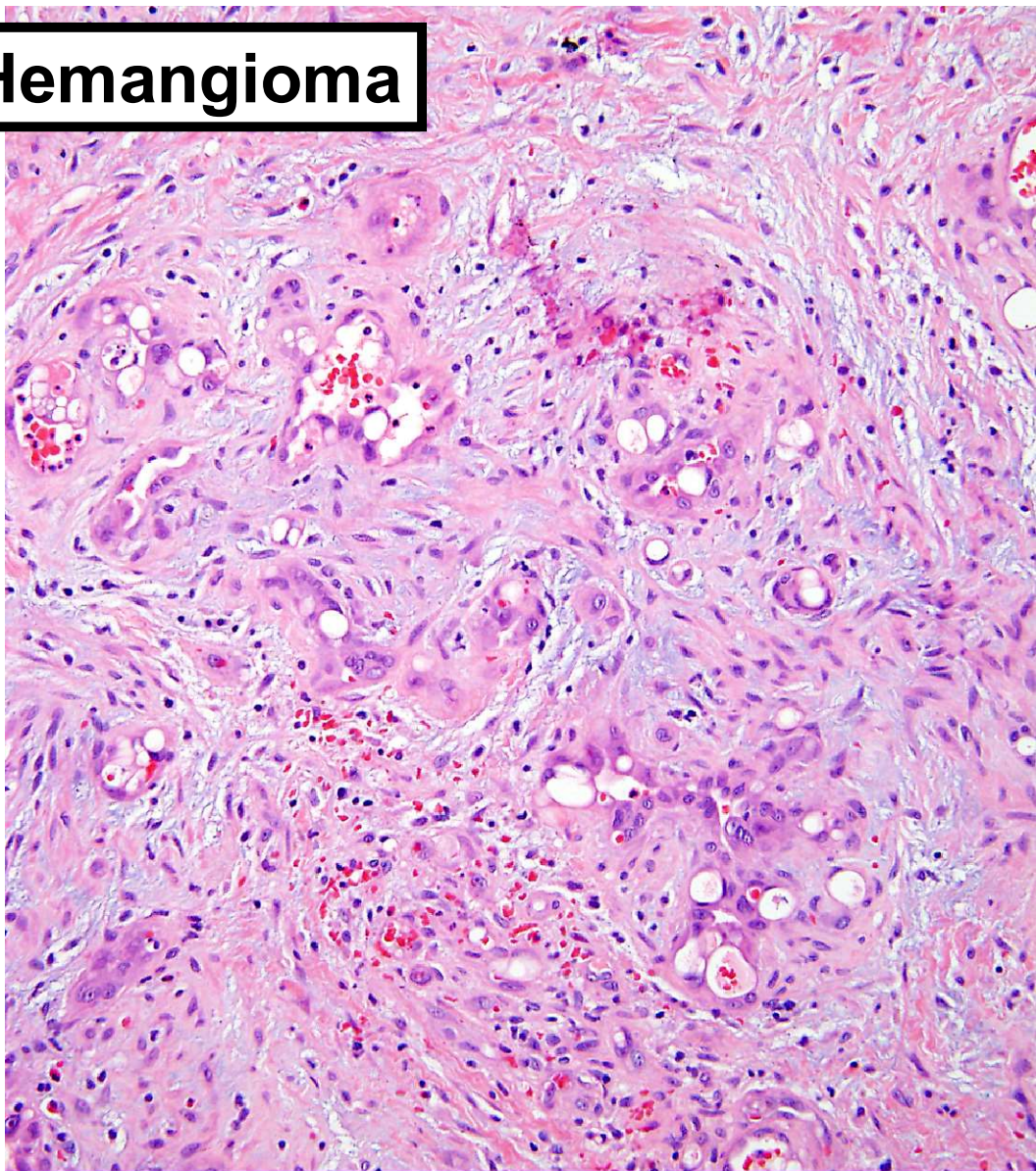
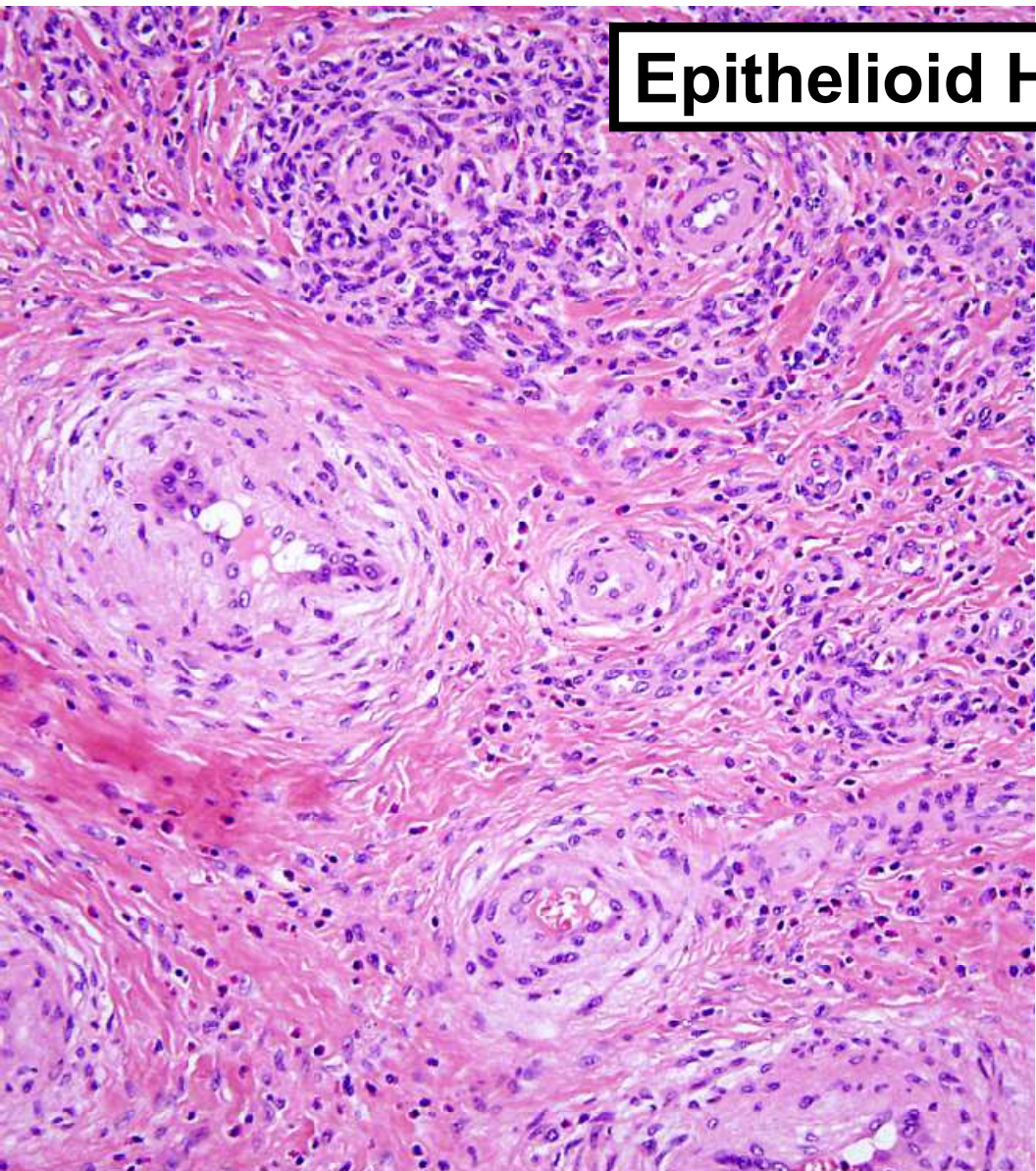
Tumor type	New genetic alterations
Spindle cell/sclerosing rhabdomyosarcoma	<i>MYOD1</i> mutations (adolescents/adults)
	<i>SRF::NCOA2</i> , <i>TEAD1::NCOA2</i> , <i>VGLL2::NCOA2</i> , <i>VGLL2::CITED2</i> (congenital/infantile)
	<i>EWSR1::TFCP2</i> , <i>FUS::TFCP2</i> (intraosseous; spindle cell and epithelioid)



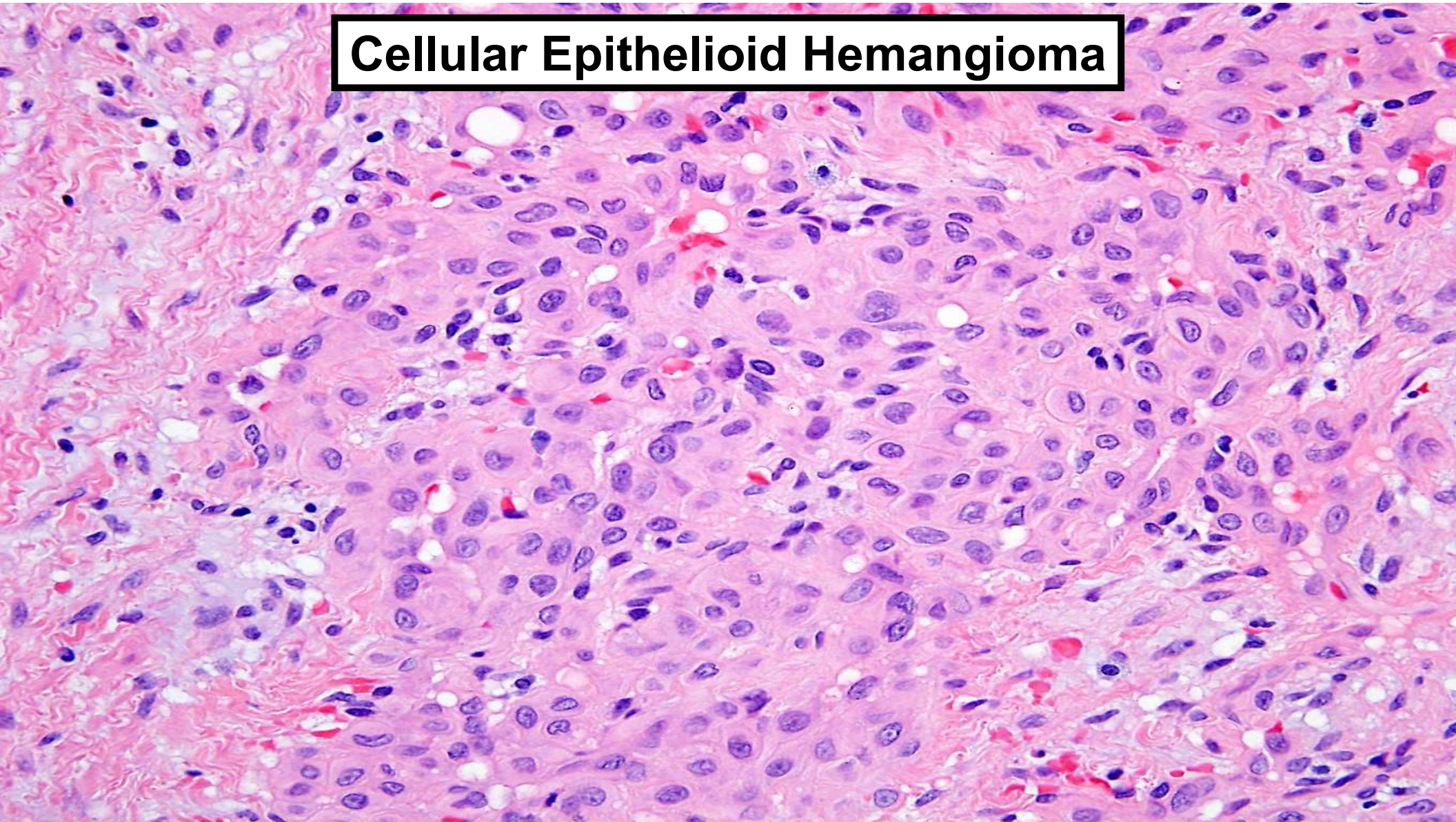
# Epithelioid Hemangioma

- **Subset also known as angiolymphoid hyperplasia with eosinophilia (head and neck)**
- **Head and neck (especially peri-auricular), bone, penis**
- **Small subcutaneous nodule +/- tender or pruritic**
- **20% multiple lesions**
- **Up to 1/3 recur locally**
- **Cellular examples with poor canalization may be misdiagnosed as malignant (but minimal nuclear atypia)**

# Epithelioid Hemangioma



## Cellular Epithelioid Hemangioma



**GENES, CHROMOSOMES & CANCER 53:951–959 (2014)**

## **ZFP36-FOSB Fusion Defines a Subset of Epithelioid Hemangioma with Atypical Features**

Cristina R Antonescu,<sup>1\*</sup> Hsiao-Wei Chen,<sup>1</sup> Lei Zhang,<sup>1</sup> Yun-Shao Sung,<sup>1</sup> David Panicek,<sup>2</sup> Narasimhan P Agaram,<sup>1</sup> Brendan C Dickson,<sup>3</sup> Thomas Krausz,<sup>4</sup> and Christopher D Fletcher<sup>5\*</sup>

## Frequent *FOS* Gene Rearrangements in Epithelioid Hemangioma

*A Molecular Study of 58 Cases With Morphologic Reappraisal*

Shih-Chiang Huang, MD,\* † Lei Zhang, MD, † Yun-Shao Sung, MSc, † Chun-Liang Chen, MSc, †  
Thomas Krausz, MD, ‡ Brendan C. Dickson, MD, § Yu-Chien Kao, MD, ||  
Narasimhan P. Agaram, MBBS, † Christopher D.M. Fletcher, MD, FRCPath, ¶  
and Cristina R. Antonescu, MD †

*Am J Surg Pathol* • Volume 39, Number 10, October 2015

**GENES, CHROMOSOMES & CANCER 54:565–574 (2015)**

## **Fusion Events Lead to Truncation of *FOS* in Epithelioid Hemangioma of Bone**

David G. P. van IJzendoorn,<sup>1</sup> Danielle de Jong,<sup>2</sup> Cleofe Romagosa,<sup>3</sup> Piero Picci,<sup>4</sup> Maria Serena Benassi,<sup>4</sup> Marco Gambarotti,<sup>4</sup> Soeren Daugaard,<sup>5</sup> Michiel van de Sande,<sup>6</sup> Karoly Szuhai,<sup>2</sup> and Judith V. M. G. Bovée<sup>1\*</sup>

# FOSB is a Useful Diagnostic Marker for Pseudomyogenic Hemangioendothelioma

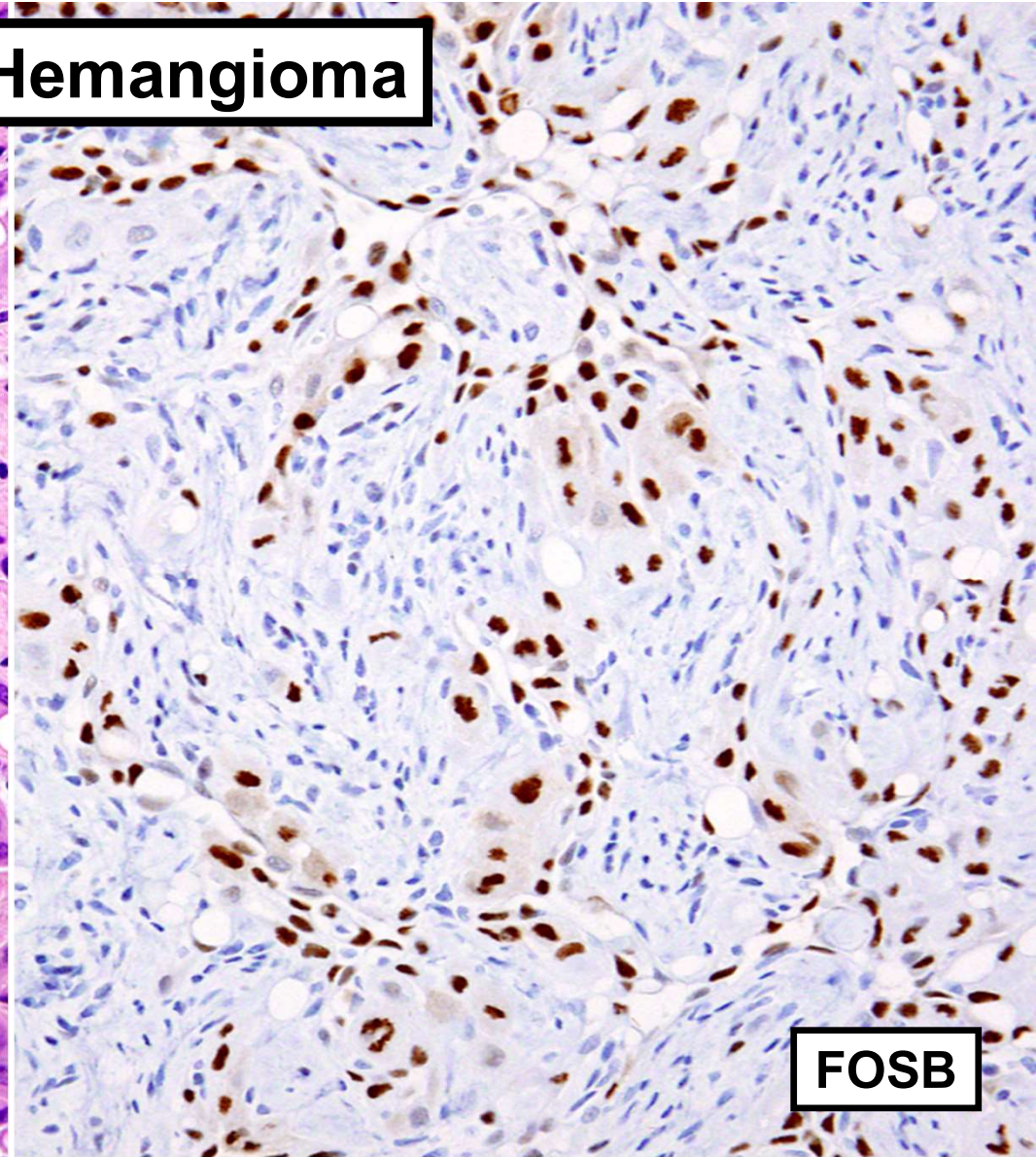
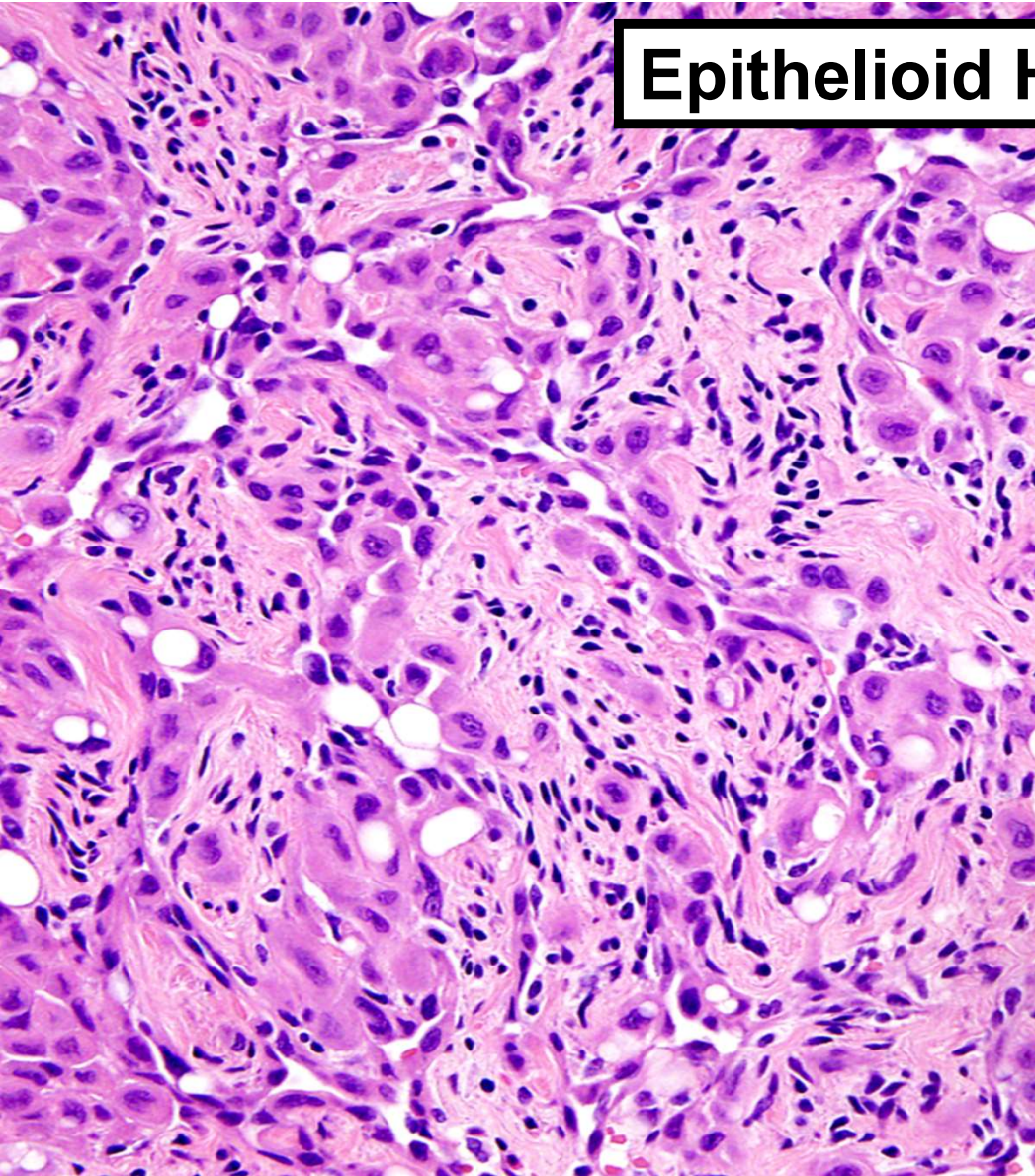
*Yin P. Hung, MD, PhD, Christopher D.M. Fletcher, MD, FRCPath,  
and Jason L. Hornick, MD, PhD*

*Am J Surg Pathol • Volume 41, Number 5, May 2017*

**TABLE 1.** Summary of Immunohistochemical Staining for FOSB

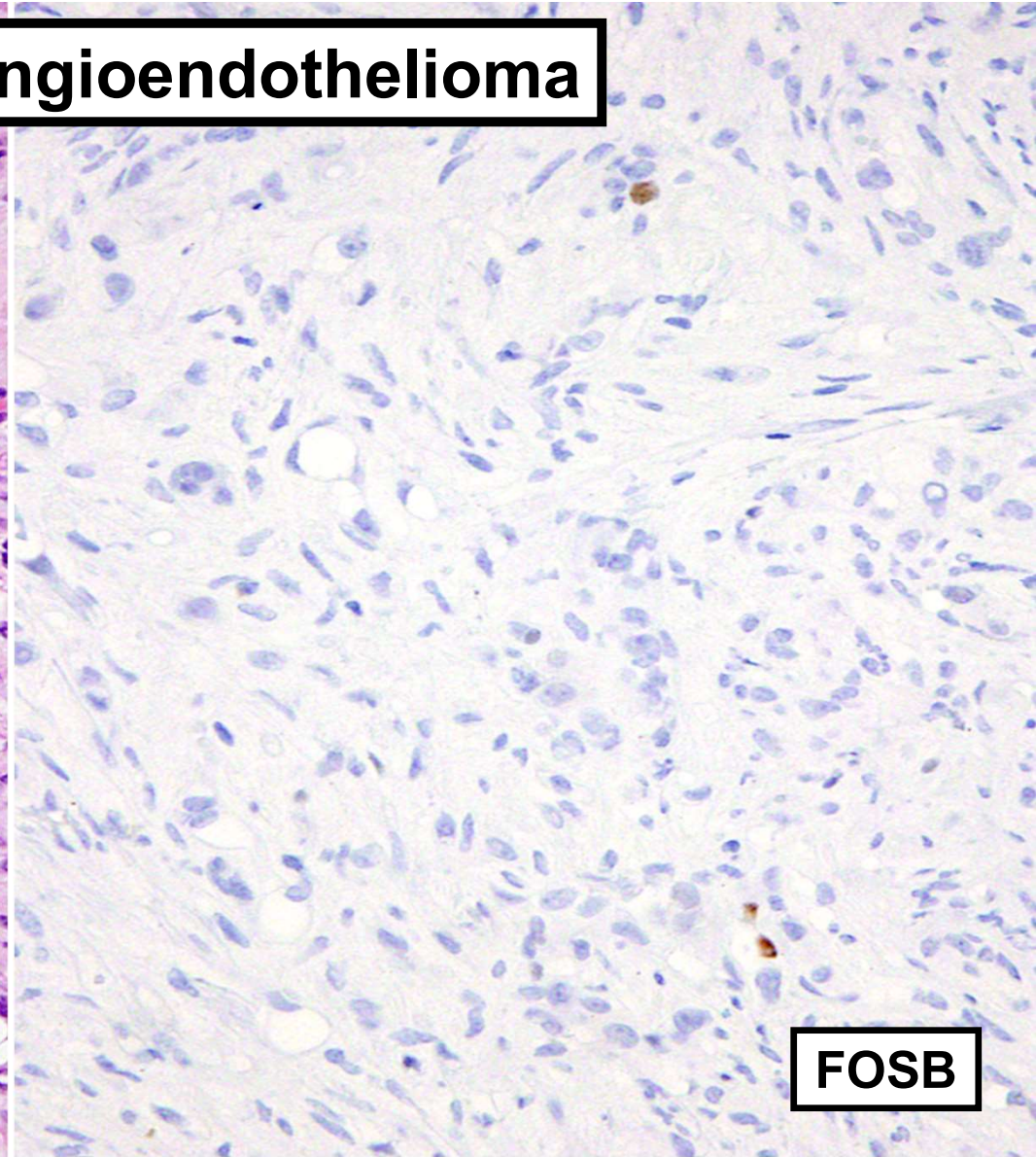
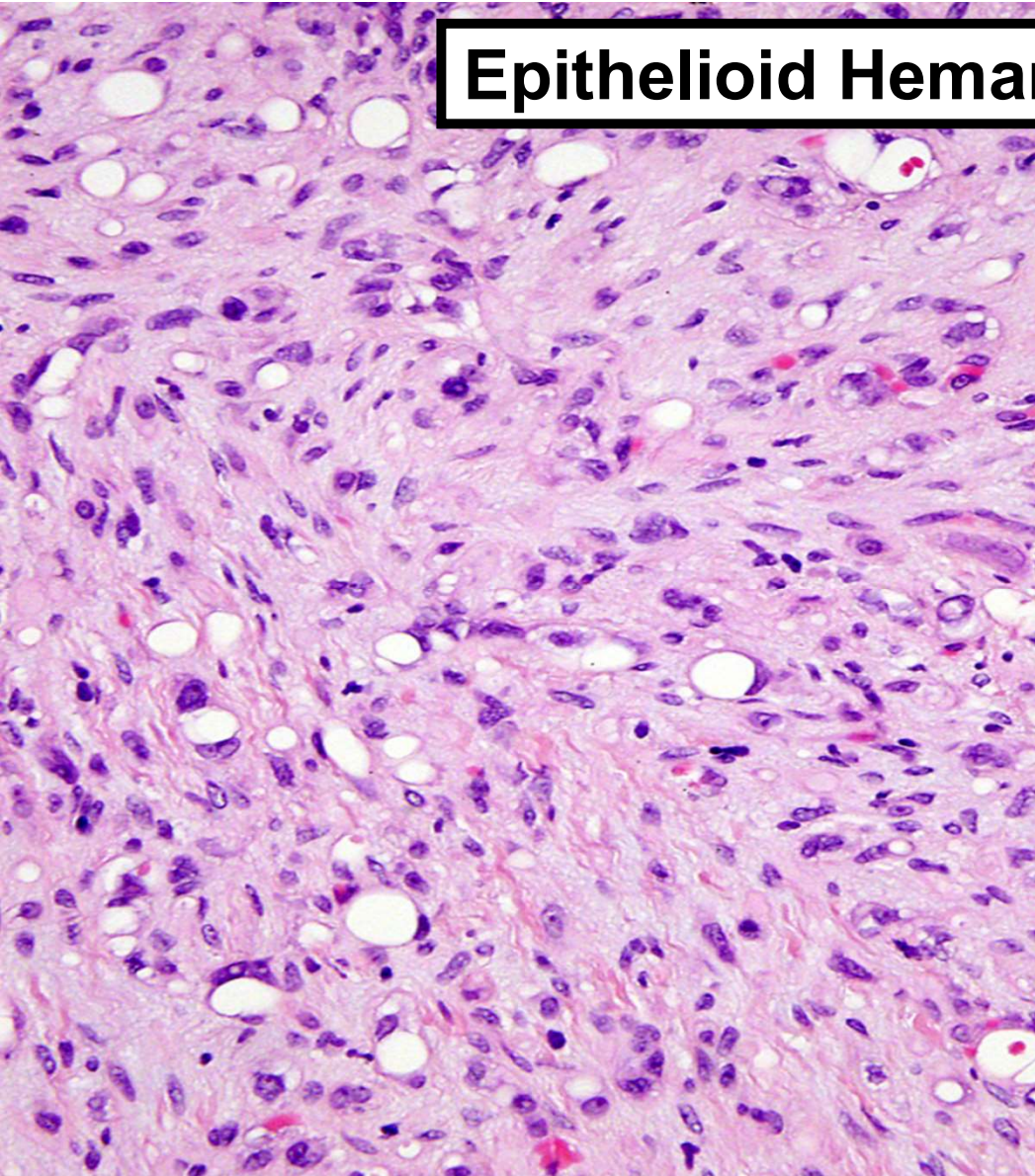
<b>Tumor Type</b>	<b>Total Cases</b>	<b>FOSB Positive (%)*</b>	<b>0</b>	<b>1+</b>	<b>2+</b>	<b>3+</b>	<b>4+</b>
Pseudomyogenic hemangioendothelioma	50	48 (96)	2	0	0	1	47
Epithelioid hemangioma	24	13 (54)	6	4	1	6	7
Conventional	8	6 (75)	0	1	1	4	2
Cellular	10	1 (10)	6	3	0	0	1
Angiolymphoid hyperplasia with eosinophilia	6	6 (100)	0	0	0	2	4
Other endothelial neoplasms and histologic mimics	200	7 (4)	142	42	9	4	3
Epithelioid angiosarcoma	20	1 (5)	11	7	1	0	1
Spindle-cell angiosarcoma	10	1 (10)	9	0	0	1	0
Epithelioid hemangioendothelioma	20	1 (5)	15	4	0	1	0
Epithelioid angiomatous nodule	10	0	9	1	0	0	0

# Epithelioid Hemangioma



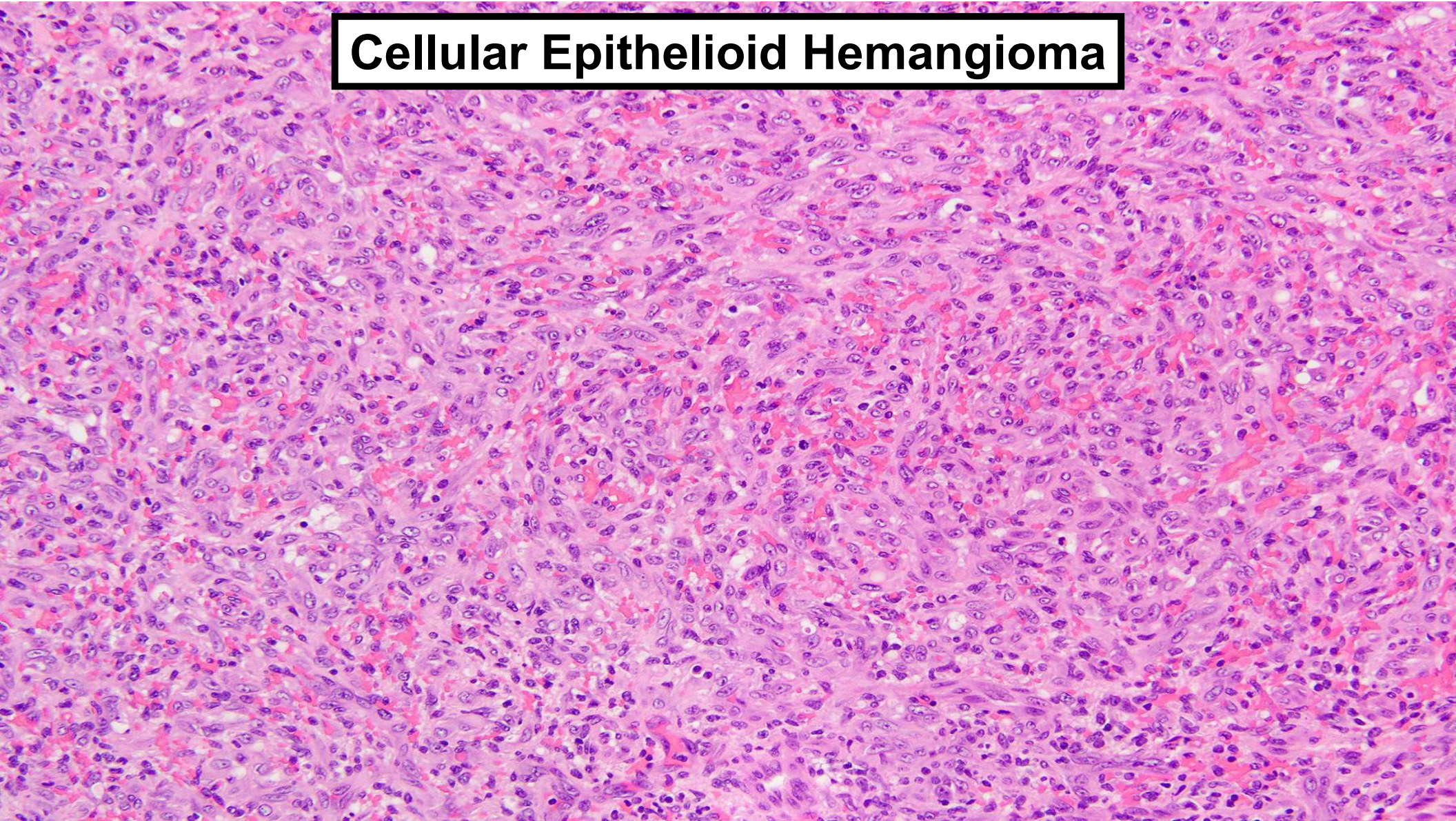
FOSB

# Epithelioid Hemangioendothelioma



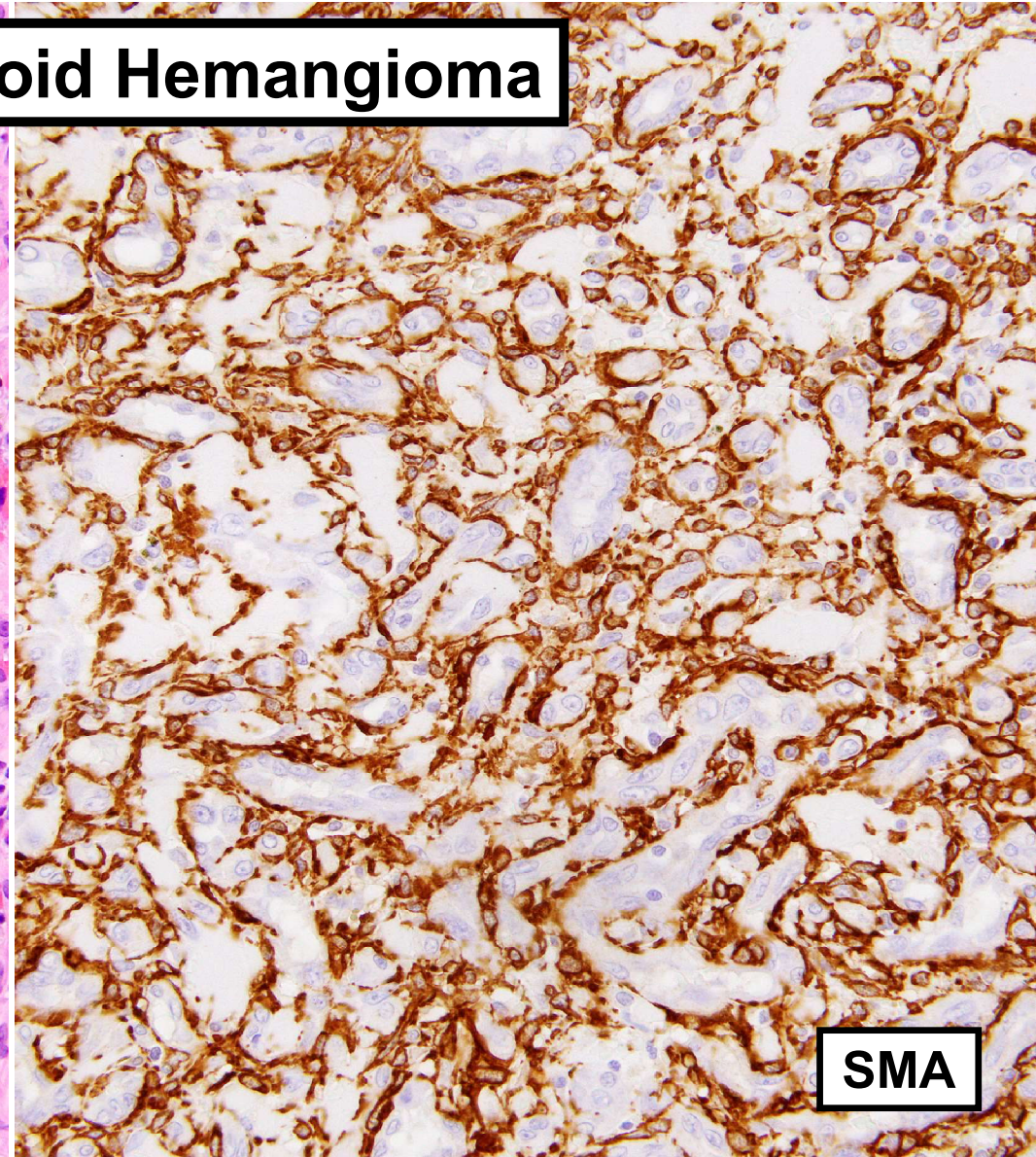
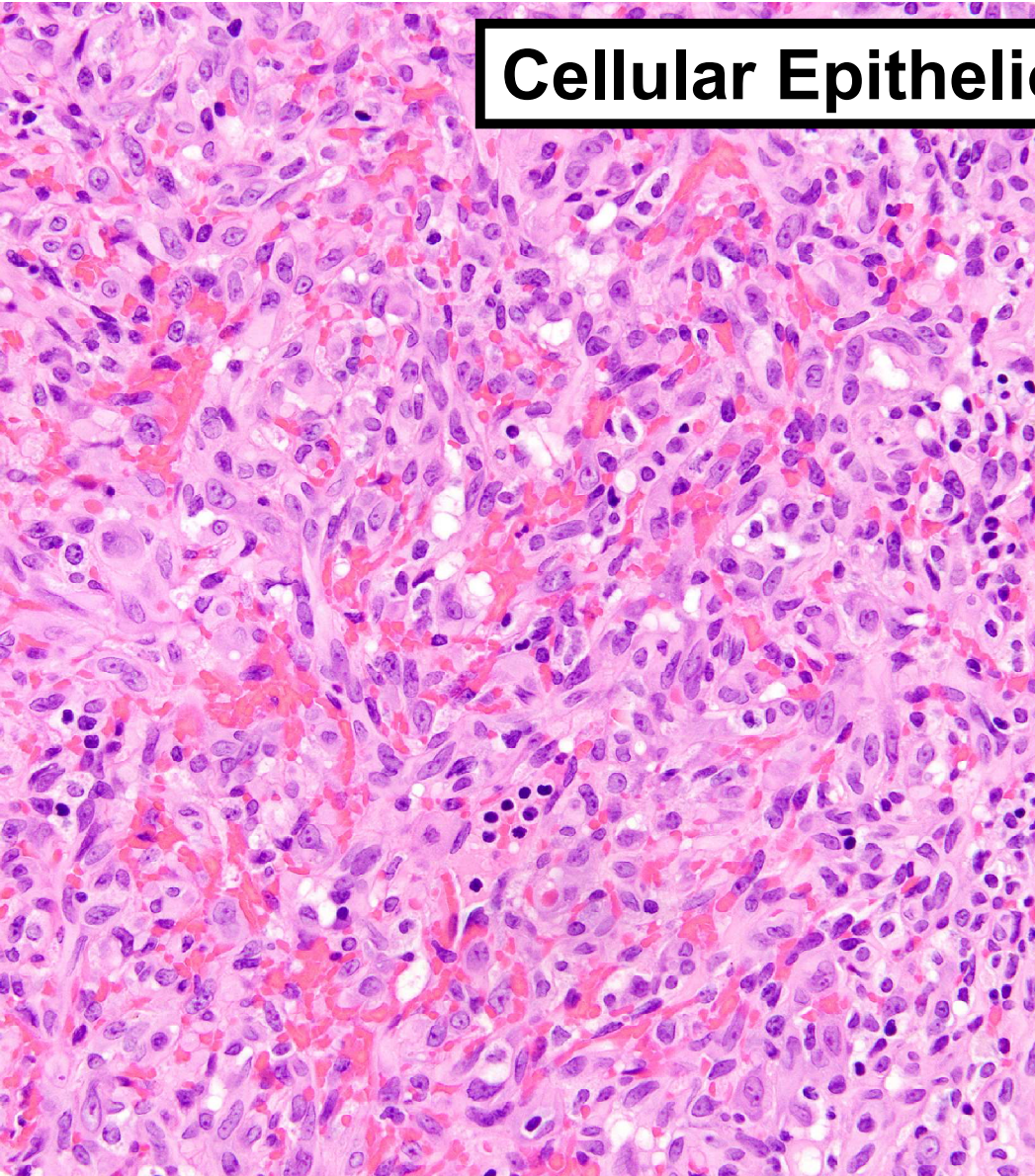
FOSB

# Cellular Epithelioid Hemangioma



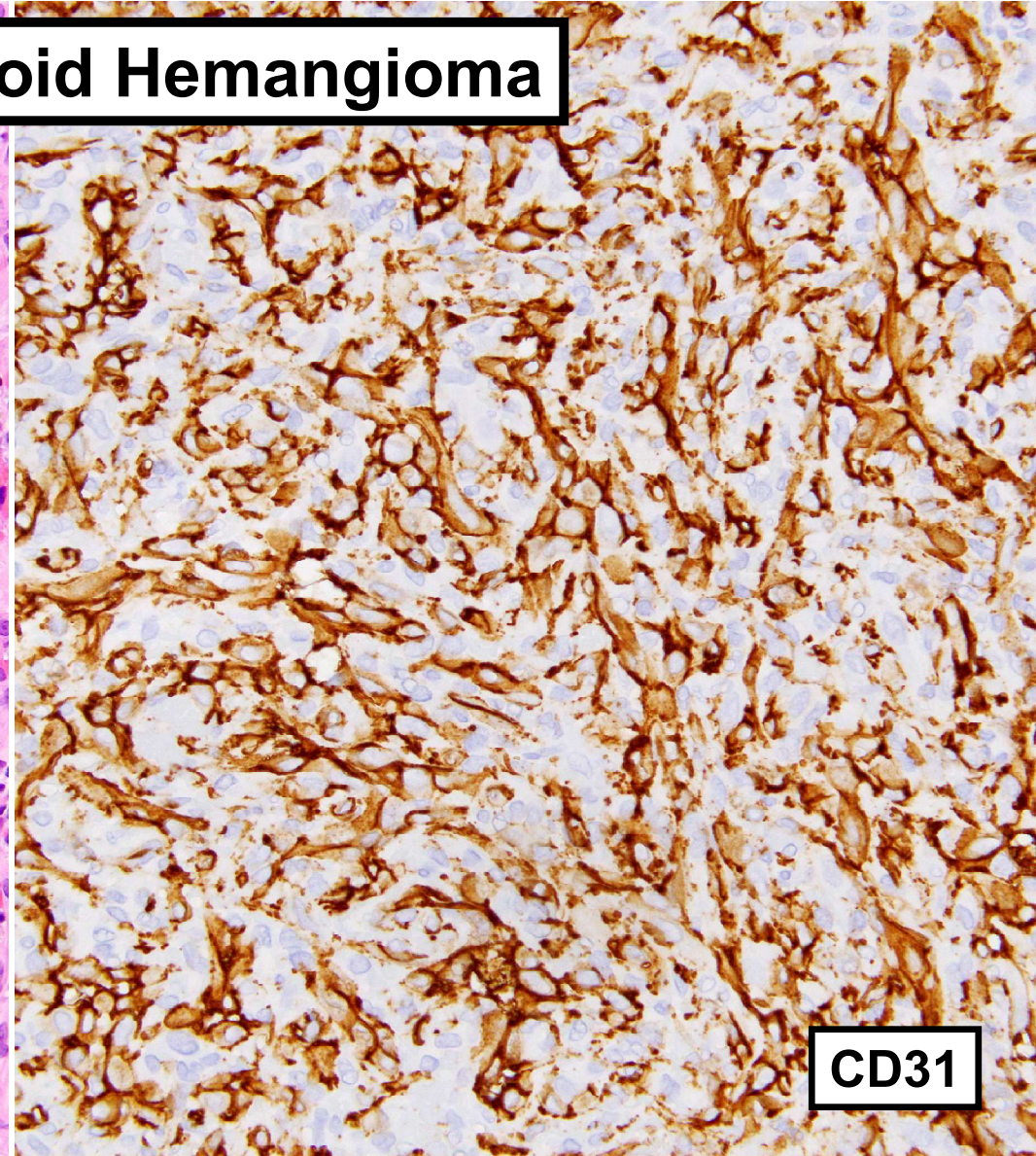
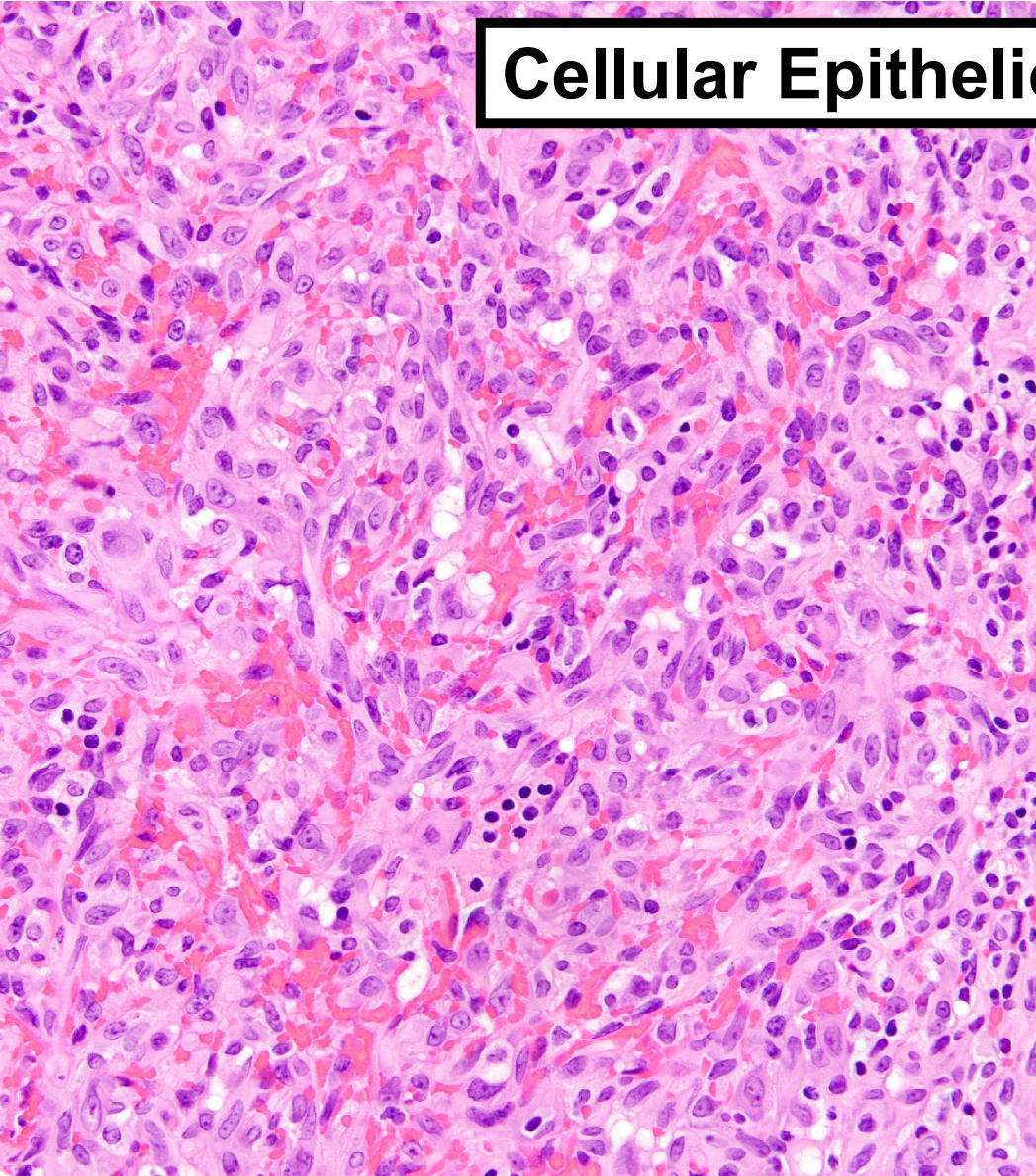


# Cellular Epithelioid Hemangioma



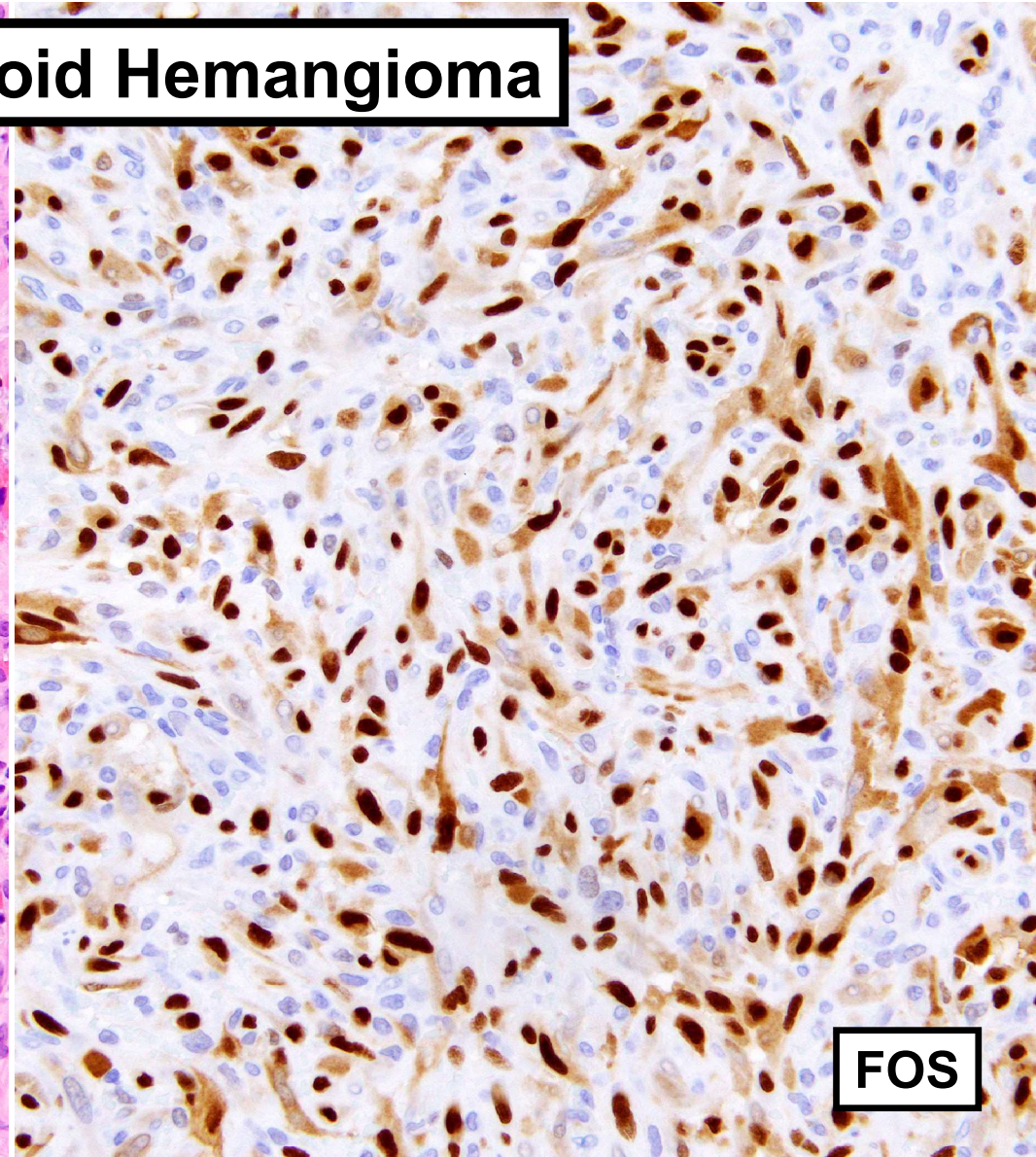
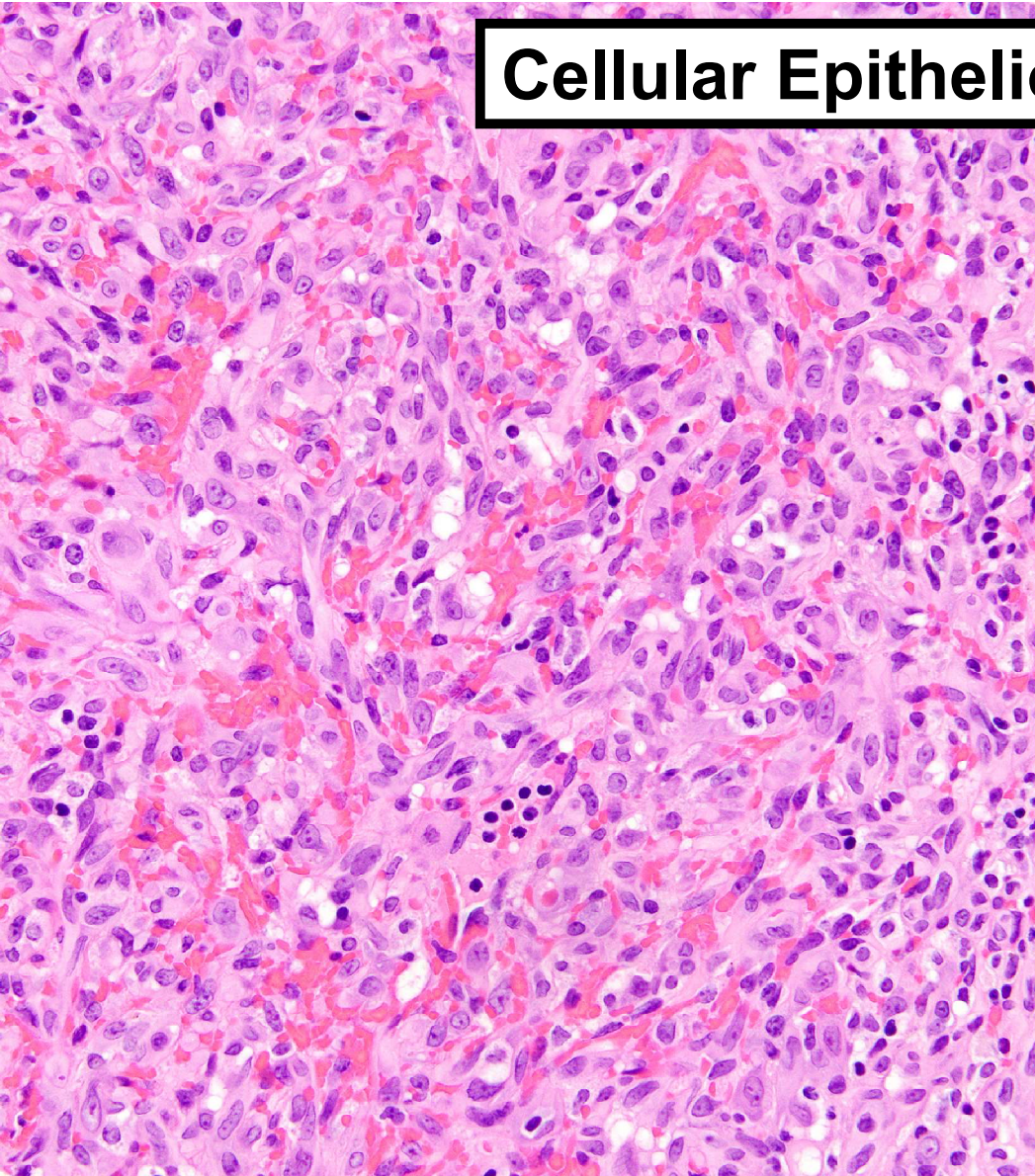
SMA

# Cellular Epithelioid Hemangioma



CD31

# Cellular Epithelioid Hemangioma



FOS

# WHO 2020: NEW GENETICS

## Peripheral nerve sheath tumors

Tumor type	New genetic alterations
Epithelioid schwannoma	<i>SMARCB1</i> mutations
Granular cell tumor	<i>ATP6AP1</i> or <i>ATP6AP2</i> mutations
Benign triton tumor (neuromuscular choristoma)	<i>CTNNB1</i> mutations
Malignant peripheral nerve sheath tumor	<i>SUZ12</i> or <i>EED</i> mutations (loss of H3K27me3)

## Tumors of uncertain differentiation

Tumor type	New genetic alterations
Phosphaturic mesenchymal tumor	<i>FN1::FGFR1</i> , <i>FN1::FGF1</i> (rare)

# Summary

- **“New” soft tissue tumor types continue to be defined, through a combination of clinicopathologic studies and molecular genetic investigations**
- **Large studies of rare soft tissue tumor types enables the development of risk stratification models to aid in prognostication**
- **Discoveries in molecular genetics and the development of correlative markers for IHC make diagnosis of soft tissue tumors more straightforward and reproducible**

 @JLHornick

# THANK YOU!

